ALLELOPATHIC INFLUENCE OF POPPY (*Papaver somniferum* L.) ON EMERGENCE AND INITIAL SEEDLING GROWTH OF RED RICE (*Oryza punctata* L.)

Muhammad Ather Nadeem¹, Bilal Ahmad Khan¹*, Sadia Afzal², Hasnain Abbas³, Muhammad Khuram Dar³, Muhammad Ehsan Safdar¹, Ishtiaq Hassan⁴, Muhammad Asif¹, Muhammad Adnan¹, Amir Aziz⁵

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ABSTARCT

Allelopathy plays crucial role in effective weed control. Opium (Papaver somniferum L.) crop release different allelochemicals at maturity which have potential to act as natural weed killer in different crops. Phytotoxic effect of poppy (P. somniferum) was examined on emergence and initial seedling growth of red rice (Oryza punctate L.). Aqueous extract of different plant parts (leaves, stem and flower) was used at various concentrations (0.25%, 0.5%, 1%, 2%, 4% and 8%) along with distilled water as control. The aqueous extracts of leaves, stem and flower of *P. somniferum* significantly inhibited the emergence, seedling growth as well as root length (cm), shoot length (cm), fresh weight (g) and dry weight (g) of O. punctata. Maximum mean emergence time (9.18 days) and minimum shoot length (1.13 cm) emergence index (0.89) and emergence percentage (6.67%) was observed under fruit extract at 8% concentration. P. somniferum aqueous extract of stem at 8% concentration took maximum time to complete 50% emergence and gave minimum root length, fresh weight, and dry weight of *O. punctata*. Based on these finding it can be concluded that the phyto-chemicals present in *P. somniferum* can be used as eco-friendly *O. punctata* growth inhibitor to manage this weed in crops especially under organic cropping.

Keywords: Emergence, inhibitory, phytotoxic, plant parts, promotor, seedling growth, weed growth.

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¹ Department of Agronomy, College of Agriculture, University of Sargodha, Sargodha, Pakistan.

^{*} Corresponding Author, Email: bilalahmadkhan678@gmail.com

² Department of Botany, University of Agriculture, Faisalabad, Pakistan.

³ Department of Agronomy, University of Agriculture, Faisalabad, Pakistan.

⁴ Inservice Agriculture Training Institute, Sargodha, Pakistan.

⁵ Department of Soil & Environmental Sciences, College of Agriculture, University of Sargodha, Pakistan.

INTRODUCTION

Crop plants suffers stress caused by unwanted weeds through competition for water, light nutrients and space. Stress created by weed plants results in an increasing mortality of whole crop plant, reduced growth, decreases in output and quality (Gallandt and Weiner, 2001). Reduction in wheat yield from 20 - 60% due to weed infestations (Turk and Tawaha, 2002). Loses in different crops due to weeds infestation in Pakistan are 20 to 30%. In cereal crops, the losses due to weed infestations are approximately 30, 40, 4 and 5 billion rupees for wheat, rice, gram and maize, respectively (Anonymous, 2005).Therefore, it is necessary to control the weeds before critical competition period to enhance the yield of crop (Macias et al., 2002).Different methods are used to control weeds such as biological, chemical, mechanical, and cultural weed control (Melander et al., 2005). In modern agriculture production system weeds efficiently controlled by use of chemical herbicides (Bajwa, 2014). The use of chemical for effective weed control result in low quality of product, weed resistance, environmental pollution (Macias et al., 2002). Extreme and continuous application of same type of herbicides has resulted in resilient weed populations and this phenomenon urged upon the misuse of allelopathic possible of crop plants (Ferreira and Reinhardt, 2010). Allelochemicals could be used as tool to control weeds and resistance (Jabranet al., 2015). Allelopathic plants have ability to inhibit the initial weeds growth (Chung et al., 2006).

(P. somniferum) Opium а common medicinal plant, belonging to family Papaveraceae. (Labancaet et al., 2018). Many researchers reported that opium crop release different allelochemicals at maturity which inhibit growth of various plants including crops and weeds. According to Alam and Islam (2002) water extract of some weeds might have promotry or inhibitory effect on germination and initial seedling growth of target weed. Different plants have different potential on target weed or crop plant.Water extract of wild onion leaves produced inhibitory impact on wheat crop germination and initial et al., growth of seedling (Baber 2009).Opium is most important dominant and competitive broad-leaves weed in winter cereals reported to have allelopathic effect, the water extracts from fresh plant parts of opium significantly inhibit germination of both barley and wheat crop (Ravlićet al., 2012). Ghodake et al., (2012) reported that allelochemicals inhibited the seed germination by blocking hydrolysis of nutrients reserve. Crop plants for example opium (*P*. somniferum), sunflower (Helianthus annuus), eucalyptus (Eucalyptus camaldulensis), turnip rape (Brassica compestris), and other species had allelopathic effects against weed growth (Skrzypek et al., 2015). It is needed to examine the phytotoxic effects of *P.somniferum* aqueous extracts on Oryza punctata germination and initial seedling growth. To control weeds many alternative strategies like as non-chemical control usina aqueous extracts (biobv herbicides) of weed plants (Cirujedaet al., 2008). Bio-herbicides are ecofriendly for weed management and easily biodegradable than synthesized herbicides (Ghafarbi et al., 2012). Water extract of opium is natural and has no chemical hazards on target crop and helpful in inhibiting weeds and enhancing the productivity of crop. This experiment was conducted to explore allelopathic potential of aqueous extracts of parts such as leaves, stem and flower of opium plant to control the Oryza punctata (red rice) weed germination and initial seedling growth specifically.

MATERIALS AND METHODS

Experiment was conducted in 2017 in weed science laboratory at Department of Agronomy, University of Agriculture Faisalabad, Pakistan. The objective of this experiment was to evaluate the allelopathic potential of aqueous extracts of *P. somniferum*on summer weed *O. punctata*. The study was arranged in completely randomized design (CRD) with factorial arrangement having three replications.

Collection of *P. somniferum* plant parts

To prepare aqueous extract of *P. somniferum* plant parts. Plants of *P.*

*somniferum*were collected from Farm, Agronomic University of agriculture Faisalabad at maturity and dried for two weeks at ambient temperature. After sun dry different parts of plants were separated and chopped into about 2 cm pieces for extract formation.

Preparation of aqueous extract

Aqueous extracts of various plants parts of P. somniferum were prepared by adding 10g of chopped dried plant material into 100ml of distilled water in bottles separately at ratio of 1:10 w/v and left for at least 24 hours at room temperature. These aqueous extracts were made from each desired part of opium such as leaves, stem, fruit and flower etc. Then the material was passed through a muslin cloth to attain the water extracts of different parts of *P. somniferum*. That process gave the 10% extract, from this 67.2ml extract were added in 16.8ml distilled water to make final volume 84ml that was considered as stock solution. From this stock solution we made further dilutions that are (0.25%, 0.5%, 1%, 2%, 4%, 8%).

These dilutions were made by using equation

$$C_1 V_1 = C_2 V_2$$

Each dilution has 42ml total volume. Each dilution of each extract placed in separate bottles and then tagged these bottles by name of each dilution with its plant name too carefully for their easy utilization in next procedure.

Laboratory experiment

To check the allelopathic effect treatments combinations of P. 21 somniferumwere applied on O. punctata. Five seeds of red rice were placed in every petri plates having filter paper double layer. Then 7ml of opium extracts dilutions of each part i.e. leaves, stem, flowers and fruits were added in recommended petri plates having 3 replications foreach dilution. One treatment was kept as control and moist with distilled water. Then to reduce evaporation petri plates were covered and rapped with scotch tape. The petri plates were kept at temperature of 30°C and each treatment

were again moistened after one week. The data regarding emergence of seeds were recorded every day for the period of 14 days. After that period removed the emerged seedlings from the petri plates and observed the different parameters like shoot length, root length, fresh and dry weight. Fresh weight was recorded immediately after harvesting while the dry weight of seedlings was observed after drying in oven for two days at 60 °C. To check allelopathic potential used different concentrations (0%, 0.25%, 0.5%, 1%, 2%, 4% and 8%) and three different plant parts (leaves, stem and flower) of *P. somniferum* on *O. punctata*.

DATA RECORDED Emergence percentage of *O. punctata* (%)

Number of emerged seeds were counted daily up to 14 days after which the emergence ceased. The emergence % was calculated by using following formula.

 $Emergence (\%) = \frac{Number of emerged seeds}{Total number of seeds} \times 100$

Emergence index of O. punctata

The emergence index was observed as per association of official seed analysis (1983) by using the following formula

$$GI = \frac{No. of \ emerged \ seeds}{Days \ of \ first \ count} + - - -$$
$$- + \frac{No. of \ emerged \ seeds}{Days \ of \ final \ count}$$

Time to 50% emergence of *O. punctata*

The time to attain 50% germination or emergence (T_{50} or E_{50}) was calculated according to the formula of Coolbear*et al.* (1984)

$$T\mathbf{50} = ti + \begin{bmatrix} \frac{N}{2} - ni \\ nj - ni \end{bmatrix} (tj - ti)$$

Mean emergence time of O. punctata

Mean emergence time (MET) was examined by the equation of Ellis and Reborts (1981).

$$\mathsf{MET} = \sum (Dn) / \sum n$$

Growth attributes of *O. punctata*

All seedlings from each petri plate separated 14 days after were germination. After that root and shoot length was measured by using meter rod from base level to top of the plants. Fresh seedlings weight was examined by separating seedlings from petri dish and measuring by using digital weight balance. Dry weight of seedlings was recorded by oven drying the seedlings at 60 °C for 48 hours then weighted to get average dry weight of seedling by using digital balance.

Statistical Analyses

Data analyses were carried out by using the Statistics software (version, 8.1Statistix, Tallahassee, FL, USA). The least significant difference (LSD) test was employed to compare the means at 5% Probability level.

RESULTS AND DISCUSSION 1. Emergence percentage of *O. punctata* (%)

Emergence percentage of red rice significantly influenced by the application of P. somniferum aqueous extracts at different concentrations. Data were presented in Table-1. Among different plant parts of P. somniferum, highest emergence was recorded from aqueous extracts of leaves (88.57%) while statistically at par with stem extract of *P*. somniferum. Lowest emergence percentage (63.81%) was recorded from fruit extract. Leaves showed the hermetic influence on emergence percentage of O. punctata. different concentration Among of aqueous extracts, highest concentration (8%) significantly reduced emergence of rice (48.89%) than control red (93.33%). The interaction effect of different concentrations and plant parts showed significant on emergence percentage. Maximum emergence (100%) of red rice was recorded from leaf extract having concentration of 4 and 8% but at par with stem extract with 1 and 2% concentrations while maximum inhibition in emergence were recorded when red rice seeds treated with fruit aqueous extract of opium having 8% concentration. Emergence inhibition of red rice was increase with the increase in aqueous extract concentrations. Our consequences are in

conformity with the observations of Jabran et al. (2015) stated that test species showed significant inhibitory effect on the emergence percentage when compared with lower concentrations, higher concentrations of significantly reduced extract the emergence percentage. Nadeem et al. (2020b) reported that Water extracts of leaf of *C. tinctorius* at 8% concentration result in lowest E. cruss-galli emergence index.

2. Emergence index of O. punctata

Emergence index (EI) was intended to assist the germination rate of germinating seed with respect to time. Aqueous extracts of *P. somniferum* significantly influenced the emergence index of *O. punctata* seedlings presented in Table-2. The highest emergence index of O. punctate seedlings was observed under control at 0% (5.03) i.e. which is similar statistically to all the concentrations and 8% except 1 concentration, while the lowest emergence index (3.07) observed at 8% which however was at par with 1%. Plant parts of *P. somniferum* have significant effect on emergence index of O. punctata seedlings. Leaves produced the stimulatory effect on emergence index of *O. punctata* seedlings whereas the fruit have ultra-low dose response at lower concentrations up to 0.5%. Fruits dose effect have low on all concentrations. Stem also have at inhibitory influence 1%. The interaction between different plant parts and their different concentrations was also significant. At all concentrations the leaf extract resulted in hiahest emergence index (9.15) while the fastest emergence was examined with fruit extracts at almost all concentration. Leaves gave the highest values of emergence index at 4%. Leaves have stimulatory effect on all concentrations and the fruit extract showed inhibitory influence on all concentrations. Stem and fruit produced the low dose response at lower concentration up to 0.5%. Like our results (Rashid et al., 2008) stated that aqueous extracts of *S*. *marianum* aqueous significantly reduced the germination index of test species than control and an increase in inhibitory effect was observed by enhancing the

extracts concentration. Khan et al. (2011) perform experiment to observe the impact of *S. marianum* aqueous extracts on *Glycine max*, *P. vulgaris*, *C. arietinum*, and *V. radiata* germination. By the influence of extracts of test species significantly decreased the germination index as compared to the control and increase in inhibitory effect was observed by enhancing the extracts concentration

3.Time to 50% emergence of *O. punctate* (days)

It is an important parameter regarding seed emergence with respect to time (Table-3). Water extracts of P. *somniferum*on exert significantly allelopathic influence on time taken for 50% germination of *O.* punctata seedlings. The different concentrations Ρ. of somniferum produced nonsignificant effect. The more time taken to 50% emergence of O. punctata (3.56 days) seedlings was given by 2% concentration whereas less time to 50% emergence was recorded at 0.25% (2.74 days). Among various parts of plant, maximum days (3.67 days) was taken by the treatment that received stem aqueous extract while leaf and fruit are statistically at par. The interaction effect of different plant parts was found significant. Stem extract of Ρ. *somniferum* produced the stimulatory effect regarding all concentrations. Highest value of time taken to 50% emergence at 8%, fruit extract produced the enhancing effect at higher (1.50)concentrations days) and maximum days was taken by the treatment receiving aqueous extract of (5.42 days) having 8% stem concentration Leaf also showed the low dose response for time to 50% emergence. Allelopathic effect of R. dentatus water extracts was shown effective in enhancing the time to 50% germination of Helianthus annus and seedlings of T. aestivum. By the effect of extracts of *R*, *dentatus*, an increase in time to 50% germination was obtained at higher concentration when compared with control (Anjum and Bajwa, 2005). According to Nadeem et al. (2020a) who reported that all the concentrations of C. *tinctorius* enhance the time to complete 50% emergence of *O. punctata* with 8%

concentration Similar inhibitory effects of aqueous extracts

4. Mean emergence time of *O. punctate* (days)

Data related to mean emergence time (MET) were presented in Table-4. Among various plant parts extracts, the leaf extract resulted in higher mean emergence time (9.07 days) while the fastest emergence was examined with fruit extracts (3.46 days) all at concentration. Among various concentrations of aqueous extract showed non-significant results while the interaction among plant parts and different concentrations showed, fruit extract with 8% concentration promote the MET of red rice (1.67 days). Leaves extract of P. somniferum at 2% concentration delayed MET of red rice (9.18 days) whereas statistically at par with all other concentrations of leave extract. The growth inhibitory effect of leaf might be due to the presence of chemicals that inhibited the growth of germinating seeds. Similar to our observations (Rose and Anitha, 2012) directed that aqueous extracts of *E.hrita* at different concentration produced inhibitory impact on groundnut and mean emergence time was inhibited by different plant parts and extracts concentration. More delayed germination (higher MET) of rice with higher (5%) than lower (2.5%) concentration of aqueous extract of Vicia sativa has also been reported by previous researchers (Zohaib et al., 2014).

5. Shoot length of *O. punctate* (cm)

Shoot length of *O. punctate* (red significantly influence by the rice) aqueous extracts of different plant parts of *P. somniferum* (opium) and their concentration (Table-5). The smallest shoot length (2.78 cm) was observed among different plant parts with the aqueous extracts of fruit of opium whereas, longest shoot lengths (3.51 cm) were recorded under stem aqueous extracts of P. somniferum. With the increase in aqueous extract concentrations, the shoot lengths were reduced. However, up to 0.5% the influence of increased concentration was non-significant. Whereas, with each unit increased in concentration from 1% to 8% the shoot length was decreased. The interaction among different concentration and plant parts was also significant. It is observed that the concentration which was kept as control gave the longest shoots (4.01 cm) that might be statically similar with the leaf extract application at the concentration of 0.25%. The stem extract of opium had stimulatory influence on the shoot length of red rice at lower concentrations at 2.00% concentration. The fruit and leaf extracts gave the stimulatory influence at lower extract concentrations. The applications of lower concentration of leaves extracts caused significantly higher concentration in associated with control. While, the application of fruit extract enhanced the shoot length up to the concentration of 0.5%. The fruit and leaves showed the stimulatory effect at low concentrations. Might have been due to hormetic effect the shoot length enhanced. The lower concentration of aqueous extract of different plant parts of opium chemicals might be act as hormones for red rice to enhance its growth. Same results were presented by Cheema et al. (2003) the lower concentrations hormetic influence of aqueous extracts of different plant parts as they act as hormones for plant growth. The shoot length was inhibited by the inhibitory influence of opium water extracts has also been stated by the verdicts of Khalig et al., (2009). The delayed germination and slow growth of seedlings can be attributed to the reduction in shoot length. The significant modifications were detected between water extract of different plant parts concerning shoot length. Baber et al. also (2009)supported that the differences in allelopathic potential of various plant parts were significant and extreme values of shoot length were recorded with stem.

6. Root length of *O. punctata* (cm)

Different plant parts and their different concentration of P. somniferum (opium) significantly influenced the root length of O. punctata (Table-6). However, among extract of parts showed various plant nonsignificant results. With the rise in aqueous extracts concentration, the root lengths were decreased. However, up to 0.5% the influence of increased

concentration was non-significant. However, increase in concentration from 1% to 8% the root length was decreased. Among different concentrations highest root length was observed in control (2.85 cm) treatment whereas lowest root length (0.88 cm) was observed from the treatment received aqueous extract with 4% concentration. The root length was inhibited by the inhibitory influence of opium water extracts has also been described by the verdicts of Khalig et al. (2009). The delayed germination and slow growth of seedlings can be attributed to the reduction in root length. Ahmad et al. (2014) also that the differences in supported allelopathic potential of various plant parts were non-significant and extreme values of root length were recorded with stem. Nadeem et al. (2020b) revealed that foliar application of *C. tinctorius* leaf extract inhibits the root length of barnyard grass.

7. Fresh weight of *O. punctata* (g)

Aqueous extracts of various plant parts and their different concentration of P. somniferum had significant influence on fresh weight of *O. punctata* (Table-7). Various plant parts, produce non-significant effect while in case of various concentrations of aqueous extract significantly reduced the red rice fresh weight was observed. Maximum value of fresh weight (56.29 q) was recorded from the treatment that were received aqueous extract having 2% concentration however statistically at par with all other concentration except 8%. While lowest fresh weight (20.22 g) was noted from the 8% Among concentration. interaction between various plant parts of opium and different concentrations the highest fresh weight was observed from stem extract having 2% concentration while fruit extract having 8% concentration killed, he tests species, it's may be due to the presence of allelochemicals that inhibit the red rice growth. Our findings are similar with the observations of Khasraw et al. (2016) documented that seedling growth of various crops was reduced by the influenced of aqueous extracts of opium. The less germination

386

and slow growth of seedlings can be attributed to the reduction in fresh weight. The non-significant modifications were detected between water extract of different plant parts concerning fresh weight. Chon *et al.* (2005)also supported that the differences in allelopathic potential of various plant parts were non-significant and extreme values of fresh weight were recorded with stem extract as seedlings grow more on stem extract application than other opium plant parts extracts.

8. Dry weight of *O. punctate* (g)

Dry weight is a vital indicator of dry matter production of plants as affected by the different allelochemicals. Different parts of plant and their different concentration of aqueous extracts of P. somniferum had significant influence on the dry weight of *O. punctate* (red rice) (Table-8). The less dry weight (3.72 g) was recorded among different plant parts with the leaves aqueous extracts of opium because of reduced growth of red rice seedlings whereas, higher values of dry weight (20.86 g) were examined with the foliar application of fruit aqueous extracts of P. somniferum. With the increase in concentration of aqueous extracts, the dry weight of red rice was decreased as at higher concentrations seedlings growth was gone to reduce due to allelopathic effects of allelochemicals of opium at higher concentration. However, up to 0.5% the influence of increased concentration was non-significant. However, with each unit increase in concentration from 1% to 8% drv weight was decreased. Lowest dry matter (2.78 g) was recorded from the high concentrated 8% agueous extract while highest dry matter was produced from the treatment that were received 2% extract (19.79 g). The lower doses produced higher biomass as compared to the untreated check perhaps due to hormesis (Table-8). The interaction among different concentrations and plant parts was nonsignificant. It is recorded that the concentration which was kept as control gave the higher values of dry weight due

to no interference of allelochemicals at 0.00% concentration which is actually distilled water called control that might be statically similar with the stem extract application at the concentration of 0.25%. According to findings of Nadeem et al., (2020b) that application of aqueous extracts of different C. tinctorius parts at higher concentration (8%) result in reduction in seedlings dry weight of E. cruss-galli. Cseke et al. (2016) also supported that the differences in allelopathic potential of various plant parts were significant and extreme values of dry weight were recorded with fruit as seedlings grow more on fruit extract application than other opium plant parts extract.

Allelopathic effect of Ρ. *somniferum* was observed on the emergence and initial seedling growth of O. punctata. The aqueous extracts of P. somniferum exert inhibitory and also in some cases stimulatory allelopathic influence (hormesis) on root length, shoot length, fresh weight and dry weight as well as on emergence of O. *punctata* weed that was depending upon the concentration of extracts. It is summed up from this study that the phyto-chemicals present in this crop at hiaher concentration can help in biological control of O. punctata weed.

CONCLUSIONS

Ρ. Allelopathic effect of *somniferum* was observed on the germination and initial seedling growth of *O. punctata*. In this research work the aqueous extracts of P. somniferum exert inhibitory and also in some cases stimulatory allelopathic influence on root length, shoot length, fresh weight and dry weight as well as on germination of О. punctata depending upon the concentration of extracts. It is summed up from this study that the phytochemicals present in this crop at 8% concentration help in biological control of O. punctata weed.

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388

Table-1.	Emergence	percentage	of	О.	punctata	as	influenced	by	aqueous	extract
concentra	tions of leaf,	stem and fru	iits	of F	P. somnifer	um.				

	Emergence percentage (%)									
Plant	Plant Concentration									
Parts	Control	0.25%	0.5%	1%	2%	4%	8%			
Leaf	86.67ab	93.33ab	80.0abc	73.33a-d	86.67ab	100.00a	100.00a	88.57a		
Stem	93.33ab	86.67ab	100.00a	100.00a	93.33a	93.33a	40.00e	86.67a		
Fruit	100.00a	86.67ab	86.67ab	53.33cde	66.67b-e	46.67de	6.67f	63.81b		
Mean	93.33a	88.89ab	88.89ab	75.56b	82.22ab	80.00ab	48.89c			

Means not sharing a letter in common differ significantly at 5% level of significance. HSD: Concentration = 16.727, Plant parts = 2.107, Concentration × plant parts = 19.781

Table-2. Emergence index of *O. punctata* as influenced by aqueous extract concentrations of leaf, stem and fruits of *P. somniferum*.

	Emergence index								
Plant	Plant Concentration								
Parts	Control	0.25%	0.5%	1%	2%	4%	8%	-	
Leaf	7.47bcd	8.32abc	6.78cd	5.98d	6.99bcd	9.15a	8.65ab	7.62a	
Stem	3.44ef	3.31efg	3.83e	4.11e	3.44ef	3.53ef	0.48hi	3.16b	
Fruit	4.17e	3.56ef	3.42ef	1.69ghi	2.06fgh	1.67hi	0.08hi	2.35c	
Mean	5.03a	5.06a	4.68ab	3.93bc	4.17ab	4.72ab	3.07c		

Means not sharing a letter in a common differ significantly at 5% level of significance. HSD: Concentration = 0.894, Plant parts = 0.725, Concentration × plant parts = 0.863

Table-3. Time to 50% emergence of *O. punctata* as influenced by aqueous extract concentrations of leaf, stem and fruits of *P. somniferum*.

	Time to 50% emergence (days)								
Plant			Co	Concentration					
Parts	Control	0.25%	0.5%	1%	2%	4%	8%	-	
Leaf	2.83bcd	2.67bcd	2.75bcd	3.17bc	3.25bc	2.50cd	2.50cd	2.81b	
Stem	3.06bc	3.25bc	3.17bc	3.50bc	3.42bc	3.92b	5.42a	3.67a	
Fruit	3.08bc	2.31cd	3.00bc	2.33cd	4.00b	3.42bc	1.50d	2.81b	
Mean	2.99 ^{NS}	2.74	2.97	3.00	3.56	3.28	3.14		

Means not sharing a letter in a common differ significantly at 5% level of significance. HSD: Concentration= NS , Plant parts = 0.482, Concentration × plant parts = 1.411 NS = Non-significant

Table-4. Mean emergence time of *O. punctata* as influenced by aqueous extract concentrations of leaf, stem and fruits of *P. somniferum.*

	Mean emergence time (days)							
Plant	Concentration							
Parts	Control	0.25%	0.5%	1%	2%	4%	8%	_
Leaf	9.08a	8.96a	9.09a	9.16a	9.18a	8.90a	9.06a	9.07a
Stem	3.70cde	3.87cde	3.87cde	4.27cd	3.84cde	4.31cd	5.92b	4.25b
Fruit	3.77cde	3.11def	3.62de	2.78ef	4.78bc	4.48bcd	1.67f	3.46c
Mean	5.52a	5.31a	5.53a	5.40a	5.94a	5.89a	5.55a	

Means not sharing a letter in a common differ significantly at 5% level of significance. HSD: Concentration= NS , Plant parts =0.535, Concentration × plant parts = 1.441 NS = Non-significant

	Shoot length (cm)								
Plant			Co	oncentratio	on			Mean	
Parts	Control	0.25%	0.5%	1%	2%	4%	8%		
Leaf	4.52ab	3.95a-d	3.77a-d	3.66 a-e	2.62d-g	2.26e-h	1.18gh	3.14ab	
Stem	3.16b-f	3.87a-d	3.96a-d	4.17abc	5.05a	2.54d-g	1.80fgh	3.51a	
Fruit	4.37abc	3.80a-d	3.55b-e	1.76fgh	2.89c-f	2.27e-h	0.80h	2.78b	
Mean	4.01a	3.87a	3.76a	3.19ab	3.52a	2.35b	1.26c		

Table-5. Shoot length of *O. punctata* as influenced by aqueous extract concentrations of leaf, stem and fruits of *P. somniferum*.

Means not sharing a letter in a common differ significantly at 5% level of significance. HSD for concentration = 1.17, for plant parts = 0.73, for Interaction = 1.263

Table-6. Root length of *O. punctata* as influenced by aqueous extract concentrations of leaf, stem and fruits of *P. somniferum*.

	Root length (cm)								
Plant			Co	oncentratio	on			Mean	
Parts	Control	0.25%	0.5%	1%	2%	4%	8%	-	
Leaf	3.97a	1.76bc	1.14c-g	1.36cde	1.06c-h	0.72e-i	0.48f-i	1.50 ^{NS}	
Stem	1.42cde	1.73bc	1.48b-e	1.26c-f	2.27b	1.53b-e	0.30hi	1.43	
Fruit	3.18a	2.28b	1.58bcd	0.88d-i	0.81c-i	0.38ghi	0.13i	1.32	
Mean	2.85a	1.92b	1.40c	1.17cd	1.38c	0.88d	0.30e		

Means not sharing a letter in a common differ significantly at 5% level of significance. HSD: Concentration = 0.518, Plant parts = NS , Concentration × plant parts = 0.911 NS = Non-significant

Table-7. Fresh weight of *O. punctata* as influenced by aqueous extract concentrations of leaf, stem and fruits of *P. somniferum*.

	Fresh weight (g)								
Plant			Co	oncentratio	n			Mean	
Parts	Control	0.25%	0.5%	1%	2%	4%	8%		
Leaf	67.33ab	51.83a- d	49.33a- d	53.61a-d	38.44bcd	34.66cd	27.33de	46.07 ^{NS}	
Stem	41.33bcd	47.33a- d	46.66a- d	60.00abc	74.00a	48.83a- d	33.33cd	50.21a	
Fruit	41.33bcd	58.00a- d	53.33a- d	54.55a-d	56.44a-d	45.55a- d	26.00e	44.17a	
Mean	50.00a	52.38a	49.77a	56.05a	56.29a	43.01a	20.22b		

Means not sharing a letter in a common differ significantly at 5% level of significance. HSD: Concentration = 6.982, Plant parts = NS , Concentration× plant parts = 28.447 NS = Non-significant

Table-8. Dry weight of *O. punctata* as influenced by aqueous extract concentrations of leaf, stem and fruits of *P. somniferum*.

	Dry weight (g)							
Plant	Concentration							
Parts	Control	0.25%	0.5%	1%	2%	4%	8%	-
Leaf	7.00 ^{NS}	3.60	4.38	3.91	3.60	3.06	1.13	3.72c
Stem	11.00	13.00	12.00	20.00	27.67	14.16	7.22	15.01b
Fruit	17.33	28.00	25.00	25.83	28.78	21.11	0.99	20.86a
Mean	11.78b	14.86ab	13.79ab	16.58ab	19.79a	12.78ab	2.78c	

Means not sharing a letter in a common differ significantly at 5% level of significance. HSD: Concentration = 7.005, Plant parts = 2.456, Concentration × plant parts = NS = Non-significant



Figure.1: Interaction between treatment means of shoot length, root length, fresh weight and dry weight of *O. Punctata* under the influence of aqueous extracts of different parts of *P. Somniferum*.

REFRANCES CITED

- Ahmad, W., M. Akbar, U. Farooq, A. Alia and F. Khan. 2014. Allelopathic effects of aqueous extracts of *Avenafatua* on seed germination and seedling growth of *Triticum aestivum* (variety GW-273). J. Envir. Sci. Tox. Food Tec., 8(1): 38-42.
- Alam, S.M. and E. Islam. 2002. Effect of aqueous extract of leaf, stem and root of nettle leaf goose foot on germination and seedling growth of rice. Pak. J. Seed Technol., 2(1): 47-51.
- Anjum, T. and R. Bajwa 2005. A bioactive annuionone from sunflower leaves. Phytochemistry, 66(1): 1919-1921.
- Anonymous. 2005. Weed Science Society of Pakistan. www. wssp.org.pk
- Baber, B.H., A. Tanveer, A. Aziz, M.M Javaid, M. Tahir, M. Sibtain and Z. Pacanoski. 2009. Phytotoxic influences of *Asphodelus tenuifolius* Cav (wild onion) on germination and seedling growth of wheat. Allelopathy J., 24(2): 341-350.
- Bajwa, A.A. 2014. Sustainable weed management in conservation agriculture. Crop Prot, 65(1): 105-113.
- Cheema, Z.A., S. Hussain and A. Khaliq. 2003. Efficacy of sorgaab? in combination with allelopathic water extracts and reduced rates of pendimethalin for weed control in mungbean (*Vigna radiata* L.). Indus J. Plant Sci., 2(1): 21-25.
- Chon, S.U., H.G. Jang, D.K. Kim,Y.M. Kim, H.O. Boo and Y.J. Kim. 2005. Allelopathic potential in lettuce (*Lactuca sativa* L.) plants. Sci. Hortic., 106(3): 309-317.
- Chung, I.M., J.T. Kim and S.H. Kim. 2006. Evaluation of allelopathic potential and quantification of momilactone A, B from rice hull extracts and assessment of inhibitory bioactivity on paddy field weeds. J. Agric. Food Chem., 54(7): 2527-2536.
- Cirujeda, A., J. Recasens, J. Torra and A. Taberner. 2008. A germination

study of herbicide-resistant field poppies in Spain. Agron. Sustain. Dev., 28(2): 207-220.

- Coolbear P., A. Francis and D. Grierson. 1984. The effect of low temperature pre-sowing treatment on the germination performance and membrane integrity of artificially aged tomato seeds. J. Exp. Bot., 35(11): 1609-1617.
- Cseke, L.J., A. Kirakosyan, P.B. Kaufman, S. Warber, J.A. Duke and H.L. Brielmann. 2016. Natural products from plants, CRC press.
- Ellis R.A. and E.H. Roberts. 1981. The quantification of aging and survival in orthodox seeds. Seed Sci. Technol., 9(2): 373-409.
- Ferreira, M.I. and C.F. Reinhardt. 2010. Field assessment of crop residues for allelopathic effects on both crops and weeds. Agron. J., 102(6): 1593-1600.
- Gallandt, E.R. and J. Weiner. 2001. Crop-weed competition. Els, 1-9.
- Ghafarbi, S.P., S. Hassannejad and R. Lotfi. 2012. Allelopathic effects of wheat seed extracts on seed and seedling growth of eight selected weed species. Int. J. Agri. Crop Sci., 4(19): 1452-1457.
- Ghodake, S.D., M.D. Jagtap and M.B Kanade. 2012. Allelopathic effect of three Euphorbia species on seed germination and seedling growth of wheat. Ann. Biol. Res., 3(10): 4801-4803.
- Jabran, K., G. Mahajan, V. Sardana and B.S. Chauhan. 2015. Allelopathy for weed control in agricultural systems. Crop Prot., 72(1): 57-65.
- Javaid, A., T. Anjum and R. Bajwa. Biological 2005. control of Parthenium II: Allelopathic effect of Desmostachyabipinnata on distribution and early seedling Parthenium growth of *hysterophorus* L. Int. J. Biol. Biotec., 2(2): 459-463.
- Kadioglu, I., Y. Yanar and U. Asav. 2005. Allelopathic effects of weeds extracts against seed germination of some plants. J. Environ. Biol., 26(2): 169-173.
- Khaliq, A., A. Matloob, M.N. Mushtaq and F. Aslam. 2009. Inhibitory effects

of sorghum and sunflower water extracts on germination and growth of Cichoriumintybus L. Proceeding, 1st Asian Allelopathy Society (AAS), December, 18(22), 110-125.

- Khan, R., M.A. Khan, M. Waqas, A.M. Khan, Z. Hussain, A. Khan and M.A. Raza. 2011. Allelopathic potential of *Silybum marianum* L. against the seed germination of edible legumes. Pak. J. Weed Sci. Res., 17(3): 293-302.
- Khasraw, M.N., K.M. Mustafa, H.M. Aziz, K.F. Mustafa, S.A. Ahmad and F.F. Khorsheed. 2016. Allelopathic effects of Euphorbia helioscopia and Papaver rhoeas on germination and initial seedling growth of cereal crops and weed seeds. J. Zank. Sul., 18(1): 145-152.
- Labanca, F., J. Ovesna and L. Milella. 2018. *Papaver somniferum* L. taxonomy, uses and new insight in poppy alkaloid pathways. Phytochem. Rev., 17(4): 853-871.
- Macias, F.A., R.M. Varela, A. Torres, J.L. Galindo and J.M. Molinillo. 2002. Allelochemicals from sunflowers: chemistry, bioactivity and applications. In Chemical ecology of plants: allelopathy in aquatic and terrestrial ecosystems, pp. 73-87.
- Melander, B., I.A. Rasmussen and P. Barberi 2005. Integrating physical and cultural methods of weed control—examples from European research. Weed Sci., 53(3): 369-381.
- Nadeem, M.A., B.A. Khan, S. Afzal, A. Aziz, R. Maqbool, M.M Amin, A. Aziz, A. Ali, M. Adnan and Durrishahwar. 2020b. Allelopathic Effects of aqueous extracts of *Carthamus tinctorius* L. on emergence and seedling growth of *Echinochloa crus-galli* L. Pak. J. Weed Sci. Res., 26(3): 365-379.
- Nadeem, M.A., B.A. khan, S. Afzal, M.A. Khan, T. Abbas, M.M. Javaid, M.M. Amin, N. Farooq and A. Aziz. 2020a. Effect of aqueous extract of *Carthamus tinctorius* L. on germination and initial seedling

growth of *Oryza punctata* L. Pak. J. Weed Sci. Res., 26(3): 331-342.

- Rashid, H., M.A. Khan, A. Amin, K. Nawab, N. Hussain and P.K. Bhowmik. 2008. Effect of *Parthenium hysterophorus* L. root extracts on seed germination and growth of maize and barley. Am. J. Plant Sci. Biotec., 2(2): 51-55.
- Ravlic, M., R. Balicevic, M. Knezevic and I. Ravli. 2012. Allelopathic effect of scentless mayweed and field poppy on seed germination and initial growth of winter wheat and winter barley. Herbologia., 13(2): 1-7.
- Rose, M.L. andS. Anitha. 2012. Effect of *Euphorbia hirta* L. extract on the germination and seedling growth of groundnut. Biotechnol. Adv., 12(1): 27-29.
- Skrzypek, E., P. Repka, A. Stachurska-Swakon, B. Barabasz-Krasny and K. Mozdzen. 2015. Allelopathic effect of aqueous extracts from the leaves of peppermint (Mentha piperita L.) on selected physiological processes of common sunflower (Helianthus annuus L.). Not. Bot. Horti. Agrobiol, 43(2): 335-342.
- Turk, M.A. and A.R.M.Tawaha. 2002. Impact of seeding rate, seeding date, and method rate of phosphorus application in faba bean (Viciafaba L. minor) in the of absence moisture stress.Biotechnol. Agron. Soc. Environ., 6(3):171–1.
- Zohaib, A., Tanveer, A., Khaliq, A. and Safdar, M. E. (2014). Phytotoxic effect of water soluble phenolics from five leguminous weeds on germination and seedling growth of rice. Pak. J. Weed Sci. Rese., 20(4): 417-429.