

# Effect of Different Environmental Stresses on the Antioxidant Capability of Air-Breathing Loach, *Paramisgurnus dabryanus* (Sauvage, 1878)

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## ABSTRACT

The effect of oxidative stress on gill and gut enzyme activity in *Paramisgurnus dabryanus* was studied under different breathing restraint treatments. A total of 150 healthy mature individuals (mean initial body weight =  $16.1 \pm 3.7$  g and mean initial total fork length =  $13.1 \pm 1.2$  cm) were selected for the experiment. The experimental specimens were separated into three modes: control group (n = 50) without any stress treatments; inhibited group (n = 50) with gut respiratory inhibition; and an air-exposed group (n = 50) with gill respiratory inhibition. After a 7 d acclimation for the inhibited and air-exposed groups, individuals were returned to the same conditions as the control group for a further 3 d acclimation. The catalase (CAT) and superoxide dismutase (SOD) activity in the gill and gut in *P. dabryanus* were examined in all three groups after the 7 d stress acclimation, 1 d later, and after a 3 d recovery. It was found that CAT and SOD activity in the gill, foregut, and midgut in the air-exposed group were much lower than in the control group. After recovery for 1 d, CAT activity in the gill and middle gut increased rapidly and was higher than in the inhibited and control groups. The SOD activity in the gill and gut also significantly increased, although there was no significant difference between the air-exposed and control groups. After a 3 d recovery, CAT and SOD activity in the gill and gut of *P. dabryanus* did not differ significantly between groups. There was a high viability of *P. dabryanus* under the air-exposed condition (in anaerobic conditions), while the level of oxidative stress in the gill, anterior, and middle gut clearly decreased. When recovering under normoxia conditions, the physical level of oxidative stress quickly returned to normal levels.

## Article Information

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## Authors' Contribution

YL and ZW designed the study. YL and JZ performed the experiments and analyzed the data. ZW and WC helped in experimental work and data analysis. YL and XL wrote the article.

## Key words

Air-exposed, Gut, Respiratory inhibition, Oxidative stress, *Paramisgurnus dabryanus*

## INTRODUCTION

The oxygen concentration in aquatic ecosystems has a direct impact on fish. It is often affected by diurnal changes in environmental factors, water temperature changes, and drought. Many fish are very sensitive to small changes in the oxygen concentration. To supplement oxygen consumption during aquatic hypoxia, some fish have evolved several adaptive strategies in terms of structure, physiology, and behavior (Catling *et al.*, 2001; Gonzales *et al.*, 2006; Huang *et al.*, 2015). For example, members of the Loricariidae and the Callichthyidae families rely on specific tissues, such as vascularized stomach or gut tissue, for aerial gas exchange under hypoxic conditions (Bickler *et al.*, 2007; Pelster *et al.*, 2016).

It has been reported that there are more than 400 species of fish that have an accessory organ for air-breathing (Graham, 2011). Some accessory air-breathing organs are

located in the digestive tract, such as the esophagus, stomach and gut (Podkowa and Goniakowska-Witalinska, 2002). Gut air-breathing as an accessory respiratory approach are mainly subordinate to Cobitidae and Callichthyidae (Nelson, 2014). Studies of the gut air-breathing of fish such as *Misgurnus anguillicaudatus* (Goncalves *et al.*, 2007), *Lepidocephalichthys guntea* (Moitra *et al.*, 1989), *Hoplosternum thoracatum* (Huebner *et al.*, 1978), *Corydoras aeneus* (Podkowa *et al.*, 2002) have been reported. *Paramisgurnus dabryanus* relies on the gut for assisted respiration, and its accessory organs are located in the posterior part of the gut, which implies regional modifications of the gut for gas exchange. Like many other air-breathing fish, the respiratory structures of *P. dabryanus* are vascularized and have a thin diffusion distance between the external and internal mediums (Moitra *et al.*, 1989; Goncalves *et al.*, 2007).

Gut air-breathing, evolved as a physiological behavior and affects the fish's normal physiological and behavioral activities and its metabolic patterns and oxidative stress when repressed. The oxygen concentration required for aerobic metabolism may also cause the production

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of reactive oxygen species (ROS) (Pelster *et al.*, 2016). The accumulation of ROS may result in oxidative stress, which is harmful for organisms (Lushchak, 2014). ROS concentrations vary with changes in oxygen availability, and organisms have developed sophisticated defense systems to eliminate reactive oxygen molecules when exposed to hypoxic and hyperoxic conditions (Welker *et al.*, 2013). Hypoxia is one of the most common conditions that can cause oxidative stress (Welker *et al.*, 2013). Catalase (CAT) and superoxide dismutase (SOD) play an important role in removing ROS. In addition, previous studies have clearly shown that the defense system against ROS is not static but is highly responsive to changing environmental conditions (Lushchak and Bagnyukova, 2006; Welker *et al.*, 2013). For example, a significant increase in ROS degrading enzyme activities and in the concentration of antioxidants is frequently observed following periods of hypoxia.

However, the response of air-breathing fish to oxidative stress under different environmental stresses has not been reported in the literature. Therefore, we studied the changes of antioxidative capacity under water and respiratory gas respiration inhibition in separate treatments to determine the effect of environmental stress on oxidative stress in air-breathing fish. Basic adaptive evolutionary studies were consulted to provide basic information.

## MATERIALS AND METHODS

### Experimental fish and holding conditions

Loach (*Paramisgurnus dabryanus*) specimens used in the experiment were collected from a local market in Dianjian, Chongqing, China (32°20'N; 107°21'E). The body weight was  $15.2 \pm 2.3$  g and fork length was  $13.3 \pm 1.2$  cm. The experimental specimens were kept in a re-circulating water tank (180 × 120 × 80 cm) in the laboratory for two weeks. In the tanks the fish had free access to air and could therefore breathe air voluntarily. During this period, the temperature of the de-chlorinated freshwater used was maintained at  $21.3 \pm 1.7^\circ\text{C}$  the oxygen level was kept above  $7.0 \text{ mg L}^{-1}$ , the pH ranged from 6.7 to 7.5. The photoperiod was maintained at 12 h light: 12 h dark and fish were fed with a commercial fish food (Xiwang Group, Chongqing, China).

### Experimental protocol

At the end of domestication, 150 healthy fish of a similar size were selected and randomly divided into three groups (Fig. 1): (1) inhibited group (n = 50), in which the *P. dabryanus* were kept active below the water surface for intestinal respiration; (2) air-exposed group (n = 50),

in which *P. dabryanus* were exposed to the air, although their bodies were kept; and (3) a control group, without any inhibition (n = 50). The details of the experimental protocol are shown in Figure 2.

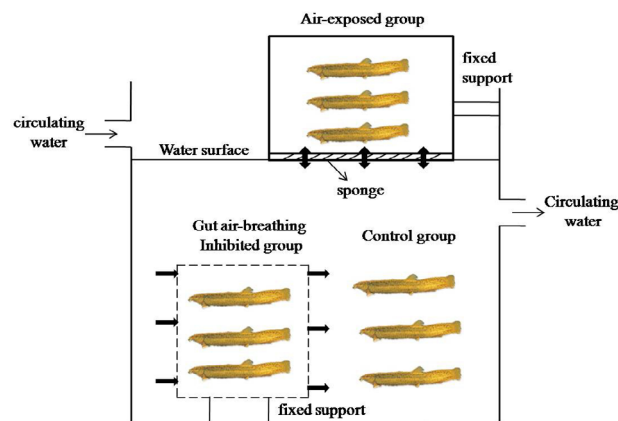


Fig. 1. Experimental device of *P. dabryanus*.

Acclimation			
1st day	Normoxia	Normoxia (Gut air-breathing restricted)	Atmospheric exposure (Gill breathing restricted)
7th day			
Recovery 0 day			
Recovery 1 day	Normoxia	Normoxia	Normoxia
Recovery 3 days	Normoxia	Normoxia	Normoxia

Fig. 2. Illustration of the experimental protocol.

### Sample collection

Six fish from each group were randomly selected for the determination of antioxidant enzyme activity. Body weight and fork length were measured after anesthesia with 0.1 g / L ethyl m-aminobenzoate. The fish were placed on an ice tray and the gills, foregut, midgut, and hindgut were quickly removed into liquid nitrogen. The organs were then placed in a cryogenic refrigerator at  $-80^\circ\text{C}$  for the determination of further enzyme activity.

### Determination of enzyme activity

The samples were removed from the refrigerator and placed on ice to thaw. After thawing was completed, each sample was weighed and homogenized. The homogenate was diluted with a F6 / 10 FLUKO hand-held

**Table I. Comparison of basic morphological parameters among different group (n=9).**

	Control group	Inhibited group	Air-exposed group
Weight (g)	15.90 ± 4.29	16.02 ± 4.10	16.19 ± 3.24
Fork length (cm)	13.04 ± 0.82	12.35 ± 1.20	13.76 ± 1.05
Gut mass (g)	0.27 ± 0.08	0.21 ± 0.05	0.25 ± 0.06
Gut length (cm)	6.36 ± 0.84	5.47 ± 0.62	6.48 ± 0.37
Gut length/fork length	0.49 ± 0.05	0.45 ± 0.06	0.47 ± 0.05
Gut mass/body mass	0.017 ± 0.01	0.013 ± 0.003	0.016 ± 0.004
Gill mass (g)	0.25 ± 0.02	0.19 ± 0.03	0.20 ± 0.02
Gill mass/body mass	0.016 ± 0.004	0.014 ± 0.005	0.013 ± 0.003

homogenizer for 5 min in an ice bath to ensure that the sample was fully ground. The volume ratio of the sample mass to the volume of the homogenate diluent was set to 1:9. The resulting homogenate was centrifuged using a cryogenic ultracentrifuge and the supernatant was used for the determination of antioxidant enzyme activity. CAT (EC1.11.1.6) and SOD (EC1.15.1.1) activity were measured in gill and gut tissue using a CAT assay kit (NO: A007, Nanjing Jiancheng Bioengineering Institute, PR China) and SOD assay kit (NO: A001, Nanjing Jiancheng Bioengineering Institute). All assays were performed on duplicate samples using a Thermo Multiskan spectrophotometer (Thermo Fisher Scientific, MA, USA). UV-permeable Corning 96-well microplates were used for all assays. All reactions were run at the saturating substrate concentrations determined for each enzyme (Li *et al.*, 2013).

#### Statistical analysis

IBM SPSS 19.0 was used for all statistical analysis of the recorded data. The effect of acclimation group (inhibited, air-exposed, control) and days on CAT and SOD activity were determined by a two-way analysis of variance (ANOVA). The effect of acclimation group (inhibited, air-exposed, control) on morphological parameters were determined by a one-way ANOVA followed by Dunnett's test. Any difference at  $P < 0.05$  was regarded as statistically significant. All values were expressed as mean ± SD.

## RESULTS

#### Morphological changes

After domestication for 7 d, there were no significant differences in the morphological parameters of *P. dabryanus* between groups (Table I).

#### Gill antioxidant activity

After domestication for 7 d, the gill CAT activity in the air-exposed group was significantly lower than that

in the control and inhibited group ( $P < 0.05$ ), while there was no significant difference in CAT activity between the control and inhibited group (Fig. 3a). After recovery for 1 d, the CAT activity in *P. dabryanus* was significantly increased in the air-exposed group ( $P < 0.05$ ), which was higher than that in the control and inhibited groups ( $P < 0.05$ ) (Fig. 3a). After recovery for 3 d, the CAT activity in *P. dabryanus* was significantly decreased ( $P < 0.05$ ), and there was no significant difference in CAT activity between the two groups (Fig. 3a). The gill SOD activity in the air-exposed group was much lower than that in the control and inhibited group ( $P < 0.05$ ), while the SOD activity in *P. dabryanus* in the control and inhibited group were not significant different (Fig. 3b). After recovery for 1 d, the SOD activity in *P. dabryanus* increased significantly in the air-exposed group ( $P < 0.05$ ), and after recovery for 3 d, there was no significant difference in SOD activity between the groups (Fig. 3b).

#### Foregut antioxidant activity

After domestication for 7 d, the foregut CAT activity in *P. dabryanus* in the air-exposed group was much lower than that in the control and inhibited groups ( $P < 0.05$ ) (Fig. 4a). The CAT activity in the foregut of *P. dabryanus* was significantly increased ( $P < 0.05$ ) after recovery for 1 d, and there was no significant difference in CAT activity between the groups (Fig. 4a). The SOD activity in *P. dabryanus* in the air-exposed group was lower than that in the control and inhibited groups ( $P < 0.05$ ) (Fig. 4b). After a 1 d recuperation, the SOD activity in *P. dabryanus* was significantly increased ( $P < 0.05$ ) in the air-exposed group and there was no significant difference in CAT activity between groups (Fig. 4b).

#### Midgut antioxidant activity

CAT activity in the midgut of *P. dabryanus* specimens in the air-exposed group was lower than that of the control group after domestication for 7 d ( $P < 0.05$ ) (Fig. 5a). After recovery for 1 d, the CAT activity in the midgut of

*P. dabryanus* sharply increased in the air-exposed group, and was much higher than that in the control and inhibited groups ( $P < 0.05$ ) (Fig. 5a). The midgut SOD activity in *P. dabryanus* in the air-exposed group was significantly lower than that in the control and inhibited groups ( $P < 0.05$ ) (Fig. 5b). After a 1 d recovery, SOD activity was significantly increased ( $P < 0.05$ ) in the midgut of *P. dabryanus* specimens that were exposed to air (Fig. 5b). After recovery for 3 d, the SOD activity in *P. dabryanus* decreased significantly in the air-exposed group ( $P < 0.05$ ). There was no significant difference in the CAT activity between groups (Fig. 5b).

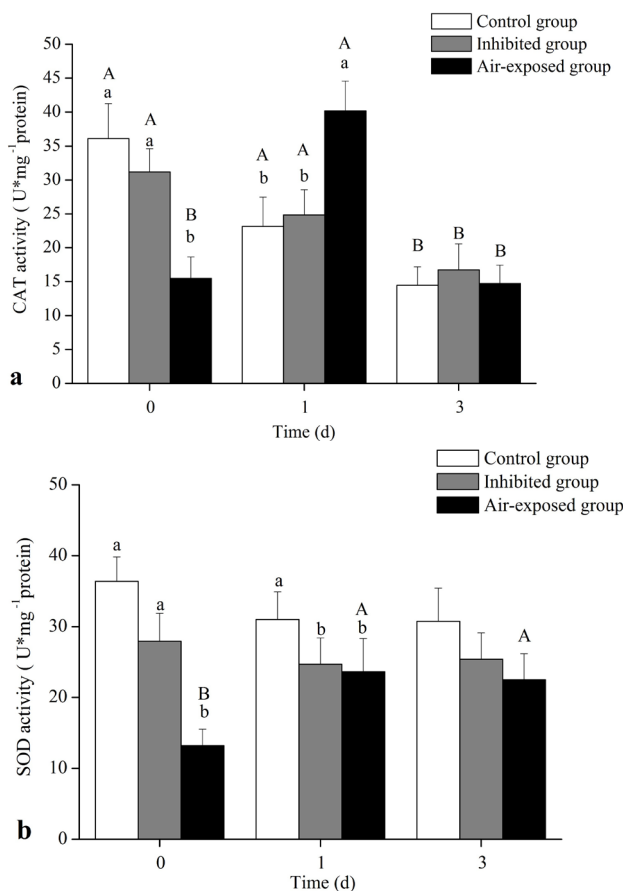


Fig. 3. Change of antioxidant capability in gill of loach. (a) catalase; (b) superoxide dismutase. Values in the table are represented as Mean  $\pm$  SD; n=6; different superscripts (lowercase letters) indicate significant differences in different group; different superscripts (capital letters) indicate significant differences in different day;  $P < 0.05$

#### Hindgut antioxidant activity

After domestication for 7 d, the CAT activity in the hindgut of *P. dabryanus* in the inhibited group was higher than that in the control and air-exposed groups ( $P < 0.05$ ),

and there was no significant difference in CAT activity between groups after recovery for 1 d (Fig. 6a). The SOD activity in *P. dabryanus* in the air-exposed group was lower than that in the control and inhibited groups ( $P < 0.05$ ) (Fig. 6b). SOD activity in *P. dabryanus* in the air exposed group increased rapidly after a 3d recovery ( $P < 0.05$ ), and there was no significant difference in the SOD activity between groups (Fig. 6b).

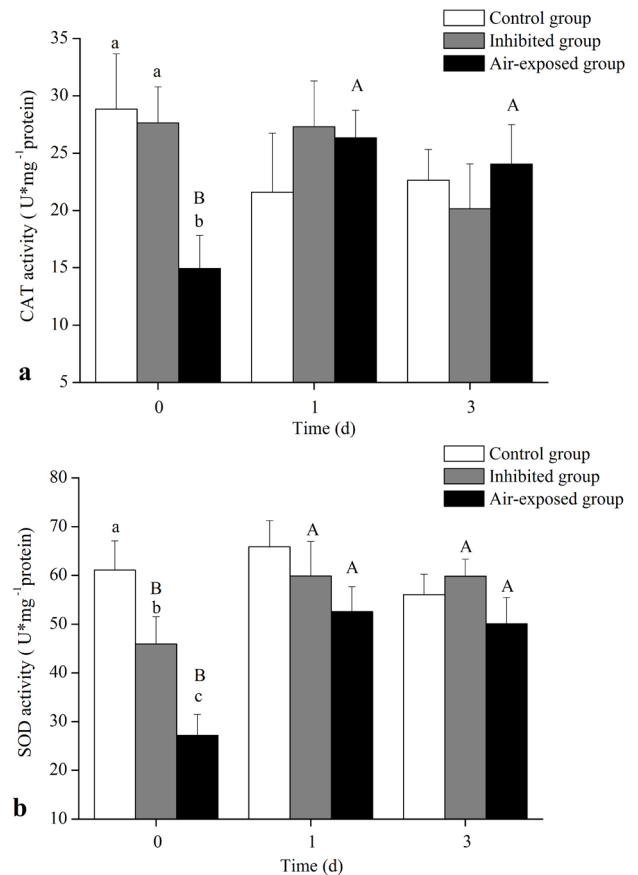


Fig. 4. Change of antioxidant capability in anterior intestine of loach. (a) catalase; (b) superoxide dismutase. Values in the table are represented as Mean  $\pm$  SD; n=6; different superscripts (lowercase letters) indicate significant differences in different group; different superscripts (capital letters) indicate significant differences in different day,  $P < 0.05$

## DISCUSSION

ROS production commonly accounts for 0.1-0.2% of the daily oxygen consumption in fish (Gorr *et al.*, 2010). Large amounts of ROS can damage tissue structures (Costantini *et al.*, 2010). Generally, the CAT and SOD systems are regarded as important cellular defense systems against oxidative stress.

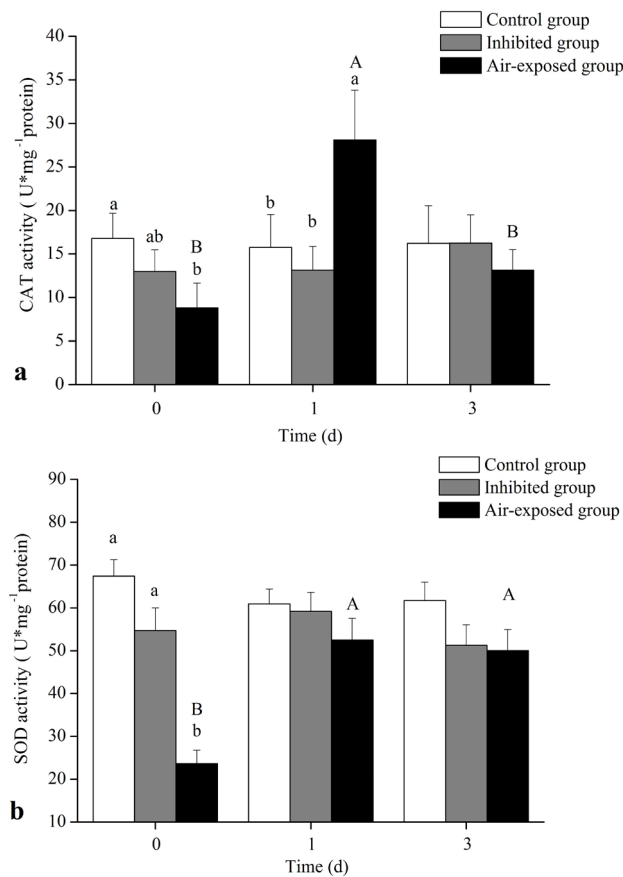


Fig. 5. Change of antioxidant capability in middle intestine of loach. (a) catalase; (b) superoxide dismutase. Values in the table are represented as Mean  $\pm$  SD; n=6; different superscripts (lowercase letters) indicate significant differences in different group; different superscripts (capital letters) indicate significant differences in different day;  $P < 0.05$

We found that the air-breathing loach *P. dabryanus* can adjust the levels of oxidative stress it experiences when confronted with different environmental conditions. In recent studies, CAT and SOD activity in the gills, foregut, and midgut of *P. dabryanus* has been shown to decrease significantly when the fish are exposed to air. In contrast, in jeju (*Hoplerythrinus unitaeniatus*), the exposure of fish tissues to air may stimulate ROS production (Pelster *et al.*, 2016). Under normal physiological conditions, organisms continually eliminate ROS and thus avoid damage to the body caused by oxidation. The mechanism of its elimination depends mainly on the body's antioxidant defense system, with CAT and SOD having important roles. We found that the CAT and SOD activity in the hindgut of *P. dabryanus* increased in the inhibited group. Studies have shown that when fish face stress conditions,

ROS are produced more than they are scavenged (Hur *et al.*, 2001). The ROS content accumulates, which results in oxidative stress, leading to the body reaching a pathological state. Demple (1999) showed that in a body under chronic stress conditions the activity of antioxidant enzymes can increase to reduce oxidative damage.

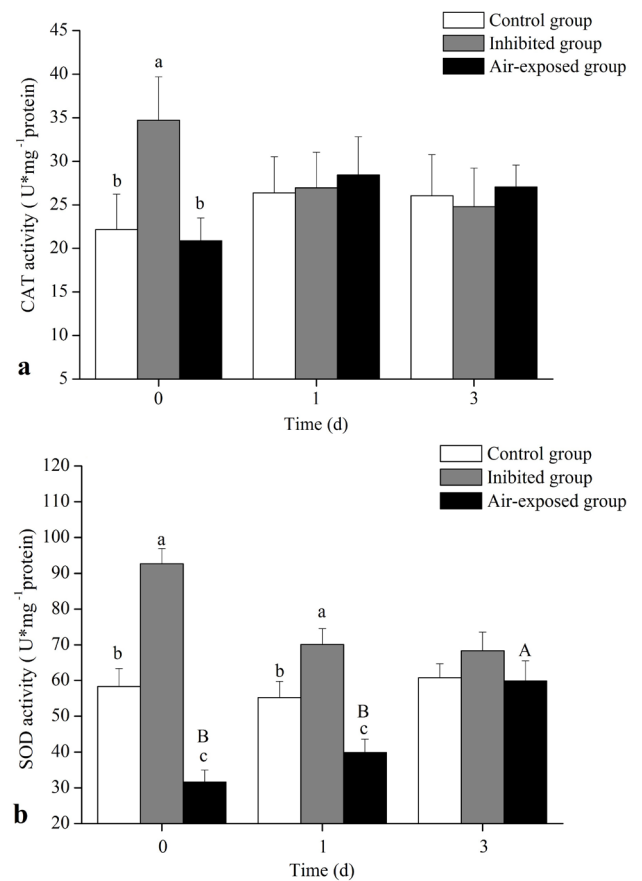


Fig. 6. Change of antioxidant capability in posterior intestine of loach. (a) catalase; (b) superoxide dismutase. Values in the table are represented as Mean  $\pm$  SD; n=6; different superscripts (lowercase letters) indicate significant differences in different group; different superscripts (capital letters) indicate significant differences in different day;  $P < 0.05$

When the fish were placed in normoxic conditions for 1 d, the CAT and SOD activities were significantly increased, which may be due to the feedback regulation of dissolved oxygen in the environment. Some studies of the antioxidant system have observed an increased activity in antioxidant enzymes under hypoxic stress or following a return to normoxic levels (Costantini *et al.*, 2010; Majmudar *et al.*, 2010). Other studies have shown that fish are accompanied by significant oxidative stress when

they recover from hypoxia, and some extremely hypoxia-tolerant animals are more susceptible to oxidative stress during recovery than under hypoxia (Bickler and Buck, 2007).

With the re-introduction of oxygen, ROS may rapidly accumulate in tissues or cells. Excess ROS can cause oxidative stress and disorders of the antioxidant system (Lushchak *et al.*, 2005). The results of this study are consistent with those of de Oliveira *et al.* (2005), who found that the antioxidant capacity of the body during reoxygenation may change significantly under hypoxia or stress, which may be an antioxidant strategy that organisms use to adapt to hypoxia and reoxygenation. After a 3 d resumption of normoxia, there was no significant difference in the antioxidant enzyme activity between groups. Under different environmental conditions, the fluctuation of the oxygen utilization rate in the matrix itself has an important effect on the ROS content (Welker *et al.*, 2013). Pannunzio (1998) found that with the prolongation of hypoxia stress, the amount of oxygen in the body decreased and the SOD was reduced to  $O_2^-$  reducing its activity in the body. When restored to normoxia for 1 d, the CAT in the air exposed group was significantly increased and was higher than that in the control group. The aerobic capacity of the hindgut in the air exposure group and the inhibition group was similar to that of the control group. Different tissues have different adaptations to overcome environmental stress. These reactions are all distinctive anti-oxidant strategies that prevent the body's ROS from damaging itself.

## CONCLUSIONS

Based on the above data, when *P. dabryanus* was exposed to the hypoxic and inhibited treatments, they had a high viability under the air-exposed condition (in anaerobic conditions), while the level of oxidative stress in the gill, foregut, and midgut clearly decreased. When exposed to normoxia conditions, *P. dabryanus* recovered quickly. The aquatic air-breathing fish *P. dabryanus* was able to regulate its physiological responses to the environmental disturbances provided in this study.

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## Statement of conflict of interest

Authors have declared no conflict of interest.

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