



Research Article

Application of Furfural in Sugarcane Nematode Pest Management

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Abstract | Sugarcane, *Saccharum officinarum* is a foremost crop which is widely grown for sugar production. A major impediment to sugarcane cultivation is the infestation by plant parasitic nematodes most notably, the cyst nematode *Heterodera sacchari*. Broadly, synthetic nematicides are utilized in the suppression of soil nematode infestation of sugarcane, with positive outcome in yield. However, emergence of resistant nematode strains and health hazards are associated with the ceaseless use of the synthetics. The efficacy of furfural (2-furfuraldehyde) from agricultural biomass waste was examined as a practicable replacement to the regular synthetic nematicide. Agricultural wastes were collected and refluxed with hydrochloric acid to produce furfural. Growth response of sugarcane plants increased significantly ($p=0.05$) with the highest concentration (75 mg) of furfural. A reduction in nematode population in soil of treated plants was remarked. The results indicated that furfural could be practically applied in the management of nematode pests of sugarcane, while safe guarding the environment from pollution.

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Keywords | Furfural, *Heterodera sacchari*, Sugarcane, Carbofuran, Agricultural wastes

Introduction

The sugarcane plant *Saccharum officinarum*, belongs to the family poaceae, which is an economically predominant grass family. It is a principal crop which accounts for 75% of global sugar production in the Sub-tropical and Tropical regions of the world (Chengarayan and Gallo-Meagher, 2001; Vilela, 2017). Brazil, India and Peru are the largest sugarcane producing countries (Bogden, 2007). The sugarcane plant has the highest bio conversion efficiency among many plants in the same family, the dry matter after harvest can be used for fuel production, while the bagasse is employed in paper making as pulp and it can serve efficiently in solar power fixing (Rainey et al., 2006). Nations with high production of sugarcane are dependent on the bagasse for electricity generation. A ton of bagasse is known to produce over 100 kWh of electricity (Ogden, 1990). Similarly, ethanol another

by product of sugarcane processing can be deployed to bio fuel and could serve as an alternative to gasoline (Da Rosa, 2005). Plant parasitic nematodes are one of the key pests of diverse crops (Fabiyi et al., 2020; Fabiyi, 2021a, b, c). Several plant parasitic nematodes are associated with the sugarcane plant, although planted crops are not as susceptible as the ratoon crops to nematode infestation (Berry et al., 2007). All crops grown are vulnerable to attack from one plant parasitic nematode or the other (Bridge and Star, 2007). *Heterodera sacchari*, the cyst nematode is salient to vital crops (Fabiyi et al., 2018). High yield losses of about \$125 billion is attributed to nematode infestation (Chitwood, 2003). Control of plant parasitic nematodes in commercial cane plant production is by the use of synthetic nematicides (Donga and Eklo, 2018). Some of the consequences of unregulated pesticide use are environmental pollution, human health problems, and pesticide residue contaminants (Fabiyi and

Olatunji, 2021). The physicochemical characteristics of most nematicides make them mobile in soil and thus contaminate ground water through run off (Pedlowski *et al.*, 2012). Many synthetic nematicides are banned due to their toxicity and negative impact in the environment (Haydock *et al.*, 2014). Hence, the aim of this research is to produce furfural from agricultural wastes and evaluate the effectiveness on the sugarcane plant nematode *Heterodera sacchari*.

Materials and Methods

Reagents

All reagents used were of analytical grade, these include dichloromethane, acetic acid, hydrochloric acid, ethanol, distilled aniline, and anhydrous magnesium sulphate.

Plant materials

Dry plantain leaves (PLT), sugarcane bagasse (SB) and dry pawpaw stem (PS) were obtained from Ilorin metropolis. The materials were air dried for two weeks and then powdered using a STEEL MAN diesel engine with 7 Horse power capacity; model R175A.

Hydrolysis of plant materials with dilute acids

300 grams each of milled sugarcane bagasse (SB), dry plantain leaves (PLT) and dry pawpaw stem (PS) was packed into 2000 mL quick-fit flask separately in batches, 1500 mL of dilute 8 % hydrochloric acid was added to the flask, the materials were allowed to soak properly into the acid solution to avoid charring, the flask was then connected to a reflux condenser and heated on a sand bath at 200°C for 2^{1/2} hours (Ali *et al.*, 2002; Fabiyi *et al.*, 2018). After hydrolysis crude furfural was decanted and filtered. The same process was repeated for each waste material using dilute 8 % hydrochloric acid. The procedure was carefully repeated sequentially until 1kg of each of the agro-waste material was exhausted.

Extraction of furfural

100 mL of the crude furfural solutions from the various hydrolysis was poured into a separating funnel, while 100 mL of dichloromethane (DCM) was added. The organic layer which contains the furfural was separated, dried with anhydrous magnesium sulphate; and filtered. The furfural was obtained by steam distillation (Montane *et al.*, 2002; Fabiyi *et al.*, 2018). The extracted furfural was collected and stored in sample bottles. This procedure was repeated for all

the crude furfural extracted from each of the agro-waste material. Each extracted furfural was coded FFR/PLT, FFR/PS and FFR/SB, that is, furfural from plantain leaves, furfural from pawpaw stem and furfural from sugarcane bagasse respectively.

Screenhouse experiment

Loamy soil was collected and pasteurized at 70°C for three hours. The soil was allowed to cool and then distributed into experimental pots of 30 cm diameter at 25 Kg each and were set aside to stabilize for two weeks. The experiment was a randomised complete block design (RCBD). There were four treatments at four levels and each with three replicates. Cane setts were planted and inoculated three weeks after planting with approximately 500 eggs and juveniles of *Heterodera sacchari* in a hole made at the base of each cane setts (Fabiyyi, 2019; Fabiyi *et al.*, 2019). Treatments were applied at 25, 50 and 75mg. The untreated plants served as the control, while all treatments were likened to carbofuran as the standard check.

Statistical analysis

Data were collected on heights of each cane plant, number of leaves of each cane plant and nematode population in 200 g soil. parameters were subjected to two-way analysis of variance (ANOVA). Treatment means, where necessary were separated using the new Duncan's multiple range test at 5 % level of probability (Gomez and Gomez, 1984).

Results and Discussion

Results from Table 1 reveals that there was no notable ($p=0.05$) difference in the numbers of leaves of cane plants treated with different furfural from each of the waste materials. The numbers of leaves were remarkably low in the untreated control plants. The highest (75 mg) dosage of application of furfural produced more leaves. Table 2 represents the response of sugarcane plants treated with furfural. Appreciable ($p=0.05$) differences were observed in the height of the cane plants. Furfural extracted from sugarcane bagasse was consequentially more effective than furfural produced from other waste materials. Significantly low height was recorded in untreated cane plants. Similarly, nematode population was reduced in cane plants treated with furfural from sugarcane bagasse. Higher nematode population was detected in the untreated control cane sugar plants (Table 3).

Table 1: Effects of treatment and treatment concentration on the number of leaves of sugarcane plants.

Treatment	9WAP	10WAP	11WAP	12WAP	13WAP	14WAP	15WAP	16WAP	17WAP	18WAP
CBFN	8 ^a	8 ^a	9 ^a	9 ^a	10 ^a	10 ^a	11 ^a	12 ^a	13 ^a	15 ^a
FFR/PLT	8 ^a	8 ^a	9 ^a	9 ^a	11 ^a	11 ^a	12 ^a	12 ^a	13 ^a	14 ^a
FFR/SB	7 ^a	8 ^a	9 ^a	9 ^a	11 ^a	11 ^a	12 ^a	13 ^a	13 ^a	13 ^b
FFR/PS	7 ^a	8 ^a	9 ^a	9 ^a	10 ^a	11 ^a	12 ^a	13 ^a	13 ^a	12 ^b
CONTROL	5 ^b	5 ^b	6 ^b	6 ^b	7 ^b	8 ^b	8 ^b	9 ^b	10 ^b	11 ^c
Dosage/Mg										
One (25 mg)	6	7	7 ^b	8	8 ^b	8 ^b	9 ^b	9 ^b	10 ^c	13 ^b
Two (50 mg)	6	7	7 ^b	8	9 ^{ab}	10 ^a	10 ^b	11 ^a	12 ^b	12 ^c
Three (75 mg)	6	8	9 ^a	9	10 ^a	11 ^a	12 ^a	12 ^a	14 ^a	16 ^a

Treatment with the same letters in the column are not significantly different at $p=0.05$, using the new Duncan's multiple range test.

Table 2: Effects of treatment and treatment concentration on the heights of sugarcane plants.

Treatment	9WAP	10WAP	11WAP	12WAP	13WAP	14WAP	15WAP	16WAP	17WAP	18WAP
CBFN	172.6 ^a	191.3 ^a	196.8 ^a	201.0 ^a	208.5 ^a	212.0 ^a	219.0 ^a	229.0 ^a	236.1 ^a	241.0 ^a
FFR/PLT	130.1 ^c	152.7 ^c	158.1 ^c	170.1 ^c	178.2 ^c	188.5 ^c	200.1 ^b	219.7 ^b	226.4 ^b	230.8 ^b
FFR/SB	159.2 ^b	165.3 ^b	173.0 ^b	182.3 ^b	187.0 ^b	196.6 ^b	202.3 ^b	228.0 ^a	235.5 ^a	240.8 ^a
FFR/PS	131.8 ^c	150.2 ^c	162.8 ^c	169.5 ^c	178.7 ^c	187.2 ^c	190.5 ^b	197.3 ^c	204.1 ^c	209.7 ^c
CONTROL	115.6 ^d	126.0 ^d	137.4 ^d	145.0 ^d	149.4 ^d	153.0 ^d	159.3 ^d	167.2 ^d	176.0 ^d	185.4 ^d
Dosage/Mg										
One (25 mg)	120.0 ^c	130.9 ^c	137.5 ^c	145.7 ^c	150.7 ^c	158.4 ^c	161.0 ^c	169.2 ^c	175.4 ^c	192.7 ^b
Two (50 mg)	131.3 ^b	143.3 ^b	151.1 ^b	159.3 ^b	166.8 ^b	179.3 ^b	185.5 ^b	190.8 ^b	196.8 ^b	201.6 ^b
Three (75 mg)	143.6 ^a	159.0 ^a	164.0 ^a	170.4 ^a	184.0 ^a	190.0 ^a	206.9 ^a	211.2 ^a	217.0 ^a	221.9 ^a

Treatment with the same letters in the column are not significantly different at $p=0.05$, using the new Duncan's multiple range test.

Table 3: Effects of treatment and treatment concentration on the population of soil nematodes of sugarcane plants.

Treatments	Soil nematode
CBFN	35 ^a
FFR/PLT	129 ^c
FFR/SB	66 ^b
FFR/PS	143 ^d
CONTROL	1579 ^c
Dosage/Mg	
One (25 mg)	155 ^c
Two (50 mg)	97 ^b
Three (75 mg)	49 ^a

Treatment with the same letters in the column are not significantly different at $p=0.05$, using the new Duncan's multiple range test.

Furfural (2-Furancarboxaldehyde), a naturally occurring compound which is prepared by acid hydrolysis of pentosans is nematicidal (Marcotullio, 2013). The efficacy of furfural at different concentrations on *Meloidogyne javanica* was evaluated by Al-Hamdany et al. (1999). They established that furfural at 1000 and 2000 ppm significantly reduce

the egg masses and root gall index of cucumber plant. With 4000 ppm of furfural there was no galling. Equivalently, Stephan et al. (2001), reported the effectiveness of furfural as a pre-planting nematicide at 4000 ppm for tomato plants, while El-Mougy et al. (2008), in green house and field experiments also affirmed that furfural is nematicidal with a sharp decline in *Meloidogyne incognita* population. Commercial furfural was tested by Kokalis-Burelle (2007) on *M. incognita* infecting tomato. Reduced galling and good vegetative growth was spotted. In laboratory experiments, furfural at 0.4 mL/L soil to small pots reduced numbers of *Helicotylenchus dihystera*, *Paratrichodorus* spp., *Pratylenchus zaeae*, *Tylenchorhynchus* sp., *Xiphinema elongatum* and *X. mampara* (Spaull, 1997). In the same vein, furfural synthesised from agro-cellulosic materials depicted strong nematicidal activity on *M. incognita* parasitizing groundnut plants (Fabiyyi, 2020) All the above corroborated the findings in this research. Considerable reduction in population of *Heterodera saccharri* was exhibited in cane plants treated with the

highest concentration of furfural. A striking increase in height of sugarcane plants was recorded in treated plants, which translates to yield increase.

Conclusions and Recommendations

Furfural can be exploited in place of the toxic synthetic nematicides in the curb of cyst nematode parasitizing cane plants. The dissimilarities observed in the results may be due to the existence of impurities in the materials utilized for hydrolysis

Novelty Statement

This is the first report on the application of furfural in the control of *Heterodera saccharri* on sugarcane plants in Nigeria.

Conflict of interest

The authors have declared no conflict of interest.

References

- Al-Hamdany M.A., Al-Noaimi H.N., Aboud H.M., and Salih H.M., 1999. Use of furfural for control of the root-knot nematode *Meloidogyne javanica* on cucumber and eggplant under greenhouse conditions. Arab. J. Plant Prot., 17(2): 84-87.
- Ali, S., Fiyaz, A.C., Najia, I. and Kiran, A., 2002. Effect of chemical treatment on the production of furfural and active carbon from rice husk. Int. J. Agric. Biol., 1: 23-25.
- Berry, S., Spaul, V.W. and Cadet, P., 2007. Impact of harvesting practices on nematode communities and yield of sugarcane. Crop Prot., 26(8): 1239-1250. <https://doi.org/10.1016/j.cropro.2006.10.022>
- Bogden, A.V., 2007. *Tropical pasture and fodder plants (tropical agriculture)* Longman group (Far East), Limited. ISBN 978-0582466760.
- Bridge, J. and Starr, J., 2007. Plant nematodes of agricultural importance Manson, London. <https://doi.org/10.1201/b15142>
- Chengarayan, K. and Gallo-Meagher, M., 2001. Effect of various growth regulation of sugarcane. *In vitro* Cell Dev. Biol., 37: 434-439. <https://doi.org/10.1007/s11627-001-0076-0>
- Chitwood, D., 2003. Research on plant-parasitic nematode biology conducted by the United States Department of agriculture agricultural research services. Pest Manage. Sci., 59: 748-753. <https://doi.org/10.1002/ps.684>
- Da Rosa, A., 2005. *Fundamentals of renewable energy processes*. Elsevier. ISBN 978-0-12-088510-7. pp. 501-502.
- Donga, T.K. and Eklo, O.M., 2018. Environmental loads of pesticides used in conventional sugarcane production in Malawi. Crop Prot., 108: 71-77. <https://doi.org/10.1016/j.cropro.2018.02.012>
- El-Mougy N.S., El-Gamal, N.G., Mohamed M.M.M., and Abdel-Kader, M.M., 2008. Furfural approaches as control measures against root rot and root-knot incidence of tomato under greenhouse and field conditions. J. Plant Prot. Res., 48(1): 93-105. <https://doi.org/10.2478/v10045-008-0010-0>
- Fabiyi, O.A. and Olatunji, G.A., 2021. Environmental sustainability: Bioactivity of *Leucaena leucocephala* leaves and pesticide residue analysis in tomato fruits. Acta Univ. Agric. Silv. Mendel. Brun., 69(4): 473-480. <https://doi.org/10.11118/actaun.2021.042>
- Fabiyi, O.A., 2019. Management of groundnut (*Arachis hypogea*) Root-knot nematode (*Meloidogyne incognita*): Effect of *Prosopis africana* Pods. Indian J. Nematol., 49(2): 214-216.
- Fabiyi, O.A., 2020. Growth and yield response of groundnut *Arachis hypogaea* (Linn.) under *Meloidogyne incognita* infection to furfural synthesized from agro-cellulosic materials. J. Trop. Agric., 58(2): 241-245.
- Fabiyi, O.A., 2021a. Evaluation of plant materials as root-knot nematode (*Meloidogyne incognita*) suppressant in okro (*Abelmoschus esculentus*). Agric. Conspec. Sci., 86(1): 51-56.
- Fabiyi, O.A., 2021b. Evaluation of nematicidal activity of *Terminalia glaucescens* fractions against *Meloidogyne incognita* on *Capsicum Chinense*. J. Hort. Res., 29(1): 67-74.
- Fabiyi, O.A., 2021c. Sustainable management of *Meloidogyne incognita* infecting carrot: Green synthesis of silver nanoparticles with *Cnidioscolus aconitifolius*: (*Daucus carota*). Vegetos, 34(2): 277-285. <https://doi.org/10.1007/s42535-021-00216-y>
- Fabiyi, O.A., Atolani, O., and Olatunji, G.A., 2020. Toxicity Effect of *Eucalyptus globulus* to *Pratylenchus* spp. of *Zea mays*. Sarhad J. Agric., 36(4): 1244-1253. <https://doi.org/10.17582/>

[journal.sja/2020/36.4.1244.1253](https://doi.org/10.2478/sab-2019-0026)

- Fabiyi, O.A., Olatunji G.A and I.O. and Daodu, I.O., 2019. Nematicidal effect of organic extract metal complex on *Meloidogyne incognita* infecting groundnuts (*Arachis hypogea*). *Sci. Agric. Bohem.*, 50(3): 191-196. <https://doi.org/10.2478/sab-2019-0026>
- Fabiyi, O.A., Osunola, O.S., Olatunji, G.A., and Umar, K.A., 2018. Efficacy of agricultural wastes in the control of rice cyst nematode (*Heterodera sacchari*). *Agric. Conspec. Sci.*, 83(4): 329-334.
- Gomez, K.A. and Gomez, A.A., 1984. Statistical procedures for agricultural research, 2nd Edn. John Wiley and Sons, New York.
- Haydock, P., Woods, S., Grove, I., and Hare, M., 2014. Chemical control of nematodes. R. Perry, M. Moens (Eds.). *Plant Nematology*. CABI. Wallingford. UK. <https://doi.org/10.1079/9781780641515.0459>
- Kokalis-Burelle, N., 2007. Effects of furfural on nematode populations and galling on tomato and pepper. *Nematropica*, 37: 307-316.
- Marcotullio, G., 2013. SEA Servizi Energia Ambiente srl-UNECE/FAO Workshop St. Petersburg 22-24/05/2013.
- Montane, D., Salvado, J., Torras, C., and Farriol, X., 2002. High temperature dilute acid hydrolysis of olive stones for furfural production. *Biomass Bioenergy*, 22: 395-304. [https://doi.org/10.1016/S0961-9534\(02\)00007-7](https://doi.org/10.1016/S0961-9534(02)00007-7)
- Ogden, 1990. Steam economy and cogeneration in cane sugar factories. *Int. Sugar J.*, 92(1099): 131-140.
- Pedlowski, M.A., Canela, M.C., Terra, M.A.C. and Faria, R.M.R., 2012. Modes of pesticides utilization by Brazilian smallholders and their implications for human health and the environment. *Crop Prot.*, 31(1): 113-118. <https://doi.org/10.1016/j.cropro.2011.10.002>
- Rainey, T., Covey, G. and Shore, D., 2006. An analysis of Australian sugarcane regions for bagasse paper manufacture. (<http://eprints.qut.edu.au/6781/>). *Int. Sugar J.*, 108(1295): 640-644.
- Spaull, V.W., 1997. On the use of furfural to control nematodes in sugarcane. *Proc. South Afr. Sugar Technol. Assoc.*, pp. 71.
- Stephan, Z.A., Al-Hamadany, M.A., Al-Din, S.S., and Dawood, H.B., 2001. Efficacy of furfural treatment in controlling the disease complex of root-knot nematode and Fusarium wilt on tomato and egg-plant under Lathouse conditions. *Arab. J. Plant Prot.*, 19: 97-100.
- Vilela, Del-Bem, 2017. Analysis of three sugarcane homo/homeologous regions suggests independent polyploidization events of *Saccharum officinarum* and *Saccharum spontaneum*. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5381655>. *Genome Biol. Evol.*, 9(2): 266-278. <https://doi.org/10.1093/gbe/evw293>