# Cardioprotective Effect of Naringenin against Myocardial Ischemia-Reperfusion Injury via Alteration of Apoptotic Signaling Pathway

SINO IBEA



Tao Ren¹, Guiqiu Cao², Xiao Han³, Feng Tan¹, Qiaoli Chen⁴, Shicheng Yang⁵ and Haiyan Zhang⁶\*

<sup>1</sup>Department of Third Cardiology, First Affiliated Hospital, School of Medicine, Shihezi University, Shihezi, Xinjiang, 832008, China

<sup>2</sup>Department of Cardiology, Fifth Affiliated Hospital of Xinjiang Medical University, Urumqi, Xinjiang, 830011, China

<sup>3</sup>Department of Cardiology, General Hospital of Fuming of Liaoning Health Industry Group, Fushun, Liaoning, 113000, China

<sup>4</sup>Department of Cardiology, First People's Hospital of Wuyi County, Jinhua, Zhejiang, 321200, China

<sup>5</sup>Department of Cardiology, Tianjin Chest Hospital, Tianjin, 300222, China <sup>6</sup>Department of Cardiology, The Second Affiliated Hospital of Nanjing Medical University, Nanjing, Jiangsu, 210011, China

Tao Ren and Guiqiu Cao are co-first authors and contributed equally to this work.

### ABSTRACT

Ischemic heart disease (IHD) is a common multiple cardiovascular disease (CVD) in clinical settings. It is a major contributor to mortality and morbidity worldwide and causes a serious threat to human life and health. Naringenin, a flavonoid possesses the potent antioxidant potential and it is proposed to be useful in the treatment of CVD. In this experimental study, we aimed to scrutinize the cardio-protective effect of naringenin against the I/R induced myocardial injury and elucidate the possible mechanism of action. In vitro studies, the H9c2 cardiomyocytes cells were treated with naringenin or without naringenin and then subjected to I/R, respectively. At end of the experimental study, the rats were anesthetized and blood samples were collected to scrutinize the various parameters such as creatine kinase myocardial band (CK-MB), high-density lipoprotein (HDL), lactate dehydrogenase (LDH), creatine, troponin-T (TRT), cholesterol, C - reactive protein (CRP), and concentration of mitochondrial enzymes viz., Ca2+, Na+ and K+ ions were estimated in blood. Heart tissue was also isolated for caspase-3 activity. Our result showed that naringenin pretreatment significantly increased cardiac dysfunction via scavenged free radicals and a reduction of inflammatory reaction. Dose-dependent treatment of naringenin significantly altered the CK-MB, HDL, LDL, creatinine, cholesterol, CRP, Ca<sup>2+</sup>, Na<sup>+</sup> and K<sup>+</sup>, respectively. A significant alleviating change in these biochemical parameters along with caspase-3 activity was noticed. Thus, in our study, we determined that I/R induced cardiac remodeling can be successfully mitigated by naringenin.

Article Information
Received 04 January 2020
Revised 23 February 2020
Accepted 04 March 2020
Available online 01 December 2021

Published 13 June 2022

**Authors' Contribution** 

TR performed the experimental study. GC and HZ estimated the biochemical data. All the authors wrote and proof-read the manuscript.

Key words Myocardial ischemia, I/R injury, Naringenin, Anti-oxidant, Antiinflammatory

# **INTRODUCTION**

Myocardial ischemic heart disease is a common cardiovascular disease (CVD) and it is having a higher incidence of mortality and morbidity worldwide. It is well documented that acute myocardial ischemia having

\* Corresponding author: ysxm53460@sina.com 0030-9923/2022/0005-2285 \$ 9.00/0



Copyright 2022 by the authors. Licensee Zoological Society of

This article is an open access  $\center{3}$  article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https://creativecommons.org/licenses/by/4.0/).

a higher rate of death. Due to sudden death, myocardial ischemia heart disease leads the numerous clinical dysfunctions including cardiac myocyte hypertrophy (Seki and Fishbein, 2014; Müller-Nordhorn and Willich, 2016). Myocardial ischemia-reperfusion is definite as tissue damage that arises when early and fast coronary flow returns to the heart tissue after the ischemia, which commonly augments the myocardial damage. During the myocardial ischemia, the organ showed a lack of nutrients and oxygen, which cause oxidative stress, apoptotic cell death and inflammatory reaction, and these circumstances will additionally deteriorate during the reperfusion (Hausenloy and Yellon, 2013; Hashmi and Al-Salam, 2015). During the I/R, it induces the structural injury and dysfunction or

metabolic disturbance. These alterations are induced via the retrieval of blood flow to the ischemic myocardium. I/R induced ischemic myocardium is very complicated, and inflammation, ERS and oxidative stress are the main targets for the current research. Previous research suggests that the excessive generation of ROS was observed during the myocardial ischemic, which further induce oxidative stress and cause harmful changes (Kalogeris et al., 2012; Raedschelders et al., 2012). Various researches have been explored to scrutinize the cardioprotective drug. It is well known that myocardial ischemia-reperfusion is an effectual treatment for acute myocardial ischemia, which can decrease the size of myocardial infarction and release the symptoms of heart disease such as heart failure (Sanada et al., 2011; Hausenloy and Yellon, 2013). Various signaling pathway dysregulationhas been induced the myocardial ischemia-reperfusion. Signaling dysregulation leading the differentiation, cell proliferation, apoptosis and autophagy under various pathological and physiological conditions may be closely related to cardiac injury (Burke and Virmani, 2007; Kalogeris et al., 2012; Ibáñez et al., 2015).

It is well proofed that flavonoids are the major phytoconstituents in various Chinese medicine and used to treat various disease especially CVD (Zeng *et al.*, 2017). Epidemiological investigations have demonstrated that a regular intake of flavonoids can decrease the risk of myocardial infarction and decrease mortality due to CVD (Zeng *et al.*, 2017; Zhu *et al.*, 2018).

Naringenin, a well-known flavonoid, that can exert a wide range of pharmacological effects including anticancer, anti-inflammatory, anti-diabetic, anti-viral, free radical scavenging effect, anti-tubercular and other biological activities (Smeriglio et al., 2017). Due to its anti-oxidant and anti-inflammatory nature, it contributes to the cardio-protective effect. Previous studies suggest that the naringenin not only reduce the peroxidase activity of cytochrome C with lipid peroxidative or free radical generation and dioleylcardiolipin, but also circulated the oxidative enzymes activities and over-generation of ROS for enhancing the I/R injury (Agouni et al., 2011; Smeriglio et al., 2017). Moreover, the exact mechanism of action is not fully elucidated. Consequently, in the current experimental study, we aimed to scrutinize the protective effect of naringenin on I/R injury and explore the possible mechanism of action.

# MATERIALS AND METHODS

Chemicals

Naringenin(95%) was purchased from the Sigma Aldrich, U.S.A. Radio-immunoprecipitation assay (RIPA) lysis buffer (Beyotime Institute of Biotechnology, Beijing,

China), 2,3,5-Triphenyl tetrazolium chloride (TTC) was purchased from the Sigma, Beijing, China. Catalase (CAT), superoxide dismutase (SOD), Malondialdehyde (MDA), glutathione (GSH), serum creatinine kinase MB (CK-MB) assay kits were purchased from the Jiancheng Bioengineering Institute, Nanjing, China. The primary antibodies against the Akt, phosphorylation of Akt (p-Akt), Bax, Bcl-2, cleaved caspase-3, caspase-3, caspase-7, caspase-9, PI3K, phosphorylation of PI3K (p-PI3K), XIAP, HrtA2/Omi, β-actin and GAPDH were purchased from the Cell Signaling Technology, Inc (CST, USA).

In vitro *experimental study* 

Cell culture

H9c2 (clonal cell lines of BDIX rat embryonic heart tissue) cells were procured from the Cell Bank of Chinese Academy of Sciences, Shanghai, China. The cell was cultured according to using the previously reported method with minor modification. Briefly, the cells H9c2 were cultured into the high glucose DMEM supplemented with L-glutamine (2 mM), streptomycin/penicillin (1% v/v) and fetal bovine serum (10% v/v) and maintained into the humidified incubator in a CO<sub>2</sub> (5%) atmosphere at 37°C. The cells were plated at a suitable density and propagate for 24-36 h before the experimental study (Shu et al., 2019).

The H9c2 cells were divided into the following groups presented in Figure 1. Briefly, high glucose DMEM medium replaced the glucose-free DMEM to ischemia and further cells were incubated for 6h at 37°C in the environment of  $H_2$  (5%),  $N_2$  (5%) and  $CO_2$  (5%) in an anaerobic glove box. After that, the medium replaced with the high glucose medium and kept for 16h to mimic reperfusion. In the treatment group, different doses of naringenin (1.25, 2.5, 5, 10, 20, 40  $\mu$ M) for 12h before the H/R. Nevertheless, different doses of naringenin pretreated group, pretreated with the H9c2 cells for 12h (Shu et al., 2019).

Cell viability assay

MTT assay was used for the estimation of cell viability via using the previously published method with minor modification (Shu *et al.*, 2019). The H9c2 cells were plated in the 96 well plates at a density of  $5 \times 10^4$  cells per well with DMEM (100  $\mu$ L) medium. After that MTT solution (20  $\mu$ L) was mixed in each well and again incubated at 37°C for 4h. The MTT medium was replaced with the DMSO (150 mL each well) and added the formazan crystals and finally estimated the absorbance at 570 nm on a microplate reader (Shu *et al.*, 2019).

Antioxidant parameters

ROS detection kit was used for the estimation of

intracellular ROS production via using the manufacture instruction (Shu et al., 2019). Briefly, H9c2 cells were cultured into the 6 well plates for 24-36h. After that, the cells were propagated, washed with washing buffer and finally centrifuged at 1g rpm for 5 min at room temperature and finally supernatant was discarded. After that, the cells were re-suspended and incubated with 5-(and-6)-carboxy-20,70-dichlorodihydrofluorescein diacetate (25 µM) at 37°C for 30 min in a dark room and washed with the PBS buffer. After the various treatments, the supernatant was utilized for the estimation of LDH level and then the cells were collected and centrifuged for 5 min at 1 g rpm at 4°C. Further, the supernatant was used for the estimation of SOD, MDA, GSH-Px and CAT level via using the manufacture instruction (Nanjing Jiancheng Bioengineering Institute, Nanjing, China).

In vivo *study* 

# Experimental animal

Sprague-Dawley (SD) rats (7-9 weeks; 250-300 g) were used for the current experimental study. The rats were received from the departmental animal house and kept in the standard experimental condition (22±5°C; 70±2 relative humidity 12/12 h dark/light cycle). The rats have received the standard pellet diet with water *ad libitum*. The current experimental study was approved from the Institutional animal ethical.

### Experimental protocol

The rats were divided into 5 groups and each group contains the 10 rats. The group as follows: Group I, sham control received the normal saline (10 mL/kg); Group II, I/R control; Group III, I/R control + naringenin (2.5 mg/kg); Group IV, I/R control + naringenin (5 mg/kg); Group V, I/R control + naringenin (10 mg/kg).

# Surgical protocol

For the surgical protocol, all the experimental rats were subjected to endotracheal and tracheotomy intubation after the anesthesia. For the ventilation performed, inserting the polyethylene-50 (PE-50) tube via trachea and link with the animal ventilator with a tidal volume (1.2 L/kg) and maintain the breath rate (70/min) and maintain the body temperature 37°C via using the heating pad. MP150 data acquisition was used for the determination of heart rate and ECG. All these parameters were monitored to the baseline, Ischemia (ischemia for 45 min) and reperfusion (reperfusion for 4 h) (Yin *et al.*, 2013).

# Myocardial infarction assessment

For the myocardial infarction, Evans Blue dye (2%) was injected into the aorta on experimental rats and all the rats were euthanized via cervical dislocation. After the euthanized, the heart tissues were immediately removed,

washed with saline and frozen and finally cut into the approximately 5 cross-sections from the apex junction site. TTC method was used to identify the myocardial infarct area into the left ventricle (Hu *et al.*, 2016).

### Biochemical parameter

For the estimation of the biochemical parameter, the heart tissues were homogenized with phosphate buffer (pH=7.4) and finally centrifuged for 15 min at 5g rpm. Collect the supernatant for the estimation of the reactive oxygen species (ROS) level. The serum parameters were scrutinized for aspartate aminotransferase (AST), creatine kinase-MB (CK-MB) and lactic acid dehydrogenase (LDH) via using the manufacture instruction's (NJJC Bio, Nanjing, China).

### Antioxidant parameter

Antioxidant parameters such as malondialdehyde (MDA), superoxide dismutase (SOD) and catalase (CAT) were estimated via using the standard kits following the manufacture instruction (NJJC Bio, Nanjing, China).

# Lipid parameter

Lipid parameters such as triglycerides total cholesterol (TC), low-density lipoprotein (LDL) and high-density lipoprotein (HDL) were estimated via using the standard kits following the manufacture instruction (NJJC Bio, Nanjing, China).

 $Na^+$ ,  $K^+$  and  $Ca^{2+}$ 

Na, K and Ca were estimated via using the previous method with minor modification (Abdalla *et al.*, 2016).

## Membrane-bound ATPases

Membrane-bound enzymes such as Ca<sup>++</sup>ATPase, Mg<sup>++</sup>ATPase and Na<sup>+</sup>K<sup>+</sup>ATPase were estimated via using the previously reported method with minor modification (Gandhi *et al.*, 2013).

## Caspase-3 activity

Caspase-3 activity and C - reactive protein (CRP) were determined by using the standard kits following the manufacture instruction (NJJC Bio, Nanjing, China).

# Statistical analysis

For the current experimental study, the whole result presented as mean  $\pm$  standard error (SD). One-way analysis of variance (ANOVA) was used for scrutinizing the statistical analysis, followed via Turkey's test method using the Graph Pad Prism 7 software. Values were considered statistically significant when P <0.05.

# **RESULTS**

Effect of naringenin against H/R induced H9c2 cell injury

MTT assay was performed for the estimation of the potential effect of naringenin on cell viability (Fig. 1). In the current study, we firstly investigated the cell proliferation or general toxicity of naringenin. In the current experimental study, the cells were treated with the naringenin in a dose-dependent manner. Figure 1A exhibited that the cell viability had no significant difference was observed between groups. After that, the capability of naringenin to reduce the H/R induced H9c2 cell injury was scrutinized. Figure 1B exhibited the considerably down-regulation of cell viability and naringenin treatment significantly maintained the cell viability at approximately (85%) and a similar trend was observed in the LDH release (Fig. 2). Moreover, in the current experimental study, we used the 10 µM naringenin for subsequent in-vitro experimental study.

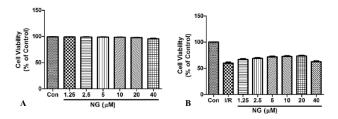


Fig. 1. Effect of naringenin on the cell viability by H/R in H9c2 cardiomyocytes. A) H9c2 cells were treated with various concentration of naringenin for 24 h and MTT assay was used for the estimation of cell viability. B) H9c2 cells were treated with for 12 h with naringenin and then exposed to 6h of hypoxia. The data presented as means±SD from three independent experiments.

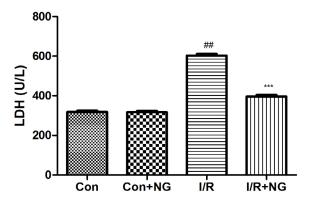


Fig. 2. Effect of naringenin on the LDH release by H/R in H9c2 cardiomyocytes. The data presented as means $\pm$ SD from three independent experiments. ##P < 0.01 versus control group; \*P < 0.05, \*\*P < 0.01 versus H/R group.

#### Antioxidant activity (in vitro)

It is well documented that oxidative stress plays an important role in the expansion of the cardiac disease.

Oxidative stress initiates apoptosis or injury. Due to this point, we have investigated the antioxidant effect of naringenin against the H/R induced oxidative damage during the cardiac disease. H/R induced oxidative stress in the H9c2 cells exhibited the reduce level of CAT (Fig. 3A), GSH-Px (Fig. 3B), SOD (Fig. 3C) and increase the level of MDA (Fig. 3D) and naringenin treatment significantly ameliorated these effects. H/R injured H9c2 cells showed the deposition of ROS and pre-treatment of naringenin significantly reduced the accumulation of ROS (Fig. 4). The result exhibited that the naringenin could be more effectively alleviate the H/R induced oxidative stress injury in H9c2 cells.

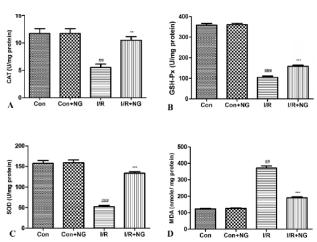


Fig. 3. Effect of naringenin on antioxidant parameter of H/R in H9c2 cardiomyocytes cells. A) CAT, B) GSH-Px, C) SOD and D) MDA. The data presented as means $\pm$ SD from three independent experiments. ##P < 0.01 versus control group; \*P < 0.05, \*\*P < 0.01 versus H/R group.

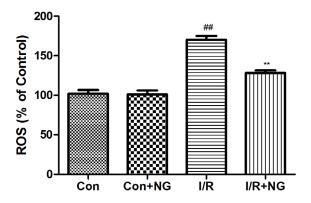


Fig. 4. Effect of naringenin on ROS concentration of H/R in H9c2 cardiomyocytes cells. The data presented as means $\pm$ SD from three independent experiments. ##P < 0.01 versus control group; \*P < 0.05, \*\*P < 0.01 versus H/R group.

Naringenin avert I/R induced myocardial injury

The myocardial enzymes such as CK-MB, LDH and AST were significantly boosted during the I/R induced myocardial injury. I/R induced myocardial damage rodent exhibited the increased level of CK-MB (Fig. 5A), LDH (Fig. 5B) and AST (Fig. 5C) and dose-dependently treatment of naringenin significantly (P<0.001) ameliorated these parameters.

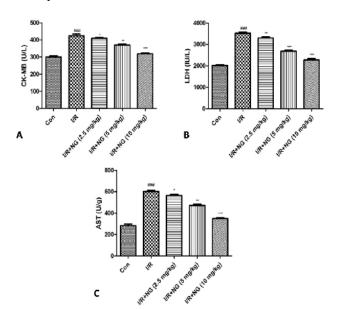


Fig. 5. Effect of naringenin on the biochemical parameter of I/R induced mice. A) CK-MB, B) LDH and C) AST. Dennett's test was used for statistically significance \*P<0.05, \*\*P<0.01 and \*\*\*P<0.001.

Naringenin avert oxidative damage during the I/R induced myocardial injury

Figure 6 exhibited the effect of naringenin on the antioxidant enzymes. I/R induced myocardial damage rats exhibited the reduce level of MDA (Fig. 6B) and increase the level of SOD (Fig. 6A), CAT (Fig. 6C) and dose-dependently treatment of naringenin significantly (P<0.001) altered the level of antioxidant enzymes.

### Naringenin altered lipid parameters

Figure 7 exhibited the effect of naringenin on the lipid parameters of I/R induced myocardial injury. I/R induced myocardial injury rats demonstrated the increased level of total cholesterol, high-density lipoprotein and low-density lipoprotein as compared to normal control. Dose dependently treatment of naringenin significantly (P<0.001) reduced the level of TC, HDL and LDL as compared to the I/R induced myocardial injury rats.

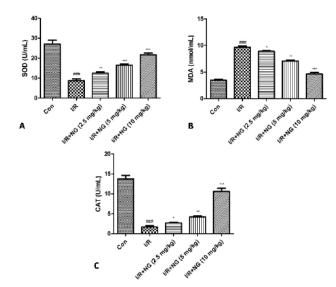


Fig. 6. Effect of naringenin on the antioxidant parameters of I/R induced mice. A) SOD, B) CAT and C) MDA. Dennett's test was used for statistically significance \*P<0.05, \*\*P<0.01 and \*\*\*P<0.001.

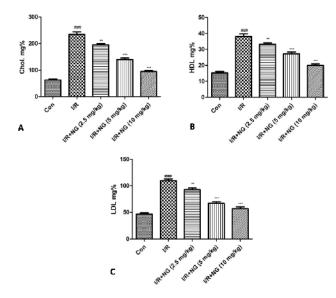


Fig. 7. Effect of naringenin on the lipid parameters of I/R induced mice. A) Cholesterol, B) High density lipoprotein and C) Low density lipoprotein. Dennett's test was used for statistically significance \*P<0.05, \*\*P<0.01 and \*\*\*P<0.001.

Effect of naringenin on Na, Ca and K level

During the myocardial cardiopathy increase the level of Na, Ca and K in the serum and similar results were obtained in the I/R induced myocardial injury rats. I/R induced group rats showed the increased level of Na, Ca and K and naringenin significantly (P<0.001) reduced the

level almost near to normal control group (Fig. 8).

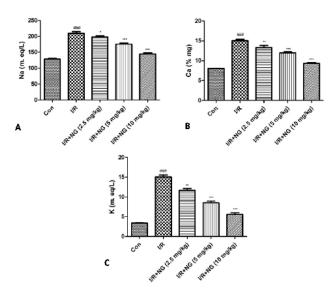


Fig. 8. Effect of naringenin on the Na, Ca and K level of I/R induced mice. A) Na, B) Ca and C) K. Dennett's test was used for statistically significance \*P<0.05, \*\*P<0.01 and \*\*\*P<0.001.

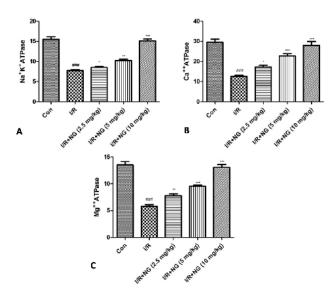


Fig. 9. Effect of naringenin on the membrane-bound enzymes of I/R induced mice. A) Mg++ ATPase, B) Na+K+ ATPase and C) Ca++ ATPase. Dennett's test was used for statistically significance \*P<0.05, \*\*P<0.01 and \*\*\*P<0.001.

# Membrane-bound enzymes

Mg<sup>++</sup> ATPase, Na<sup>+</sup>K<sup>+</sup> ATPase and Ca<sup>++</sup> ATPase all enzymes are considered as the membrane ATP dependent enzymes. I/R induced group rats exhibited the reduced

level of membrane-bound enzymes such as  $Mg^{++}$  ATPase,  $Na^+K^+$  ATPase and  $Ca^{++}$  ATPase and dose-dependently treatment of naringenin significantly (P<0.001) increased the level of membrane-bound enzymes such as  $Mg^{++}$  ATPase,  $Na^+K^+$  ATPase and  $Ca^{++}$  ATPase (Fig. 9).

Naringenin avert caspase-3 activity and CRP level

Figure 10 showed the increased level of caspase-3 activity in the I/R induced control group rats. Naringenin significantly (P<0.001) decreased the activity of caspase-3 in a dose-dependent manner.

A similar momentum was observed in the I/R induced myocardial injury rats and concentration-dependent treatment of naringenin significantly (P<0.001) downregulated the level of CRP (Fig. 11).

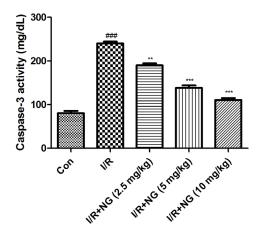


Fig. 10. Effect of naringenin on the caspase-3 marker of I/R induced mice. Dennett's test was used for statistically significance \*P<0.05, \*\*P<0.01 and \*\*\*P<0.001.

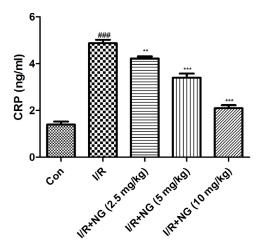


Fig. 11. Effect of naringenin on the CPR level of I/R induced mice. Dennett's test was used for statistically significance \*P<0.05, \*\*P<0.01 and \*\*\*P<0.001.

### **DISCUSSION**

Ischemic CVD is considered the most common CVD and it predominantly affected human health (Nordlie *et al.*, 2005). I/R is a clinical phenomenon that induces the structure injury and metabolic or dysfunction interruption. These alterations are induced via the recovery of blood flow to the ischemic myocardium (Darby *et al.*, 2013). The mechanism of I/R induced myocardial injury is not clear, but few of the research suggest that the oxidative stress, inflammatory and apoptosis play an important role during the disease.

It is well documented that flavonoids play a preventive role in the expansion of the CVD. Flavonoids such as naringenin, a well-known Phyto-constituents of various Chinese herbal medicine used in the inflammatory and oxidative stress-induced various diseases via potent antiapoptotic effects (Zeng et al., 2017). Naringenin is a major ingredient of flavonoids and it belongs to the flavanonol subclass. As per the report of the Chinese herbal medicine, various medicinal and edible fruits and vegetables have been reported to contain naringenin (Zygmunt et al., 2010). Previous research suggests showed the cardio-protective effect of naringenin against various animal models (Jiang et al., 2005; Zygmunt et al., 2010). Also, the essential mechanism of naringenin's myocardial protection has still not been fully explained. However, in the current experimental study, we explored the preventive effects of naringenin against the I/R induced myocardial injury and explore the possible mechanism of action.

Previous studies suggest that the reduced level of creatinine assuage the remodelled cardiac cells (Moghimian *et al.*, 2013; Anderson *et al.*, 2011). Analogously, in the current experimental study, the destruction of the myocardial cell was further controlled via reduce level of creatinine level brought by naringenin. Various researches suggest that the CRP is a significant marker for evaluating the cardiac remodelling and it is well documented that diabetes to be responsible for the cardiac remodeling (Slavíková *et al.*, 2007). In the current experimental study, we found that the naringenin prevents cardiac remodelling via down-regulation of the CRP level in a dose-dependent manner. Naringenin also reduced the level of CRP via inhibiting the vascular obstruction in the myocardial cell.

Numerous ions play a significant role in governing the physiology functional nature of cardiac cells (Wright *et al.*, 2001). Previous research suggests that basic ions such as Ca<sup>2+</sup>, Na<sup>+</sup> and K<sup>+</sup>directly or indirectly are related to contraction and conduction of assorted phenomenon in myocardiocytes (Shekhawat *et al.*, 2009). The alterations in the level of these ions to a significant level have a

direct implication on the cardiac structure (Pertoldi et al., 2006). Our results demonstrate there is a significant effect of our molecule on K+ ion. This molecule decreased the concentration of K+ ions in the serum when compared with diseased control. Suggesting its role on K<sup>+</sup> channels, the release of K+ion from myocardiocytes in the blood serum decreases the cationic concentration of cardiac cells (Wang et al., 2012). The decrease in cationic concentration relaxes the myocardial cell giving more relaxation time to cardiac cells and reducing the load of contraction. Another important parameter of evaluating cardiac remodelling is through analyzing the Na<sup>+</sup> and Ca<sup>2+</sup> concentrations which may also contribute to cardiac remodelling (Wang et al., 2009, 2012). There is no significant alteration in that Ca<sup>2+</sup> and Na<sup>+</sup> ions. The channels for these ions are spared by this compound.

It is well proofed that the I/R enhanced all types of blood pressure; mean systolic or diastolic along with heart rate in the pathogenic rodents as compared to normal rats, this trend of I/R induced myocardial injury is well correlated with our experimental study (Plosker and McTavish, 1995). Increase in mean BP and heart rate in diseased rats can be attributed to increase in release of catecholamines and renin, which in turn initiates the formation of Angiotensin II from Angiotensin I (Moghadam et al., 2013). CK-MB enzyme is a significant marker for identifying the changes in the remodeled heart, as its normally increased in during the heart disease. In the current experimental study, we observed the increased activity of CK-MB and dosedependent treatment of naringenin significantly increased the activity of CK-MB. The protective effect may be due to a reduction in myocardial contractility.

It is well documented that HDL is an important parameter of cardiomyopathy and it is directly responsible for the clearance of cholesterol via bile (Villani *et al.*, 1991). The clearance of cholesterol significantly decreases the load on cardiomyocytes. In the current experimental study, the level of HDL and cholesterol level considerably increased in I/R induced myocardial injury rats and dose-dependent treatment of naringenin significantly decreased the level of HDL and cholesterol (Villani *et al.*, 1991; Sarvari *et al.*, 2010). Based on the result, we can say that the HDL and flavonoids correlating.

Previous research suggests that oxidative stress plays an important role in the expansion of various diseases (Malorni et al., 1998). During the oxidative stress, observed the imbalance between the free radicals and ROS, which lead to injurious effects on the body. It is well proved that oxidative stress plays an important role in the expansion of I/R injury (Nader et al., 2010; Vishnupriya and Padma, 2017). Various research suggests that the ROS are the foremost factors that induce oxidative stress, which in turn

result in cell injury to biological macromolecules, influence the normal physiological state of cells and instigates the oxidative stress reaction. In the current investigation, we observed that the H/R and I/R significantly enhanced ROS levels, induced H9c2 dell damage and pathological changes, and enhanced MDA production (an indicator of LPO) and decreased the level of SOD and CAT (Tang et al., 2017; Yu et al., 2019). Dose-dependent treatment of naringenin significantly (P<0.001) decreased the level of MDA and increased the level of SOD and CAT in the H9c2 and cardiac muscle tissue. Our result revealed that the naringenin considerably reduced the oxidative stress in the H9c2 cells and the cardiac muscle and based on the result; we can conclude that the naringenin reduced the myocardial injury via inhibition of oxidative stress.

Apoptosis of cardiac myocardial injury induced via oxidative stress and inflammation plays an important role in the pathogenesis of CVD, such as ischemic heart disease, heart disease and I/R injury (Zhang et al., 1999; Kikuchi et al., 2013). Moreover, the preclusion of oxidative stress and inflammation-induced apoptosis may serve as a favorable effect for the treatment of the myocardial injury. It is well documented that apoptosis is regulated via two pathways such as extrinsic and intrinsic cell death pathways, which are correspondingly characterized via caspase and Bcl-2 family (Zhang et al., 1999). In this experimental study, we focused on the intrinsic pathway. The intrinsic pathway is arbitrated through the activation of caspase and mitochondrial dysfunction. The last apoptosis pathway is attributed via activation of caspase-12. During the I/R injury, ischemia, particularly combined with the reperfusion, triggers translocation of Bax into the outermost layer of mitochondrial membrane, which is related to the increased level of Bax and decreased level of Bcl-2/ Bax ratio (Zhang et al., 2017; Xu et al., 2019). Among all of the proteins, caspase is the significant endogenous inhibitor of apoptosis. During the apoptosis, XIAP binds with the caspase especially caspase-3, 7 and 9, a process that may be regulated via Smac/Diablo and HtrA2/Omi (both are the inhibitor of XIAP). It has been shown as the increased level of caspase-3 reduces the cardiomyocyte apoptosis and also decreases the myocardial infarct area. In the current experimental study, naringenin significantly reduced the level of caspase-3 and also contribute to its observed anti-apoptotic mode of action.

## **CONCLUSION**

In the current experimental study, we scrutinized the cardio-protective effect of naringenin against the I/R injury and also scrutinized the potential effect against the H9c2 cells (*in-vitro*) via an antioxidant mechanism. Naringenin could be a promising therapeutic drug against myocardial ischemic cardiovascular disease. Moreover, a further detailed investigation is conducted to elucidate the mechanisms involved in the cardioprotective effect of naringenin in detail.

Statement of conflict of interest

The authors have declared no conflict of interest.

#### REFERENCES

- Abdalla, A.N., Almaliki, W.H., Mukhtar, M.H., Anwar, F., Shahid, I., Menshawi, S.A. and Alsulimani, T., 2016. Ameliorative influence of dietary dates on doxorubicin-induced cardiac toxicity. *Pharmacol. Pharm.*, 7: 343-353. https://doi.org/10.4236/pp.2016.78042
- Agouni, A., Chalopin, M., Martinez, M.C. and Andriantsitohaina, R., 2011. Protection by red wine polyphenols against metabolic and cardiovascular alterations associated with obesity: A possible link with estrogen alpha receptor. *J. Wine Res.*, **22**: 151-157. https://doi.org/10.1080/09571264.2011.603245
- Anderson, L., Brown, J.P., Clark, A.M., Dalal, H., Rossau, H.K., Bridges, C. and Taylor, R.S., 2011. Patient education in the management of coronary heart disease. *Cochrane Database Syst. Rev.*, **6**:CD008895.
- Burke, A.P. and Virmani, R., 2007. Pathophysiology of acute myocardial infarction. *Med. Clin. North Am.*, **91**: 553-572. https://doi.org/10.1016/j.mcna.2007.03.005
- Darby, S.D., Ewertz, M., McGale, P., Bennet, A.M., Blom-Goldman, U., Brønnum, D., Correa, R.N., Cutter, D., Gagliardi, G., Gigante, B., Jensen, M. and Nisbet, A., 2013. Risk of ischemic heart disease in women after radiotherapy for breast cancer. *N. Engl. J. Med.*, **368**: 987-998. https://doi.org/10.1056/NEJMoa1209825
- Gandhi, H., Patel, V.B., Mistry, N., Patni, N., Nandania, J. and Balaraman, R., 2013. Doxorubicin mediated cardiotoxicity in rats: Protective role of felodipine on cardiac indices. *Environ. Toxicol. Pharmacol.*, 36: 787-795. https://doi.org/10.1016/j. etap.2013.07.007
- Gibellini, L., Pinti, M., Nasi, M., Montagna, J.P., Biasi, S.D., Roat, E., Bertoncelli, L., Cooper, E.L. and Cossarizza, A., 2011. Quercetin and cancer chemoprevention: Evidence-based complement. *Altern. Med.*, **2011**: 591356. https://doi.org/10.1093/ecam/neq053

- Hashmi, S. and Al-Salam, S., 2015. Acute myocardial infarction and myocardial ischemia-reperfusion injury: A comparison. *Int. J. clin. exp. Pathol.*, 8: 8786-8796.
- Hausenloy, D.J. and Yellon, D.M., 2013. Myocardial ischemia-reperfusion injury: A neglected therapeutic target. *J. clin. Invest.* **123**: 92-100. https://doi.org/10.1172/JCI62874
- Hu, T., Wei, G., Xi, M., Xi, M., Yan, J., Wu, X., Wang, Y., Zhu Y., Wang, C. and Wen, A., 2016. TincSynergistic cardioprotective effects of Danshensu and hydroxysafflor yellow A against myocardial ischemia-reperfusion injury are mediated through the Akt/Nrf2/HO-1 pathway. *Int. J. mol. Med.*, 38: 83-94. https://doi.org/10.3892/ijmm.2016.2584
- Ibáñez, B., Heusch, G., Ovize, M., Van, D. and Werf, F., 2015. Evolving therapies for myocardial ischemia/ reperfusion injury. J. Am. Coll. Cardiol., 65: 1454-1471. https://doi.org/10.1016/j.jacc.2015.02.032
- Jiang, H., Wood, K.V. and Morgan, J.A., 2005. Metabolic engineering of the phenylpropanoid pathway in saccharomyces cerevisiae. *Appl. environ. Microbiol.*, 71: 2962–2969. https://doi. org/10.1128/AEM.71.6.2962-2969.2005
- Kalogeris, T., Baines, C.P., Krenz, M. and Korthuis, R.J., 2012. Cell biology of ischemia/reperfusion injury. *Int. Rev. Cell mol. Biol.*, **298**: 229-317. https://doi.org/10.1016/B978-0-12-394309-5.00006-7
- Kikuchi, K., Tancharoen, S., Takeshige, N., Yoshitomi, M., Morioka, M., Murai, Y. and Tanaka, E., 2013. The efficacy of edaravone (radicut), a free radical scavenger, for cardiovascular disease. *Int. J. mol. Sci.*, 14: 13909-13930. https://doi.org/10.3390/ ijms140713909
- Maheshwari, A., 2015. Cardiovascular outcomes in diabetes and strategies to minimize? *World Heart J.*, 7: 1-2.
- Malorni, W., Testa, U., Rainaldi, G., Tritarelli, E. and Peschle, C., 1998. Oxidative stress leads to a rapid alteration of transferrin receptor intravesicular trafficking. *Exp. Cell Res.*, **241**: 102-116. https://doi.org/10.1006/excr.1998.4020
- Moghadam, M.H., Imenshahidi, M. and Mohajeri, S.A., 2013. Antihypertensive effect of celery seed on rat blood pressure in chronic administration. *J. med. Fd.*, **16**: 558-563. https://doi.org/10.1089/jmf.2012.2664
- Moghimian, M., Faghihi, M., Karimian, S.M., RezaImani, A., Houshmand, F. and Azizi, Y., 2013. The role of central oxytocin in stress-induced cardioprotection in ischemic-reperfused

- heart model. *J. Cardiol.*, **61**: 79-86. https://doi.org/10.1016/j.jjcc.2012.08.021
- Müller-Nordhorn, J. and Willich, S.N., 2016. Coronary Heart Disease. *Int. Encycl. Public Hlth.*, 159-167. https://doi.org/10.1016/B978-0-12-803678-5.00090-4
- Nader, M.A., El-Agamy, D.S. and Suddek, G.M., 2010. Protective effects of propolis and thymoquinone on development of atherosclerosis in cholesterol-fed rabbits. *Arch. Pharm. Res.*, **33**: 637-643. https://doi.org/10.1007/s12272-010-0420-1
- Nordlie, M.A., Wold, L.E. and Kloner, R.A., 2005. Genetic contributors toward increased risk for ischemic heart disease. *J. mol. cell. Cardiol.*, **39**: 667-679. https://doi.org/10.1016/j.yjmcc.2005.06.006
- Pertoldi, C., Breyne, P., Cabria, M.T., Halfmaerten, H.A.H. and Jansman, K., 2006. Genetic structure of the European polecat (*Mustela putorius*) and its implication for conservation strategies. *J. Zool.*, **270**: 102-115. https://doi.org/10.1111/j.1469-7998.2006.00095.x
- Plosker, G.L. and McTavish, D., 1995. Captopril: A review of its pharmacology and therapeutic efficacy after myocardial infarction and in ischaemic heart disease. *Drugs Aging*, 7: 226-253. https://doi.org/10.2165/00002512-199507030-00007
- Raedschelders, K., Ansley, D.M. and Chen, D.D.Y., 2012. The cellular and molecular origin of reactive oxygen species generation during myocardial ischemia and reperfusion. *Pharmacol. Ther.*, **133**: 230-255. https://doi.org/10.1016/j.pharmthera.2011.11.004
- Sanada, S., Komuro, I. and Kitakaze, M., 2011. Pathophysiology of myocardial reperfusion injury: Preconditioning, postconditioning, and translational aspects of protective measures. *Am. J. Physiol.*, **301**: H1723-H1741. https://doi.org/10.1152/ajpheart.00553.2011
- Sarvari, S.I., Haugaa, K.H., Anfinsen, O.G., Leren, T.P., Smiseth, O.A., Kongsgaard, E., Amlie, J.A. and Edvardsen, T., 2010. Malignant arrhythmias in patients with arrhythmogenic right ventricular cardiomyopathy are related to right ventricular mechanical dispersion. *Eur. Heart J.*, **32**: 1089-1096. https://doi.org/10.1093/eurheartj/ehr069
- Seki, A. and Fishbein, M.C., 2014. Ischemic Heart Disease. *Pathobiol. Hum. Dis. Dyn. Encycl. Dis. Mech.*, 995-1013. https://doi.org/10.1016/B978-0-12-386456-7.03305-0
- Shekhawat, G.S., Kumar, D., Verma, K., Singh, K., Jana, S.J., Teotia, P. and Yadav, S., 2009. Copper as a biometal: An insight on transport, toxicity

and tolerance in photosynthetic organisms. In: *Biometals: Molecular structures, binding properties and applications* (eds. G. Blanc and D. Moreau). Nova Science Publishers Inc, New York, USA. pp. 23-34.

- Shu, Z., Yang, Y., Yang, L., Yang, L., Jiang, H., Yu, X. and Wang, Y., 2019. Cardioprotective effects of dihydroquercetin against ischemia reperfusion injury by inhibiting oxidative stress and endoplasmic reticulum stress-induced apoptosis: Via the PI3K/Akt pathway. *Fd. Funct.*, **10**: 203-215. https://doi.org/10.1039/C8FO01256C
- Slavíková, J, Kuncová, J. and Topolčan, O., 2007. Plasma catecholamines and ischemic heart disease. *Clin. Cardiol.*, **30**: 326-330. https://doi.org/10.1002/clc.20099
- Smeriglio, A., Barreca, D., Bellocco, E. and Trombetta, D., 2017. Proanthocyanidins and hydrolysable tannins: occurrence, dietary intake and pharmacological effects. *Br. J. Pharmacol.*, **174**: 1244-1262. https://doi.org/10.1111/bph.13630
- Tang, J.Y., Jin, P., He, Q., Lin-He, Lu, L.H., Ma, J.P., Gao, W.L., Bai, H.P. and Yang, J., 2017. Naringenin ameliorates hypoxia/reoxygenation-induced endoplasmic reticulum stress-mediated apoptosis in H9c2 myocardial cells: involvement in ATF6, IRE1α and PERK signaling activation. *Mol. cell. Biochem.*, 424: 111-122. https://doi.org/10.1007/s11010-016-2848-1
- Villani, F., Galimberti, M. and Zunino, F., 1991. Prevention of doxorubicin-induced cardiomyopathy by reduced glutathione. *Cancer Chemother: Pharmacol.*, **28**: 365–369. https://doi.org/10.1007/BF00685691
- Vishnupriya, P. and Padma, V., 2017. A review on the Antioxidant and therapeutic potential of *Bacopa monnieri*. *React. Oxyg. Species*, **3**: 111–120. https://doi.org/10.20455/ros.2017.817
- Wang, F., Smith, N.A., Xu, Q., Fujita, T., Baba, A., Matsuda, T., Takano, T., Bekar, L. and Nedergaard, M., 2012. Astrocytes modulate neural network activity by Ca 2+-dependent uptake of extracellular K +. Sci. Signal, 5: ra26. https://doi.org/10.1126/ scisignal.2002334
- Wang, Y., Guo, L., Golding. I., Cox, E.C. and Ong, N.P., 2009. Quantitative transcription factor binding kinetics at the single-molecule level. *Biophys. J.*, **96**: 609-620. https://doi.org/10.1016/j.bpj.2008.09.040
- Wright, D.H., Abran, D., Bhattacharya, M., Hou, X., Bernier, S.G., Bouayad, A., Fouron, J.C., Vazquez-Tello, A., Beauchamp, M.H., Clyman,

- R.I., Peri, K., Varma, D.R. and Chemtob, S., 2001. Prostanoid receptors: Ontogeny and implications in vascular physiology. *Am. J. Physiol. Regul. Integr. Comp. Physiol.*, **281**: R1343-R1360. https://doi.org/10.1152/ajpregu.2001.281.5.R1343
- Xu, H., Wang, J., Cai, J., Feng, W., Wang, Y., Liu, Q. and Cai, L., 2019. Protective effect of lactobacillus rhamnosus gg and its supernatant against myocardial dysfunction in obese mice exposed to intermittent hypoxia is associated with the activation of nrf2 pathway. *Int. J. biol. Sci.*, **15**: 2471-2483. https://doi.org/10.7150/ijbs.36465
- Yin, Y., Guan, Y., Duan, J., Wei, G., Zhu, Y., Quan, W., Guo, C., Zhou, D., Wang, Y., Xi, M. and Wen, A., 2013. Cardioprotective effect of Danshensu against myocardial ischemia/reperfusion injury and inhibits apoptosis of H9c2 cardiomyocytes via Akt and ERK1/2 phosphorylation. *Eur. J. Pharmacol.*, **699**: 219-226. https://doi.org/10.1016/j.ejphar.2012.11.005
- Yu, L.M., Dong, X., Zhang, J., Li, Z., Xue, X., Wu, H., Yang, Z., Yang, Y. and Wang, H., 2019. Naringenin attenuates myocardial ischemia-reperfusion injury via cgmp-pkgiα signaling and *in vivo* and *in vitro* studies. *OxiMed. cell. Longev.*, **2019**: 7670854. https://doi.org/10.1155/2019/7670854
- Zeng, Y., Li, Y. and Yang, J., 2017. Therapeutic role of functional components in alliums for preventive chronic disease in human being. *Evidence-based complement. Altern. Med.*, **2017**: 9402849. https://doi.org/10.1155/2017/9402849
- Zhang, L., Xiao, Y. and Jiale, H.E., 1999. Cocaine and apoptosis in myocardial cells. *Anat. Rec.*, **257**: 208–216. https://doi.org/10.1002/(SICI)1097-0185(19991215)257:6<208::AID-AR6>3.0.CO;2-0
- Zhang, Z., Zhang, Y. and Li, J., 2017. MicroRNA-26b protects against the H2O2-induced injury on cardiac myocytes via its target gene HGF. *Int. J. clin. exp. Pathol.*, **90**: 149–156.
- Zhu, B., Zhang, Q-L., Hua, J-W., Chang, W-L. and Qin, L-P., 2018. The traditional uses, phytochemistry, and pharmacology of *Atractylodes macrocephala* Koidz: A review. *J. Ethnopharmacol.*, **226**: 143-167. https://doi.org/10.1016/j.jep.2018.08.023
- Zygmunt, K., Faubert, B., MacNeil, J. and Tsiani, E., 2010. Naringenin, a citrus flavonoid, increases muscle cell glucose uptake via AMPK. *Biochem. biophys. Res. Commun.*, **398**: 178-183. https://doi.org/10.1016/j.bbrc.2010.06.048