Antiviral Activities of *Capsicum annuum*Methanolic Extract against Herpes Simplex Virus 1 and 2





Taghreed A. Hafiz¹, Murad A. Mubaraki^{1*}, Mohamed A. Dkhil^{2,3} and Saleh Al-Quraishy²

¹Clinical Laboratory Sciences Department, College of Applied Medical Sciences, King Saud University, Saudi Arabia

²Department of Zoology, College of Science, King Saud University, Saudi Arabia ³Department of Zoology and Entomology, Faculty of Science, Helwan University, Egypt

ABSTRACT

This study was conducted to quantify the antioxidant and anti-viral activities of *Capsicum annuum* methanolic extracts. The extract of *Capsicum annuum* was prepared by maceration method and the total polyphenolic and flavonoids contents were determined using a spectrophotometric method. The antiradical scavenging property and the maximum nontoxic concentration of the extract was determined using DPPH assay and MTT method, respectively. Serial concentrations of the extract not exceed the maximum nontoxic concentration were used against Vero cells for anti-HSV-1 and ant-HSV-2 effects by plaque assay. The results of the phytochemical analysis showed that the extract has high polyphenolic content while has relative low flavonoids contents. In the antiradical scavenging property test the extract showed at 64.1 μ g/ml 50% DPPH radical scavenging activity and in the MTT assay the extract has relatively low cytotoxicity where the IC₅₀ value of extract against Vero cells was 1078.69 μ g/ml. Furthermore, the extract exhibited a considerable anti-HSV-1 and anti-HSV-2 activities at the concentration of 25 μ g/ml. In conclusion, our findings indicate that the methanolic extract of *Capsicum annuum* could be used in combination with standard drug to treat herpes infection.

Article Information
Received 27 July 2016
Revised 21 August 2016
Accepted 13 October 2016
Available online 02 January 2017

Authors' Contributions

TH, MM, MD and SQ designed and performed the experiments. TH and MM wrote the paper.

Key words

Capsicum annuum, Herpes simplex virus type-1, Herpes simplex virus type-2, Antioxidant, Antiviral.

INTRODUCTION

Tatural products and medicinal herbs have demonstrated to be a principle source of forefront molecules and many plant extracts as crude or an active ingredient with antiviral activity have been documented (Alvarez et al., 2015; Visintini Jaime et al., 2013). Capsicum is a genus belongs to family Solanaceae or nightshades that are an economically important family of flowering plants. Capsicum is highly rich in vitamins A, B, C and E and also contains many minerals such as molybdenum, potassium and manganese. According to Levy et al. (1995) and Szolcsanyi (2004), liquid chromatographymass spectrometry analysis revealed the presence of violaxanthin (C₄₀H₅₆O₄), capsaicinoids (C₁₈H₂₇O₃N), lutein $(C_{40}H_{56}O2)$, β -cryptoxanthin $(C_{40}H_{56}O)$ and β -carotene (C₄₀H₅₆) in *Capsicum* fruits. Furthermore, Liu *et al.* (2013) found three capsaicinoids like capsaicin, dihydrocapsaicin and nordihydrocapsaicin in extracts of Capsicum annuum. In folk medicine, Capsicum is used to relief neuralgia,

rheumatic disorders and non-allergic rhinitis and as irritant in lumbago. Moreover, *Capsicum* has been shown to have an antimicrobial activity inhibiting the growth of gram positive and gram negative bacteria such as *Staphyllococcus* sp., and *Escherichia coli*, respectively (Siehta *et al.*, 1984). In addition, *Capsicum* traditionally been used in Indonesia to treat *Candida albicans* (Soetarno *et al.*, 1997). Moreover, it has been demonstrated by Koffi-Nevry *et al.* (2012) the bactericidal effects of *Capsicum annuum* and *Capsicum frutescens*.

Acute and recurrent infections with herpes simplex virus type-1 (HSV-1) and type-2 (HSV-2) are widely spread worldwide, leading to a wide range of symptoms from weak to severe particularly in immunocompromised patients in whom the infection may become life threatening (Verma et al., 2008). Acyclovir is a nucleoside derivative drug has been widely used to cure HSVs infections (Vijayan et al., 2004). However, due to the high incidence of HSV infections and the widely use of this drug, new strains from the HSVs appear with high resistance to this class of treatment (Visintini Jaime et al., 2013). Thus, it is of highly importance to develop novel antiviral agents acting upon different targets to overcome the problem of viral resistance. Therefore, the current study was aimed to

^{*} Corresponding author: mmubaraki@ksu.edu.sa 0030-9923/2017/0001-0267 \$ 9.00/0 Copyright 2017 Zoological Society of Pakistan

268 T.A. Hafiz et al.

describe the antiviral activity of *Capsicum annuum* against HSV-1 and HSV-2 using Vero cells as an *in vitro* approach.

MATERIALS AND METHODS

Preparation of Capsicum annuum methanolic extract (CaME)

Briefly, the whole *Capsicum annuum* dried fruits were collected from a local market at Riyadh, Saudi Arabia during the months of March, 2016. The fruits were extracted with 70% methanol by shaking for 24 h at room temperature. The methanolic extract was then sieved using Whatman® grade 2 filter paper. After then, supernatant was evaporated under vacuum and dissolved in dimethyl sulfoxide (DMSO).

DPPH radical scavenging assay

Antiradical activity of Capsicum annuum methanolic extract was determined by using 1,1-diphenyl 1-2-picryl hydrazyl (DPPH) radical scavenging assay as described previously by Abdel Moneim (2013). A 2.5% DPPH was freshly made by dissolving 2.5 mg of DPPH in 100 ml of 100% methanol and an aliquot of 100 µl of the extract at the concentrations of 25, 50, 100, 150, 200 and 250 µg/ml were mixed with 3 ml of the DPPH solution, separately in clean tubes. Corresponding blank sample was prepared and ascorbic acid (1-100 µg/ml) was used as reference standard. The mixtures were incubated at room temperature in dark place for 20 min and the change in absorbance was read at 517 nm. The scavenging activity percentage of the sample was calculated by the equation, %DPPH scavenging = [(A control - A sample)/A control]-100, where A is the absorbance. Ascorbic acid was used as positive control and comparing with its' IC50.

Cell culture

Kidney epithelial cells extracted from an African green monkey (84113001/SIGMA, Vero Cell Line) were maintained in Dulbecco's modified Eagle's growth medium (DMEM, Invitrogen, Carlsbad, CA) with 10% FBS (fetal bovine serum, Invitrogen) and supplemented with 0.1% penicillin/0.1% streptomycin obtained from Invitrogen. Cell lines were maintained at 37°C, humidified atmosphere with 5% CO₂ according to the standard procedures and used for cytotoxicity and antivirus assays when approximately 80% confluent.

Cytotoxicity assay

Cell cytotoxicity was carried out using 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay to check whether the methanol extract from *Capsicum annuum* at various concentrations

showed toxicity to Vero cells. Vero cells were plated at an initial density of 1×10^4 cell/ml in a 96-well plate and treated with various concentrations of CaME (50–2500 $\mu g/ml)$ or vehicle for 48 h. 0.5 mg/ml MTT (Sigma) was added to the cells were then incubated with at 37 °C for 4 h. The formed formazan crystals were extracted and dissolved using isopropanol. The absorbance after then was estimated with an ELISA reader at 562 nm. The resultant data were expressed as the percentage of viable cells relative to untreated controls.

Antiviral activity assay

80% confluent cell monolayers seeded in 96-well plates were incubated with increasing non-cytotoxic concentration of CaME, thus, 10, 25, 50, 100, 200 and 500 μ g/ml of CaME were used. Six wells at least were used for every concentration. After 2 h, the cell were infected with HSV-1 or HSV-2 at a multiplicity of infection (MOI) of 0.001 pfu/cell, and re-incubated at 37 °C in a humidified atmosphere of 5% CO₂ in air. The viral cytopathic effect (CPE) was examined daily using a light microscope. When CPE was observed in all virus control wells, the 50% inhibitory concentration (IC₅₀) of viral CPE was estimated in comparison to the virus control, as described previously. Acyclovir (Sigma®) at concentration of 0.5, 1, 2, 4 and 5 μ g/ml served as the positive control.

Viral plaque number reduction assay

Inhibition of HSV-1 and 2 replications were estimated with plaque reduction assay. Vero cells were distributed in 24-well plates at a density of 10×10^4 cells. The plaque reduction assay was done in infecting cell monolayers with HSV-1 or HSV-2 at a MOI of 0.001 pfu/cell in presence of serial dilutions of the non-cytotoxic concentration of CaME (10, 25, 50, 100, 200 and 500 µg/ml) for 2 h at 37°C in a humidified atmosphere of 5% CO₂ in air. The medium was subsequently discarded from the wells, and the cells were washed with clean medium twice and covered again with a fresh medium containing 1.2% methylcellulose (Sigma) with CaME at the same concentration. After incubation for 24 h at 37°C in humidified atmosphere with 5% CO₂, the supernatant was discarded, and the cells were fixed in 10% formalin in phosphate-buffered saline and then stained with 0.8% crystal violet in 50% ethanol and the viral plaques were counted microscopically at low power.

Statistical analysis

One-way ANOVA and statistical comparisons between the groups were performed with Duncan's test, using a statistical package program (SPSS version 17.0). Results represent the mean of three independent experiments ± standard deviations.

Table I.- Total polyphenolic content, total flavonoid content and antiradical activity of *Capsicum annuum* methanolic extract.

Extract	Capsicum annuum methanolic extract
Total polyphenolic ^a	76.3±4.36
Total flavonoid ^b	12.1±0.72
DPPH IC ₅₀	64.1±3.2

^a, Total phenolic content in *Capsicum annuum* methanolic extract is determined as mg/g gallic acid equivalent of phenols/g dried extract; ^b, Total flavonoid content in *Capsicum annuum* methanolic extract is determined as mg/g quercetin equivalents of flavonoids/g dried extract. Radical scavenging activity was measured changes in absorbance at 517 nm. Data are expressed as mean ± standard deviations of three independent experiments each performed in duplicate.

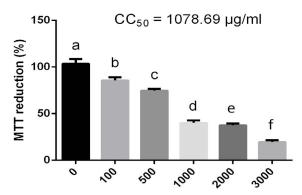
RESULTS AND DISCUSSION

As shown in Table I, the total polyphenolic content of Capsicum annuum methanolic extract was 76.3±4.36 mg/g gallic acid equivalent of phenols/g dried extract while, the total flavonoid content was 12.1±0.72 mg/g quercetin equivalents of flavonoids/g dried extract. Capsicum annuum is known as a good source of phytochemicals compounds such as polyphenols, carotenoids and ascorbic acid, and is recognized as beneficial to health. These phytochemicals display high antioxidant property, and their consumption has been linked to a decreased risk of developing chronic and degenerative diseases such as cancer and diabetes (Bode and Dong, 2011; Kwon et al., 2007). Among phytochemicals, polyphenols are particularly interesting for their free radical scavenging activities and in vivo biological activities. The major active compounds found in red pepper are capsaicinoids such as capsaicin, dihydrocapsaicin, and nordihydrocapsaicin. In accordance with our findings, Nascimento et al. (2014) found that the phenolic content ranged from 3.2 ± 0.22 to 110.6 ± 1.03 mg GAE g-1 of whole fruits of C. frutescens extract. On other hand, Yazdizadeh Shotorbani et al. (2013) found that the red pepper contains polyphenols ranged from 63 to 78 mg GAE g-1 of whole fruits of C. annuum extract. However, our finding in flavonoids is greater than that of recorded by Yazdizadeh Shotorbani et al. (2013) who reported the maximum flavonoids found in red pepper is 7.8 mg/g quercetin equivalents of flavonoids/g dried extract.

Furthermore, we determined the antioxidant activity of CaME by using DPPH radical scavenging activity assay. The highest antiradical activity based on the DPPH model system was found for CaME, which was comparable to the activity of vitamin C. The obtained data for the DPPH radical scavenging assay was shown in Table I. The

data indicated that the 50% DPPH radical scavenging activity of CaME was 64.1 µg/ml while, the 50% DPPH radical scavenging activity of vitamin C was 68.5 µg/ ml. The whole fruit of Capsicum annuum methanolic extract presented higher phenolics and flavonoids contents, and these contents could be correlated with the DPPH radical scavenging potential. The present finding was in agreement with Chavez-Mendoza et al. (2015), who reported that bell peppers contain high concentration of antioxidant activity. In this regard, Materska and Perucka (2005) attributed this activity to capsaicin and dihydrocapsaicin found in the fruit. The antioxidant properties of phenolic compounds originate from their properties of proton loss, chelate formation, and dismutation of radicals. Phenols are compounds that have the ability to destroy radicals because they contain hydroxyl groups.

In addition, the cytotoxicity assay was performed using MTT to assess the metabolically active cells which convert the dye to a water-insoluble dark blue formazan by reduction process of splitting tetrazolium ring (Alvarez et al., 2015). The cytotoxicity of CaME against Vero cells is summarized and shown in Figure 1. The CC₅₀ value of CaME against Vero cells was 1078.69 µg/ml. According to our findings, CaME has low cytotoxicity and Vero cells can tolerate it well. The low toxicity of CaME was reported by Viktorija et al. (2013) who noted that capsicum extract did not show any cytotoxic effect at the examined concentrations while, capsaicin alone (100 µM) showed high cytotoxicity and they assumed that the other compounds (carotenoids, vitamins, and other polyphenolic substances) within the extract interact antagonistic with the cytotoxic effect of capsaicin. However, Park et al. (2012)



Capsicum annuum methanolic extract Conc. µg/ml

Fig. 1. The cytotoxic effect of *Capsicum annuum* methanolic extract. CC_{50} is the concentration of the 50% cytotoxic effect. Data are showed as Mean of three independent experiments each performed in duplicate. Means indicated with different letters differ significantly.

270 T.A. Hafiz et al.

Table II.- The antiviral activity of Capsicum annuum methanolic extract.

CaME	Cell Cytopathic Effect (CPE)		
	HSV-1	HSV-2	
10 μg/ml	-	-	
25 μg/ml	-/+	-	
$50 \mu g/ml$	+	-/+	
$100 \ \mu g/ml$	+	+	
$200~\mu g/ml$	+	+	
500 μg/ml	+	+	

^{+,} Virus inhibition; +/-, Partial inhibition; -, No inhibition, presence.

found that the proliferation of Vero cells was significantly reduced by CaME and they attributed the cytotoxicity of red pepper to its antioxidant activity.

Results of the antiviral activity and the cytopathic inhibitory assay of CaME against HSV-1 and HSV-2 were presented in Table II. Concentrations from 50-500 µg/ml completely inhibit *in vitro* growth of HSV-1 with a partial inhibition at 25 µg/ml, while concentrations from 100-500 µg/ml completely inhibit *in vitro* growth of HSV-2 with a partial inhibition at 50 µg/ml. The present findings were in accordance with Ozcelik *et al.* (2011) and Carole *et al.* (2015). However, Bourne *et al.* (1999) stated that capsaicin does not have a direct anti-herpes effect, whereas civamide (cis-capsaicin) exhibited remarkable inhibition against genital HSV.

As illustrated in Figure 2, CaME at 40 µg/ml showed 50% inhibition of plaque of HSV-1 and provided 100% inhibition against HSV-1 at 200-500 µg/ml as indicated by the plaque inhibition assay. Likewise, the anti-HSV-2 potential of the CaME was performed and the results were illustrated in Figure 3. The HSV-2 was more tolerate to CaME and according to the obtained results, CaME at 40 ug/ml provided 40% inhibition of plaque of HSV-2 and provided 100% inhibition against HSV-2 at 200 - 500 ug/ml. The present findings might be indicating the antiherpes simplex virus types-1 and 2 of Capsicum annuum and these activities due to the ability of phytochemical compounds in the extract to interfere at the attachment points and penetration of the virus in cells. The principal act of the extract is probably modified the cellular and viral receptors, preventing the virus from attaching to cells. According to Obi et al. (2006), if the receiving cells are modified before the viral infection, the virus's ability to bind and penetrate the cells would be greatly reduced. Indeed, the study of Kouassi and Koffi-Nevry (2012) showed that Capsicum annuum contains biological active

compounds and vitamins such as ascorbic acid and the β-carotene are responsible for these activities.

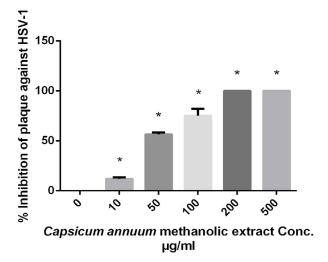
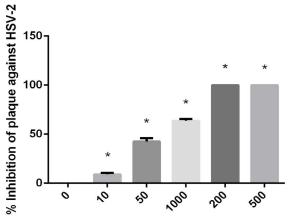


Fig. 2. Inhibition activity of *Capsicum annuum* methanolic extract against HSV-1. IC₅₀ is the concentration of the extract required to inhibit 50% virus-induced cytopathic effect. Results are represented as Mean of two independent experiments each performed in triplicate. Means indicated with (*) differ significantly.



Capsicum annuum methanolic extract Conc. µg/ml

Fig. 3. Inhibition activity of *Capsicum annuum* methanolic extract against HSV-2. IC₅₀ is the concentration of the extract required to inhibit 50% virus-induced cytopathic effect. Results are represented as Mean of two independent experiments each performed in triplicate. Means indicated with (*) differ significantly.

CONCLUSION

In conclusion, the crude extract of Capsicum annuum

was active against HSV-1 and HSV-2 and demonstrated antiviral activity with low cytotoxicity which may be used in combination with standard drug to treat herpes infection.

ACKNOWLEDGMENT

The authors would like to appreciate the Deanship of Scientific Research at King Saud University for funding this study through the research group project No. RG-198.

Conflict of interest statement

We declare that we have no conflict of interest.

REFERENCES

- Abdel Moneim, A.E., 2013. The neuroprotective effects of Purslane (*Portulaca oleracea*) on rotenone-induced biochemical changes and apoptosis in brain of rat. *CNS Neurol. Disord. Drug Targets*, **12**: 830-841. http://dx.doi.org/10.2174/187152731131 29990081
- Alvarez, A.L., Habtemariam, S., Abdel Moneim, A.E., Melon, S., Dalton, K.P. and Parra, F., 2015. A spiroketal-enol ether derivative from *Tanacetum vulgare* selectively inhibits HSV-1 and HSV-2 glycoprotein accumulation in Vero cells. *Antiviral Res.*, 119: 8-18. http://dx.doi.org/10.1016/j.antiviral.2015.04.004
- Bode, A.M. and Dong, Z., 2011. The two faces of Capsaicin. *Cancer Res.*, **71**: 2809-2814. http://dx.doi.org/10.1158/0008-5472.CAN-10-3756
- Bourne, N., Bernstein, D.I. and Stanberry, L.R., 1999. Civamide (cis-capsaicin) for treatment of primary or recurrent experimental genital herpes. *Antimicrob. Agents Chemother.*, **43**: 2685-2688.
- Carole, K.A., Valery, A.E., Kouadio, Y., Clement, K.K., Guillaume, Y.Y., Rose, K. and Guillaume, L.Y., 2015. Cytotoxic and antiviral activity of methanolic, ethanolic and acetate extracts of six varieties of Capsicum. *Int. J. Curr. Microbiol. appl. Sci.*, **4**:76-88.
- Chavez-Mendoza, C., Sanchez, E., Munoz-Marquez, E., Sida-Arreola, J.P. and Flores-Cordova, M.A., 2015. Bioactive compounds and antioxidant activity in different grafted varieties of bell pepper. *Antioxidants*, 4: 427-446. http://dx.doi.org/10.3390/antiox4020427
- Koffi-Nevry, R., Kouassi, K.C., Nanga, Z.Y., Koussémon, M. and Loukou, G.Y., 2012. Antibacterial activity of two bell pepper extracts: *Capsicum annuum* L. and *Capsicum frutescens. Int. J. Fd. Proper.*, **15**: 961-971. http://dx.doi.org/10.1080/10942912.2010.509896

- Kouassi, C.K. and Koffi-Nevry, R., 2012. Evaluation of knowledge and use of pepper varieties (Capsicum) grown in the Côte d Ivoire. *Int. J. Biol. Chem. Sci.*, **6**: 175-785.
- Kwon, Y.I., Apostolidis, E. and Shetty, K., 2007. Evaluation of pepper (capsicum annuum) for management of diabetes and hypertension. *J. Fd. Biochem.*, **31**: 370-385. http://dx.doi.org/10.1111/j.1745-4514.2007.00120.x
- Levy, A., Harel, S., Palevitch, D., Akiri, B., Menagem, E. and Kanner, J., 1995. Carotenoid pigments and beta-carotene in paprika fruits (*Capsicum* spp.) with different genotypes. *J. agric. Fd. Chem.*, **43**: 362-366. http://dx.doi.org/10.1021/jf00050a019
- Liu, A., Han, C., Zhou, X., Zhu, Z., Huang, F. and Shen, Y., 2013. Determination of three capsaicinoids in Capsicum annuum by pressurized liquid extraction combined with LC-MS/MS. *J. Sep. Sci.*, **36**: 857-862. http://dx.doi.org/10.1002/jssc.201200942
- Materska, M. and Perucka, I., 2005. Antioxidant activity of the main phenolic compounds isolated from hot pepper fruit (*Capsicum annuum* L). *J. agric. Fd. Chem.*, **53**: 1750-1756. http://dx.doi.org/10.1021/jf035331k
- Nascimento, P.L., Nascimento, T.C., Ramos, N.S., Silva, G.R., Gomes, J.E., Falcao, R.E., Moreira, K.A., Porto, A.L. and Silva, T.M., 2014. Quantification, antioxidant and antimicrobial activity of phenolics isolated from different extracts of *Capsicum frutescens* (Pimenta Malagueta). *Molecules*, 19: 5434-5447. http://dx.doi.org/10.3390/molecules19045434
- Obi, R.K., Iroagba, I.I. and Ojiako, O.A., 2006. Virucidal potential of some edible Nigerian vegetables. *Afr. J. Biotechnol.*, **5**: 1785-1788.
- Ozcelik, B., Kartal, M. and Orhan, I., 2011. Cytotoxicity, antiviral and antimicrobial activities of alkaloids, flavonoids and phenolic acids. *Pharm. Biol.*, **49**: 396-402. http://dx.doi.org/10.3109/13880209.201 0.519390
- Park, J.H., Jeon, G.I., Kim, J.M. and Park, E., 2012. Antioxidant activity and antiproliferative action of methanol extracts of 4 different colored bell peppers (*Capsicum annuum* L.). *Fd. Sci. Biotechnol.*, 21: 543-550. http://dx.doi.org/10.1007/s10068-012-0069-2
- Siehta, A.S., Magdoub, M.W.I., Fayed, E.O. and Hoft, A.A., 1984. Effect of added salt and *Capsicum tincture* on non lactic acid bacteria in pickled Domiati Cheese. *Annls. Agric. Sci.*, **29**: 793-810.
- Soetarno, S., Sukrasno, S., Yulinah, E. and Sylvia,

T.A. Hafiz et al.

S., 1997. Antimicrobial activities of the ethanol extracts of *Capsicum* fruits with different levels of pungency. *J. math. Sci.*, **2**: 57-63.

- Szolcsanyi, J., 2004. Forty years in capsaicin research for sensory pharmacology and physiology. *Neuropeptides*, **38**: 377-384. http://dx.doi.org/10.1016/j.npep.2004.07.005
- Verma, H., Patil, P.R., Kolhapure, R.M. and Gopalkrishna, V., 2008. Antiviral activity of the Indian medicinal plant extract Swertia chirata against herpes simplex viruses: a study by *in vitro* and molecular approach. *Indian J. med. Microbiol.*, 26: 322-326. http://dx.doi.org/10.4103/0255-0857.43561
- Vijayan, P., Raghu, C., Ashok, G., Dhanaraj, S.A. and Suresh, B., 2004. Antiviral activity of medicinal

- plants of Nilgiris. *Indian J. med. Res.*, **120**: 24-29.
- Viktorija, M., Liljana, K.G. and Karen, N., 2013. Cytotoxic effects of capsaicin and capsicum extracts on neuroblastoma cells. International conference on natural products utilization: From plants to pharmacy shelf, Bansko, Bulgaria.
- Visintini Jaime, M.F., Redko, F., Muschietti, L.V., Campos, R.H., Martino, V.S. and Cavallaro, L.V., 2013. *In vitro* antiviral activity of plant extracts from Asteraceae medicinal plants. *Virol. J.*, **10**: 245. http://dx.doi.org/10.1186/1743-422X-10-245
- Yazdizadeh Shotorbani, N., Jamei, R. and Heidari, R., 2013. Antioxidant activities of two sweet pepper *Capsicum annuum* L. varieties phenolic extracts and the effects of thermal treatment. *Avicenna J. Phytomed.*, **3**: 25-34.