

Research Article



Exploring Enhanced Insecticidal Activity of Mycelial Extract of *Trichoderma harzianum* against *Diuraphis noxia* and *Tribolium castaneum*

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Abstract | Fungi are important organisms that produce potent secondary metabolites with antimicrobial, herbicidal, insecticidal and other such beneficial activities. Production of such compounds can be increased or decreased depending on growth conditions. For these reasons a *Trichoderma harzianum* was selected for its growth optimization conditions and extraction of insecticidal compounds effective against two major insect pests including *Diuraphis noxia* and *Tribolium castaneum*. The pure fungal strain of *Trichoderma* was grown using two different nutrient media including Glucose Peptone Yeast Broth (GPYB) and Potato Dextrose Broth (PDB) and the effect of organic extract was investigated against selected insects' pest. From this study it was found that the mycelial extract of *T. harzianum* fermented on PDB showed enhanced insecticidal mortality (73% and 76%) against *Diuraphis noxia* and *Tribolium castaneum* respectively while the extract obtained from GPYB media showed merely 46% and 56% mortality against *Diuraphis noxia* and *Tribolium castaneum* respectively at highest concentration after 2 hours exposure. Increased potency of extracts was observed with the increased exposure time. The LC_{50} value calculated for organic extract obtained from *T. harzianum* grown on PDB against *Diuraphis noxia* was $398.69 \mu\text{g mL}^{-1}$ after 3 hours with increased potency ($237.46 \mu\text{g mL}^{-1}$) after 24 hours, whereas against *Tribolium castaneum* it was $213.85 \mu\text{g mL}^{-1}$ after 24 hours while its decreased potential was recorded ($1153.5 \mu\text{g mL}^{-1}$) after 3 hours. Similar pattern of toxicity level was observed for organic extract exposure of *T. harzianum* grown on GPYB viz increasing exposure time increased the mortality rate, The LC_{50} value against *Diuraphis noxia* was reduced to $1083.49 \mu\text{g mL}^{-1}$ after 24 hours from $2269.08 \mu\text{g mL}^{-1}$ after 3 hours, whereas against *Tribolium castaneum* it reduced significantly from $1849.69 \mu\text{g mL}^{-1}$ after 3 hours to $677.21 \mu\text{g mL}^{-1}$ after 24 hours. From the results it is concluded that *T. harzianum* grown on PDB produced potent secondary metabolites that could be used for the development of target specific insecticides against *Diuraphis noxia* and *Tribolium castaneum*. Further studied towards the isolation and structural elucidation of lead compounds is recommended for the development of local agro-chemical product as environmental friendly biodegradable organic insecticides.

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Introduction

Agricultural productivity is adversely affected by insects' pathogen as they damage both standing

crops as well as stored commodities. These insect pests can be controlled by various ways, among them use of insecticides is very common. These insecticides generally contain an active ingredient along with a

carrier substance, the active ingredient can be from chemical or/and biological source. Chemically synthesized pesticides do have some advantage but possess serious threat to environment and can have serious side effects as well, pesticides like Aldrin and Azodrin have been banned and their application limit have reduced in most countries. For this reason, a work towards the development of natural products based agrochemicals is needed. The advantage of naturally occurring bioactive metabolites over synthetic molecules is that they are environmental friendly and least harmful to human health. Natural products or bioactive secondary metabolites are considered as most effective ingredients of potent drugs. These molecules could be obtained from various sources including, plants animals and microbes.

Fungi are diverse group of organisms found in both terrestrial and aquatic environment and can produce variety of bioactive metabolites (Iqbal et al., 2014). Metabolites from microbes are host specific and having broad range of activity (Berdy, 2005). The derivatives of secondary metabolites are used as pesticides having antiparasitic, antihelminthic agents, antifeedant, insecticides, nematocides, cestocides, antiworm compounds, acaricides, phytotoxins, phytohormones (Cole and Rolinson, 1972), plant growth regulators, chlorosis inducers, algicide, germination inhibitors, and as biocontrol agents (El-Hasan et al., 2009). The most useful microbial compounds are monensin, bialaphos and avermectin isolated from different species of *Streptomyces*. Commercially various agricultural products are available which are produced from microorganisms like *Beauveria* species, *Bacillus thuringiensis*, *Metarhizium* species, *Baculovirus*, *Trichoderma* species and *Verticillium* species (Dayan et al., 2009).

Trichoderma spp are free living fungi found in root, soil as well as in foliar environments. It has numerous antibiotic producing and bio-controlling characteristics (Reino et al., 2008a). The isolates of *Trichoderma* are good source of antibiotics having diverse effects against various pathogenic microbes (Chruma et al., 2009). The azaphilone and butenolide isolated from *Trichoderma harzianum* were reported with potent antifungal activities against different yeast (Reino et al., 2008b). Its use as a biocontrol agent was proposed against soil borne fungal and other bacterial pathogens along with its use as a potent bio-pesticide (Osmanova et al., 2010).

The biosynthesis of secondary metabolites depends primarily on growth conditions. This current study was designed to identify cost effective growth medium for the biosynthesis of insecticidal secondary metabolites of *Trichoderma harzianum* effective against *Diuraphis noxia* and *Tribolium castaneum*.

Materials and Methods

Collection of soil samples

Surface level soil samples (from up to a 6 cm soil depth) were collected randomly from Peshawar, Khyber Pakhtunkhwa using standard protocol (Rohilla and Salar, 2012). The collected samples were packed in a sterilized polyethylene bag, labelled properly and stored at 4 °C further use (Gaddeyya et al., 2012).

Isolation of *Trichoderma harzianum* from soil sample

The separation of fungal species from soil samples were carried out using standard serial dilution method under aseptic conditions. The seed suspension was prepared by adding 1 g of soil sample in 10 mL potato dextrose broth (PDB) and the solution was further diluted to 1×10^{-4} concentration suspension. To avoid bacterial infection Streptomycin sulfate ($1 \mu\text{g mL}^{-1}$) was added to each test tube and incubated at 28 °C for 05 days. The obtained fungal species were transferred to the petri dishes containing pre-sterilized potato dextrose agar media (PDA) and streptomycin sulfate ($1 \mu\text{g mL}^{-1}$) and incubated at 30 °C in dark for 7 days. (Gupte et al., 2002) (Atalla et al., 2008). The obtained fungal biomass contained variety of fungal species which was further purified by transferring and growth a colony of each fungi using repeated growth on potato dextrose agar (PDA) as described above till the growth of one specie per petri dish was achieved. The fungal strains were identified by observing it under binocular microscopic through microscopic characteristics (color, texture appearance and diameter of colonies) (Rohilla and Salar, 2012). The *Trichoderma harzianum* selected for growth, extraction of bioactive metabolites and their insecticidal activities against *Diuraphis noxia* and *Tribolium castaneum* was further purified and identified by mycologist at department of plant pathology, The University of Agriculture-Peshawar.

Growth and extraction of fungal biomass

Two different media i.e. potato dextrose broth (PDB) and glucose peptone yeast broth (GPYB) were used for the fermentation of *Trichoderma harzianum* (Iqbal

et al., 2018). The biomass of *Trichoderma harzianum* obtained from both PDA and GPYB was extracted separately by following the procedure described by (Begum et al., 2018). The fermented biomass of *Trichoderma harzianum* was separated from the broth through filtration, washed with distilled water and 10 g was transferred to conical flasks containing 100 mL ethyl acetate (EtOAc). The fungal biomass was homogenized through blender and kept on magnetic stirrer for 24 hours for maximum extraction of biomolecules. The fungal cells were separated via filtration, 50 mL distilled water was added to liquid fraction and transferred to separating funnel. Both aqueous and organic fractions were separated and the aqueous fraction was re-extracted with EtOAc (100 × 3 mL). The combined organic fractions were dried over anhydrous MgSO₄, filtered and concentrated under reduced pressure to obtain brown oil as crude extract. The extract was stored at 4 °C in glass vial for further use.

Insecticidal assay

Organic extract of *Trichoderma harzianum* obtained from both media was separately screened for insecticidal activity (Noshad et al., 2015) against *Diuraphis noxia* and *Tribolium castaneum*. The stock solution (1000 µg mL⁻¹) from crude of each extract was prepared (8 mg of crude per 8 mL of EtOAc) and further diluted to 500 µg mL⁻¹ and 250 µg mL⁻¹ concentration. Each concentration (3 mL) was added to clean petri dish containing a filter paper and solvent was allowed to evaporate at room temperature. Ten insects were transferred in each petri dish along with their normal feed (fresh wheat leaves for *Diuraphis noxia* and grounded wheat flour for *Tribolium castaneum*). The experiments were performed in triplicate and the mortality of insects was recorded at 3 hr and 24 hr interval and the data was compared with both positive control (Malathion 1000 µg mL⁻¹) and negative control (Blank, 0 µg mL⁻¹) experiment. The LC₅₀ was calculated by probit analysis (Finney and Stevens, 1948) and percent mortality was calculated using following formula (Equation 1) (Begum et al., 2018).

$$\text{Mortality \%} = \frac{I_{Ts} - I_b}{T_i} \times 100 \dots (1)$$

Where;

I_{Ts} = Killing of insects by test solution; I_b = Killing of insects by blank; T_i = Total insects.

Statistical analysis

The data collected during the experimental analysis was analyzed using Statistix 8.1® and the result is presented as mean of triplicate. Statistical descriptive comparison of more values for experiment were analyzed by CRD design and ANOVA followed by LSD at 5% level of significance (Steel et al., 1997).

Results and Discussion

Effect of mycelial extract of *Trichoderma harzianum* against *Diuraphis noxia*

Aphids as a vector of plant viruses are considered serious insect pest of various crops including, wheat, tomato, pepper, cucumber etc all over the world. Indiscriminative use of insecticides have also caused resistance among various insects pests. Biological control of aphids using entomopathogenic fungi is an emerging strategy. Fungi produce important bio molecules that can be used for the development of bio-insecticides for the control of insect pests. The organic extract of *T. harzianum* grown on PDB media was tested against *Diuraphis noxia* in three different concentrations. The results clearly shows both time and concentration dependent effect on the mortality of aphids (Table 1), although the results were non-significant in comparison to positive control experiment (Melathion, 1000 µg mL⁻¹) while significantly increased mortality (73% after 24 hours using 1000 µg mL⁻¹ concentration of organic extract of *T. harzianum* grown on PDB media) of *Diuraphis noxia* was recorded in comparison to blank experiment (3.3% after 24 hours). The effect of organic extract of *T. harzianum* fermented on GPYB media showed appreciate mortality against aphids (Table 1). In first 3 hours increased aphids' mortality from 6.66% to 23% was recorded with the increase in concentration of organic extract from 250 µg mL⁻¹ to 1000 µg mL⁻¹. Similarly increase in exposure time of organic extract from 3 hours to 24 hours also showed increased mortality. The highest percent mortality (23.3%) was observed with 1000 µg mL⁻¹ crude extract after 3 hours, which significantly increased (46.6%) after 24 hours. The LC₅₀ calculated after 24 hours (237.46 µg mL⁻¹) clearly shows the high aphicidal potential of organic extract of *Trichoderma harzianum* (Table 1). The LC₅₀ calculated (1083.49 µg mL⁻¹) clearly showed that the organic extract have very least effect on aphids mortality after 24 hours, while as the mortality increased significantly (2269.05 µg mL⁻¹) after three hours. From the results it is concluded

Table 1: Effect of organic extract of *T. harzianum* obtained from PDB and GPYB media against *Diuraphis noxiar-values* in parenthesis show in percentage.

Concentrations ($\mu\text{g mL}^{-1}$)	PDB*		GPYB*		
	After 3hr	After 24hr	After 3hr	After 24hr	
Negative **	0.33f (3.33%)	0.33f (3.33%)	0.33f (3.33%)	0.33f (3.33%)	
250	4.33d (43.3%)	5.00c (50%)	0.66f (6.66%)	2.33e (23.3%)	
500	5.33c (53.3%)	6.33b (63.3%)	1.66e (16.6%)	3.66d (36.6%)	
1000	6.66b (66.6%)	7.33b (73.3%)	2.33e (23.3%)	4.66c (46.6%)	
Positive ***	6.66b (66.6%)	9.66a (96.6%)	7.66b (76.6%)	9.66a (96.6%)	

* Media used for the growth of *T. harzianum*; ** Blank experiment was run same as all except use of organic extract of *T. harzianum*; *** A known insecticide Malathion ($1000 \mu\text{g mL}^{-1}$) was used in positive control experiment; Values with different letters are significantly different ($P < 0.05$).

Table 2: Effect of organic extract of *T. harzianum* obtained from PDB and GPYB media against *Tribolium castaneum* values in parenthesis show in percentage.

Concentrations ($\mu\text{g mL}^{-1}$)	PDB*		GPYB*		
	After 3hr	After 24hr	After 3hr	After 24hr	
Negative**	0.00h (0%)	0.00h (0%)	0.00h (0%)	0.00h (0%)	
250	2.33g (23.3%)	5.33d (53.3%)	1.66g (16.6%)	3.33e (33.3%)	
500	3.33f (33.3%)	6.00c (60%)	2.33f (23.3%)	4.66d (46.6%)	
1000	4.66e (46.6%)	7.66b (76.6%)	3.66e (36.6%)	5.66c (56.6%)	
Positive***	7.66b (76.6%)	9.66a (96.6%)	7.66b (76.6%)	9.66a (96.6%)	

* Media used for the growth of *T. harzianum*; ** Blank experiment was run same as all except use of organic extract of *T. harzianum*; *** A known insecticide Malathion ($1000 \mu\text{g mL}^{-1}$) was used in positive control experiment; Values with different letters are significantly different ($P < 0.05$).

that the organic extract of *T. harzianum* fermented on GPYB media possess statistically less aphidicidal potential as compare to *T. harzianum* grown on PDB media. This is due to the production of more potent bioactive compounds when *T. harzianum* was fermented on PDB. The carbon source available in nutrient media is responsible for the bio-synthesis of secondary metabolites having increases or decreased biological activities. The use of soil borne fungus *Beauveria bassiana* *Conidia* is reported for its effective use to control aphids (Mingguang and Shenghua, 2003). Similarly, Feng has described the formulation of various fungi including *Omuraea rileyi*, *Paecilomyces fumosoroseus*, and *Verticillium lecanii* for the control of insects' pests (Dun et al., 2003). Copping and Duke have reported the efficient use mycelium of *Myrothecium verrucaria* as a potent nematocidal highly effective against the plant parasitic nematodes residing the field of tobacco, grapes, citrus etc (Copping and Duke, 2007).

Effect of mycelial extract of Trichoderma harzianum against Tribolium castaneum

Many of the existing insecticides and other pesticides were originated initially from various

biological sources. Even now natural products have been considered as an important source for the development of new agrochemicals. Recently Sparks et al have reported the importance of fungal natural products towards the discovery and formulation of new insecticidal agrochemicals (Sparks et al., 2017). The effect of crude extract of *T. harzianum* mycelia against red flour beetle (*Tribolium castaneum*) is presented in Table 2. Results showed that the mortality rate of red flour beetle by fungal extract (all tested concentrations) had shown significantly increased mortality as compare to negative control experiment. The ethyl acetate extract of *T. harzianum* obtained from the mycelia cells cultured on PDB media showed overall improved insecticidal activity against *Tribolium castaneum* in comparison to mycelia cells cultured on GPYB media (Table 2). The mortality of *Tribolium castaneum* significantly increased with the increase in concentration (from $250 \mu\text{g mL}^{-1}$ to $1000 \mu\text{g mL}^{-1}$) and exposure time (3 hours to 24 hours). The organic extract of *T. harzianum* grown on GPYB culture media showed 56.6% mortality of *Tribolium castaneum* (at $1000 \mu\text{g mL}^{-1}$ concentration) whereas significantly improved mortality (76.6%) was recorded for mycelium extract

of *T. harzianum* fermented using PDB media. The LC_{50} values clearly shows that the insecticidal effect of extract obtained from *T. harzianum* grown on PDB media increased from $1153.5 \mu\text{g mL}^{-1}$ after 3 hours to $213.85 \mu\text{g mL}^{-1}$ after 24 hours which suggested that the bioactive molecules present in the extract possess appreciable activity against red flour beetle. In comparison the LC_{50} efficacy (Table 2) of organic extract of *T. harzianum* grown on GPYB media was 3 fold less ($677.21 \mu\text{g mL}^{-1}$) at highest tested concentration after 24 hours of exposure. Two fungal strains i.e. *Beauveria bassiana* and *Verticillium* were reported for the mortality of red flour beetles, where *Beauveria bassiana* showed 61.67% mortality through infection via adhesion, spore germination and mycelium colonization in cadavers of *Tribolium castaneum* (Hanan, 2017). Another study shows the significant effect of *B. bassiana* isolate 22292A against three stored grain pests including red flour beetles (Cogburn and Rice, 1999) where up to 100% mortality of pests was achieved.

Conclusions and Recommendations

Secondary metabolites are bio-molecules with significant importance in routine life, these compounds are produced by the living organisms such as plants, animals and microbes. The exploration of biologically potent secondary metabolites where fungi is known to produce more potent compounds. These bioactive molecules can be used for the development of various pharmaceuticals as well as Agro chemicals. From present study it is concluded that the biosynthesis of insecticidal molecules by *T. harzianum* depends on growth media. The organic extract of *T. harzianum* grown on potato dextrose broth in comparison to glucose peptone yeast broth showed improved insecticidal activities against both *Diuraphis noxia* and *Tribolium castaneum*. The organic extract of *T. harzianum* might contain more potent biomolecule as a good candidate towards the development of environmental friendly insecticides for the control of both *Diuraphis noxia* and *Tribolium castaneum*. To our knowledge this is the initial report for the control of *Diuraphis noxia* and *Tribolium castaneum* using organic extract of *T. harzianum*. Further studies on the isolation of targeted compound(s) is needed towards the

development of bio-based insecticides.

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Novelty Statement

The use of biological extract towards the control of various pathogens is the need of the day. To our knowledge this is the first report where the organic extract of *Trichoderma harzianum* is used for the control of apids and red flour beetles.

Author's Contribution

Mudassar Iqbal: Principal author, conceive the idea, planned and supervised the research.

Sadaf Rahim Awan: Researcher, conducted the research, collected the data.

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