#### http://dx.doi.org/10.18681/pjn.v35.i02.p157-173

# Two new records of entomopathogenic nematodes (Nematoda: Steinernematidae) from Gilgit-Baltistan, Pakistan

K. A. Tabassum,<sup>†</sup> J. Salma and H. Sagir

National Nematological Research Centre, University of Karachi, Karachi-75270, Pakistan

<sup>†</sup>Corresponding author: tabassumak@uok.edu.pk

#### Abstract

A survey of District Hunza, Gilgit and Nager, Gilgit-Baltistan, Pakistan was conducted during January, 2017. Out of 20 soil samples, 6 samples were found positive for *Steinernema* species. One sample of *Prunus avium* had *Steinernema affine* from Hunza, two samples had *S. cholashanense* from *Juglans nigra* and *Malus pumila* from Gilgit, and three samples harboured *S. feltiae* from *Prunus armeniaca*, *Prunus persica* and *Malus pumila* from Nager. This paper deals with re-description of *S. affine* based on comparative study of morphological and molecular characteristics with brief study of *S. cholashanense* and *S. feltiae*. *S. affine* Pak. G.S.356 deposited in NCBI with accession number MF150033; *S. cholashanense* Pak.G.S.350 and Pak. G.S.355 with accession numbers MF125282 and MF0339642, respectively; *S. feltiae* Pak. G.S. 354, 357, 358 deposited with accession numbers MF150034, MF158314 and MF144570, respectively.

Keywords: Taxonomy, Steinernema affine, S. cholashanense, S. feltiae, new records, Gilgit - Baltistan.

**N**ational Nematological Research Centre (NNRC), University of Karachi, Pakistan, has conducted many surveys in Pakistan to isolate EPNs for effectively utilized to control important insect pests. Studies of EPN in Pakistan have discovered six new species along with seven known species (Shahina *et al.*, 2017).

This paper deals with re-description of *S. affine* with brief study of *S. cholashanense* and *S. feltiae.* According to Nguyen & Hunt, 2007 *Steinernema affine* was described in 1937 by Bovien as *Neoplactana affinis* from diseased fly larvae of bibionid fly in Denmark. In 1971, *N. affinis* re-isolated from larvae of Danish bibionid by Poinar & Lindhardt and deposited in the Davis Nematode Collection at University of California, Davis, CA, USA. Both aforesaid scientists studied *N. affinis* and *N. binionis*, they play the same role as parasites of bibionid flies in Denmark. Where as in 1979, Poinar re-

described and differentiated both species on morphological basis. In 1988, Poinar designated lectotypes and two paralectotypes, one with male and one female. After that it was redescribed by Mracek et al., (2005a) as S. affine from British, Columbia and Canada. S. affine is widely distributed species and found throughout Europe (Sturhan & Liskova, 1999; Sturhan & Mracek, 2000; Mracek et al., 2005a), UK and Ireland (Griffen et al., 1991; Hominick et al., 1995; Hominick, 2002), Vancouver area, British Columbia, Canada (Mracek et al., 2005b). Hazir et al., (2003) reported S. affine from Turkey and et al., (2015) from Italy. S. Tarasco cholashanense was described by Nguyen et al., (2008) collected from Chola Shan Mountain covered by xerophytic vegetables, Sichuan Province, China. S. feltiae originally described from the larvae of Agrotis (syn. Feltia) sgetum by Filipjev (1934). From Pakistan, it was reported by Anis et al., (2002) from Turbella

Dam, Khyber Pakhtunkhwa after that this species was re-described by Tabassum & Shahina in 2004. During the present study, EPN were further explored from Gilgit-Baltistan, Pakistan. The samples were collected from three districts: Gilgit (35.9197°N. 74.3881°E), Hunza (36.3082°N. 74.6195°E) and Nager (35.24°N. 73.48°E), Gilgit-Baltistan having minimum temperature -1-10 °C during January, 2017. The climate of all three districts varies from region to region, in District Gilgit, maximum temperature ranges between 20-30 °C whereas minimum temperature ranges -0.4- 1 °C (www. wikipedia. org), in Hunza 10-26 Max., -1 to -10 Min. (www. tiptop gleb.com) whereas in Nager 10-15 Max., and-1 to -14 Min. (www. wikipedia.org) have notified.

### **Materials and Methods**

**Nematodes collection and isolation:** Soil samples were collected from different areas of Gilgit-Baltistan by removing 6-10 cm layer of ice at the depth of 6 cm in the rhizosphere of different fruits (Fig. 1). Nematodes were isolated from soil samples using the insect baiting with the last instars of *Galleria mellonella* (L.) following the technique described by Bedding & Akhurst (1975) and kept at 5-10 °C. For the emergence of infective juveniles (IJs) dead larvae were placed in White traps (White, 1927). Nematodes have been maintained through soil assays.

Morphology and morphometrics of the isolated nematodes: Only IJ collected from White traps on the fourth day of emergence were used for measurement. The first and second generation adults were obtained at 3-4 and 6-7 days, respectively. All the different stages were killed by hot water (85 °C), then fixed in TAF (Courtney *et al.*, 1955), water in specimens replaced by glycerin using the Seinhorst technique (Seinhorst, 1959), after which they were mounted in pure glycerin. Permanent slides were made for measurements. Measurements and Photomicrographs were taken using a Nikon E-400 and Nikon DS-Fil, respectively.

Molecular characterization and phylogenetic relationship: DNA was extracted from a single female using the modified method reported by Joyce et al., (1994). Primers TW81 and AB28 were used for amplification of internal transcribed regions ITS1-5.8S-ITS2. For D2D3 expansion segment of the 28S rRNA region, the primer 501 and 391 were used as reported by Nguyen et al., 2006. The method reported by Khatri-Chhetri et al., 2011 were used for PCR amplification. PCR products were purified using DNA purification kit (Promega purification kit) and sequenced. The resulting sequences were edited and analyzed using software packages Chromas 2.23 (Technelysium, Tewantin, QLD, Australia) and Bio Edit 7.0.4.1 (Hall, 1999) and deposited in GenBank with accession numbers MF116307 and MF125282 for S. cholashanense isolate by using ITS; MF150034, MF144570 and NF158314 for S. feltiae, whereas MF150033 for S. affine using D2D3 28S RNA region. The resulting sequences of other species of Steinernema available in GenBank were aligned using Clustal X1.83 (Thompson et al., 1997). Phylogenetic analyses based on Maximum parsimony or pair wise distance of sequences, were done by using PAUP\* v 4.0b10 (Swofford, 2002). Through MrEnt 2.0 (Zuccon & Zuccon, 2010) the resulting trees were visualized. Caenorhabditis *elegans* (EU131007) was applied as an out-group during calculation of the trees based on ITS regions. Cervidellus alutus (AF331911), together with Panagrellus redivivus (AF331910) were used as out group for calculation of the trees based on D2D3 region.

### **Results and Discussion**

Steinernema affine (Bovien, 1937) Wouts, Mracek, Gerdin & Bedding, 1982 (Fig. 2. A-F)

# Measurements: Table 1

**Female (First Generation)**: Body variable in shape, usually C shaped sometimes coiled on

Two new records of entomopathogenic nematodes from Gilgit-Baltistan



Fig.1. Surveyed localities and collection of samples from Gilgit, Baltistan.

heat relaxation. Head rounded some time offset from body, cuticle smooth, six lips each bearing a single labial papilla. Four cephalic papillae present. Amphids distinct, crescent shaped located between labial and cephalic papilla. Lateral field and phasmid inconspicuous. Stoma short shallow, cheilorhabdions distinct. Excretory pore located mostly at the base of metacorpus and always anterior to nerve ring.

Pharynx muscular with cylindrical procorpus extending into a slightly swollen metacorpus, a narrower isthmus surrounded by nerve ring and large valvated basal bulb. Genital tract amphidelphic with reflexed ovaries. Vulva transverse slit like with protruding lips. Narrow rectum, distinct anal opening, postanal swelling reduced or slightly present. Tail usually with a terminal knob at base.

**Female (Second Generation):** Similar to first generation females in general morphology but smaller. Vulval aperture located posterior to mid body, lips protruding. Tail straight conoid which often ends in a fine mucron. Post anal swelling absent or slightly present.

**Males**: Cephalic region, cuticle and pharynx similar to females, except nerve ring usually located at anterior portion of basal bulb. Testis single, well developed, ventrally reflexed. Spicules paired, moderatelly curved and colorless. Spicule head longer than wide, shaft short, velum present. In ventral view, gubernaculum was forked at proximal portion. The distal portion varied from simple to hook shaped. There are 23 genital papillae comprises eleven pairs and a single ventral preanal papillae. Five pairs subventral, anterior to cloaca, three pairs in the region of spicule and gubernaculum, two pairs ventral subterminal, one pair postcloacal, subdorsal.

**Infective juveniles**: Third stage juvenile is enclosed in the second stage cuticle. Cephalic

region finely rounded and offset from body. The mouth and anus are closed, intestine collapsed. Bacterial cells are filled in a modified vesicle in the anterior portion of the intestinal lumen. Lateral field with six ridges (7 lines), having prominent sub-marginal pair. Excretory pore located anterior to nerve ring or at middle of pharynx. Tail sharply pointed, hyaline portion short.

Remarks: Steinernema affine was found from cherry Prunus avium from Village Murtazabad District Hunza, Gilgit-Baltistan, characterized genetically by D2D3 region of rDNA. The having the accession number sequence. MF150033 was deposited in GenBank. The sequence length of S. affine is 598 bp having nucleotide composition, A = 0.23810, C =0.17619, G=0.33810, and T = 0.24762. For D2D3 sequence, MP trees were calculated based on 1059 characters, of which 102 are constant, 183 are variable characters are parsimony uninformative and 774 characters (included) are parsimony informative. The phyogenetic relationships between 46 species of Steinernema are presented in Fig. 3 (For MP, tree length = 3083, Cl = 0.5800, RI = 0.7673, RC = 0.4450,HI = 0.4520). The sequence length composition and pair wise distance have shown in Table 3 and 4, respectively.

The population of S. affine Pak.G.S.356 has some variation in morphometric data. In male shorter body in length (1362 vs 1400-1733) µm; greater distance from head to excretory pore (100-125 vs 82-114) µm; shorter anal body (53-60 VS 52-72) width μm; shorter gubernaculum in length (37-45 vs 37-56); lower D % (61 vs 69); higher SW (151 vs 117). In IJs and female slight differences have been found (Table 1). S. affine is widely distributed throughout Europe, UK, Ireland, Coloumbia, and Canada. From Asian countries S. affine was reported only from Italy and now this species has recovered from Gilgit-Baltistan, Pakistan.

Characters	First Ge	neration	Second G	eneration	Infective stage
	Male	Female	Male	Female	juveniles
TL	$1489 \pm 123$	$3824 \pm 328$	$1293 \pm 120$	$1646 \pm 126$	$675 \pm 35$
	(1362-1733)	(3117-4212)	(1150-1500)	(1460-1820)	(610-835)
Lip width	$14.5 \pm 1.2$	$33.5 \pm 2.7$	$12.5 \pm 1.1$	21 ± 1.6	$10 \pm 0.8$
1	(14-16)	(28-35)	(12-13)	(19-25)	(9-11)
MBW	$93 \pm 5.2$	$179 \pm 37.8$	$64 \pm 15.4$	$91.8 \pm 5.3$	$29 \pm 2.4$
	(80-105)	(140-255)	(60-72)	(85-100)	(27-33)
EP	$112 \pm 10$	$122.4 \pm 15.3$	$94.8 \pm 46$	$98.3 \pm 4.6$	$61 \pm 7.5$
	(100-125)	(107-150)	(85-106)	(90-105)	(50-68)
NR	$124 \pm 5.5$	$158.7 \pm 9.1$	$117 \pm 35$	$112 \pm 8.4$	$95 \pm 12$
	(115-145)	(150-175)	(108-123)	(98-123)	(85-100)
ES	$161 \pm 4.8$	$238 \pm 29$	$159 \pm 55$	$162 \pm 9.8$	$128 \pm 6.5$
	(155-168)	(213-317)	(149-169)	(150-179)	(110-140)
TL (Ijs with	$47 \pm 3.0$	66.4 ± 8	$35.7 \pm 23.6$	$54\pm 6$	67 ± 7
sheath)	(45-50)	(50-75)	(30-42)	(48-64)	(60-75)
Anal body	$55 \pm 2.7$	$60.9 \pm 6.2$	45 ±17.2	$40 \pm 1.5$	$16.5 \pm 2.3$
width	(53-60)	(55-72)	(40-51)	(38-42)	(15-19)
V%		$51.7 \pm 3.7$		$53.7 \pm 2$	
		(46-59)		(51.5-57.6)	
а	$18.4 \pm 1.3$	$22.2 \pm 4.8$	$20 \pm 3.8$	$17.7 \pm 0.68$	$26 \pm 1.8$
	(16.5-20.7)	(16-30)	(18.5-24.5)	(17-19)	(20-28)
b	$9.0 \pm 0.8$	$16 \pm 1.8$	$8 \pm 0.6$	$10 \pm 0.79$	$5.8 \pm 1.2$
	(8-10.2)	(12.8-18.7)	(6.8-9.6)	(8.3-11)	(5.1-6.0)
с	$30 \pm 3.0$	$59.4 \pm 11.6$	$36.3 \pm 11.5$	$30.6\pm3.2$	$10 \pm 1.5$
	(27-34)	(52-70)	(30-40.6)	(25-35.7)	(9-11.5)
c'	$0.78\pm0.01$	$1.08\pm0.13$	$0.78\pm0.07$	$1.34\pm0.11$	$0.9\pm0.07$
	(0.75 - 0.89)	(0.9-13)	(0.72 - 0.9)	(1.2-1.5)	(0.7-1.0)
D %	$69 \pm 6.4$	$52\pm9.6$	$59.5\pm5.1$	$60.5 \pm 5$	$48 \pm 3.2$
	(61-78)	(50.8-64.7)	(50-68)	(51-66)	(41-50)
Е%	$234 \pm 22$	$189.5 \pm 27$	$267 \pm 27$	$183.4\pm21.7$	$93\pm8$
	(200-250)	(147-214)	(232-306)	(154-210)	(70-110)
Spicules	$81 \pm 3.6$		$69 \pm 4.6$		
-	(75-88)		(65-72)		
Gubernaculum	41 ± 5		$37.8 \pm 18.6$		
	(37-45)		(30-42)		
SW %	$151 \pm 39$		$152 \pm 10.4$		
	(131-166)		(139-172)		
GS %	$60 \pm 6.3$		$55 \pm 5.2$		
	(51-66)		(44-58.5)		
H%					$39 \pm 0.7$
					(35-44)

Table 1.	Morphometric values of <i>Steinernema affine</i> ( $n = 20$ each). All measurements are in $\mu m$
	and in the form of mean $\pm$ SD (range).

L = Total body length; MBW= Maximum body width; EP= Excretory pore; ES= Distance from anterior end to oesphagus; TL= Tail length; D%= Distance from anterior end to excretory pore divided by pharynx length  $\times$  100; E % = Distance from anterior end to excretory pore divided by tail length  $\times$  100; SW = Spicule length divided by anal body diam.; GS = Gubernaculum length divided by spicule length; H = Hyaline portion  $\times$  100/tail length.



**Fig. 2.** Light micrograph of *Steinernema affine*. Female first generation A-D: A, B. Anterior region; C. Vulval region; D. Tail region; Male first generation: E, F. Tail region (Scale bars: A-D, F= 10  $\mu$ m, E = 20  $\mu$ m).



**Fig. 3.** Phylogenetic relationships of new records of *Steinernema* species, 1 isolate of *S. affinae* and 3 isolates of *S. feltiae* with 46 species of *Steinernema* based on the D2-D3 rDNA sequences from GenBank. *Cervidellus alutus* (AF331911) was used as out group. Numbers at the nodes represent bootstrap proportion for Maximum Parsimony of 50% or more.

## S. *cholashanense* Nguyen, Puza & Mracek, 2008 (Fig. 4. A-F)

## Measurements: Table 2

During the survey of Gilgit-Baltistan, Pakistan a new record *S. cholashanense* Pak.G.S.350 and Pak.G.S.355 were isolated from apple (*Malus pumila*) of village Nasirabad, District Hunza and from walnut (*Juglans nigra*) of Village Danyour, District Gilgit, respectively. These isolates were morphologically and genetically characterized by ITS rDNA region.

**Remarks:** The sequences were deposited in GenBank under accession numbers MF125282 and MF039642. Maximum parsimony of ITS regions shows that the alignment resulted in 1094 characters, 91 are constant, 196 are variable characters are parsimony-uninformative and 807 characters (included) are parsimony-informative. The phylogenetic relationships between 46 species of *Steinernema* are presented in Fig. 5 (for MP, tree length = 4812, Cl = 0.4362, RI = 0.6342, RC = 0.2766, HI = 0.5638). The sequence length composition (Table 3) and pair wise distance have shown in Table 4. *S. cholashanense* was molecularly identified by utilizing ITS rDNA region.

## S. feltiae Filipjev, 1934

Three isolates of *S. feltiae* Pak.G.S.354 (MF150034), Pak.G.S.357 (MF144570) and Pak.G.S.358 (MF158314) were also found during survey of *Prunus armeniaca*, *Prunus persica* and *Malus pumila* from Village Hoper and Thole, District Nager, Gilgit-Baltistan. These isolates were identified through morphologically and molecularly with D2D3 rDNA regions (Table 5).

### References

Anis, M., Shahina, F. & Maqbool, M. A. (2002). Identification of *Steinernema feltiae*  Filipjeve, 1934 (Rhabditida: Steinernematidae) from Pakistan. *Pakistan Journal of Nematology*, 20, 65.

- Bedding, R. A. & Akhurst, R. J. (1975). A simple technique for the detection of insect parasitic rhabditid nematodes in soil. *Nematologica*, 21, 109-110.
- Bovien, P. (1937). Some types of association between nematodes and insects. *Videnskabelige Meddelelser Fra Dansk Naturhistorisk Forening Bd.*, 101, 1-114.
- Courtney, W. D., Polley, D. & Miller, V. I. (1955). TAF an improved fixative in nematode technique. *Plant Disease Reporter*, 39, 570-571.
- Filipjev, I. N. (1934). Miscellania Nematologica
  1. Eine neueart der Gatung Neoaplectana
  Steiner nebst Bermerkungen uber die systematische Stellung der letzteren.
  Magasin de Parasitologie de l'Institut
  Zoologique de l'Académie d'USSR, 4, 229-240.
- Hall, T. A. (1999). BioEdit: A user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. *Nucleic Acids Symposium Series*, 41, 95-98.
- Hazir, S., Keskin, N., Stock, S. P., Kaya,
  H. K. & Özcan, S. (2003). Diversity and distribution of entomopathogenic nematodes (Rhabditida: Steinernematidae and Heterorhabditidae) in Turkey. *Biodiversity & Conservation*, 12, 375-386.
- Hominick, W. M., Reid, A. P. & Briscoe, B. R. (1995). Prevalence and habitat specificity of steinernematid and heterorhabditid nematodes isolated during soil surveys of the UK and the Netherlands. *Journal of Helminthology*, 69, 27-32.
- Hominick, W. M. (2002). Biogeography. In: Gaugler, R. (Ed.). Entomopathogenic Nematology. CABI Publishing, Wallingford, UK, pp. 115-143.
- Joyce, S. A., Reid, A. P., Driver, F. & Curran, J. (1994). Application of polymerase chain reaction (PCR) methods to identification of entomopathogenic nematodes. In: Burnell, A. M., Ehlers, R. U. & Masson, J. P. (Eds.),



**Fig. 4.** Light micrograph of *Steinernema cholashanense*: Female first generation A-D, F: A, B. Anterior region; C. Vulval region; D, F. Tail region; Male first generation: E. Tail region. (Scale: A-D, F=  $10 \ \mu m$ , E =  $20 \ \mu m$ ).

Characters	First Ge	eneration	Second	Generation	Infective
	Male	Female	Male	Female	stage
					juveniles
TL	$1536\pm206$	$4280\pm549$	$830\pm28$	$1982 \pm 195$	$815 \pm 45$
	(1185-2047)	(1302-4975)	(712-950)	(1435-2758)	(700-875)
MBW	$143 \pm 35.$	$235 \pm 25$	$71\pm8.8$	$125 \pm 18$	$26 \pm 1.5$
	(85-178)	(133-262)	(50-89)	(95-156)	(24-30)
EP	$110 \pm 18$	$99 \pm 11$	$59 \pm 5.3$	$86 \pm 5.3$	$65 \pm 1.3$
	(90-125)	(75-125)	(45-72)	(65-105)	(53-68)
NR	$115 \pm 11$	$132 \pm 12$	$94 \pm 6.4$	$112 \pm 8.8$	$79 \pm 2.1$
	(90-130)	(125-150)	(82-104)	(90-142)	(65-94)
ES	$162 \pm 14$	$185 \pm 10$	$119 \pm 5.8$	$176 \pm 12$	$121 \pm 3.6$
	(125-184)	(158-225)	(108-129)	(145-200)	(105-129)
Testis reflection	$450\pm58$		$212 \pm 9.3$		
	(300-680)		(200-256)		
TL (Ijs with sheath)	$37 \pm 4$	$55 \pm 5.3$	$39 \pm 4.1$	$61 \pm 3.8$	$103\pm6.3$
	(32-45)	(45-62)	(27-48)	(45-72)	(95-115)
Anal body width	$45 \pm 5.6$	$73 \pm 12.5$	$40 \pm 3.5$	$40 \pm 1.5$	$16 \pm 0.8$
·	(35-56)	(62-87)	(35-47)	(33-47)	(14-18)
V%		$50 \pm 3.2$		$51 \pm 2.3$	
		(49-52)		(45-56)	
a	$13 \pm 3.6$	$21 \pm 1.2$	$12 \pm 2.1$	$14.3 \pm 1.5$	$26.5\pm1.5$
	(8.2-21)	(20-24.5)	(9-14)	(11-16)	(22-31)
b	$8.3 \pm 1.0$	$19 \pm 1.8$	$5.9 \pm 1.2$	$11.1 \pm 1.4$	$6.3 \pm 0.5$
	(6.5-9.5)	(18-21.3)	(5.5-6.8)	(9-12.5)	(6.2-7.5)
с	$39.7 \pm 4.2$	$63 \pm 9$	$23 \pm 1.8$	$35 \pm 2.4$	$11 \pm 1.0$
	(34-48)	(49-79)	(17-30)	(25-43)	(10-13)
c'	$0.7 \pm 0.1$	$0.7 \pm 0.1$	$1.0 \pm 0.1$	$1.1 \pm 0.1$	$4.1 \pm 0.2$
	(0.5 - 0.85)	(0.6-0.9)	(0.8-1.3)	(1-1.5)	(3.2-4.9)
D %	$67 \pm 8.5$	$51 \pm 7.2$	$49 \pm 3.7$	$48 \pm 3.1$	$47 \pm 2.3$
	(51-79)	(33-60)	(45-57)	(45-56)	(42-51)
E %	$274 \pm 35$	$183 \pm 22$	$187 \pm 18$	$153\pm9.6$	$84 \pm 4.3$
	(193-350)	(125-240)	(130-210)	(130-170)	(75-95)
Spicules	$63 \pm 3.7$		$45 \pm 2.1$		
-	(58-70)		(40-50)		
Gubernaculum	$36 \pm 2.5$		$29 \pm 1.8$		
	30-40		(25-37)		
SW %	$120 \pm 13$		$107 \pm 7.3$		
	(90-140)		(85-130)		
GS %	$69 \pm 4.5$		$53 \pm 2.1$		
	(55-79)		(51-68)		
H%					$35 \pm 4.8$
					(33-49)

Table 2. Morphometric values of *S. cholashanense* (n=25 each). All measurements are in  $\mu$ m and in the form of mean ± SD (range).

L = Total body length; MBW= Maximum body width; EP= Excretory pore; ES= Distance from anterior end to oesphagus; TL= Tail length; D%= Distance from anterior end to excretory pore divided by pharynx length  $\times$  100; E % = Distance from anterior end to excretory pore divided by tail length  $\times$  100; SW = Spicule length divided by anal body diam.; GS = Gubernaculum length divided by spicule length; H = Hyaline portion  $\times$  100/tail length.



**Fig. 5.** Phylogenetic relationships of new record of *S. cholashanense* with 47 species of *Steinernema* based on the ITS- rDNA sequences from GenBank. *Cervidellus elegans* (EU131007) was used as out group. Numbers at the nodes represent bootstrap proportion for Maximum Parsimony of 50% or more.

Species	٨	C	С	т
ITS region	A	C	G	I
S. cholashanense Pak. G. S.350	0.26667	0.13333	0.25000	0.35000
S. cholashanense Pak. G. S.355	0.36667	0.23333	0.13333	0.21667
S. jollieti	0.26667	0.12500	0.25000	0.35833
S. weiseri	0.26250	0.13333	0.25000	0.35833
S. ichnuase	0.26667	0.13750	0.24583	0.35000
S. feltiae	0.26667	0.12917	0.25000	0.34167
S. xueshanense	0.26617	0.12500	0.25000	0.35833
S. litorale	0.26250	0.12917	0.25000	0.35417
S. cholashanense	0.26667	0.12917	0.24167	0.36250
S. kraussei	0.26667	0.14583	0.25417	0.36250
S. oregonense	0.26667	0.12917	0.25000	0.35417
S. taxanum	0.26667	0.12917	0.24167	0.36250
S. thani	0.27500	0.17350	0.24583	0.34167
S. akhursti	0.27500	0.12917	0.24583	0.35000
S. everstense	0.27500	0.12917	0.24583	0.35000
S. kushidai	0.27500	0.13333	0.24583	0.34583
S. monticolum	0.28750	0.12500	0.23750	0.35000
S. ashiuense	0.29583	0.11667	0.22917	0.35833
S. scarabaei	0.26667	0.13750	0.25833	0.33750
S. unicornum	0.30000	0.13750	0.22917	0.33333
S. longicoedum	0.28750	0.14167	0.25417	0.31667
S. karrii	0.28333	0.15000	0.25417	0.31250
S. carpocapsae	0.27917	0.14583	0.25000	0.32500
S. balochiense	0.27500	0.14167	0.24583	0.33750
S. siamkayai	0.28750	0.13750	0.24583	0.32917
S. tami	0.29167	0.15000	0.24167	0.31667

Table 3. Sequence length and composition of ITS and D2D3 regions of species of *Steinernema* species.

т	· · · 1 · · · C ·			. C	C'1. '. D. 1.'.
i wo new re	cords of e	ntomonathoge	nic nematode	s from	Cillett-Balfistan
1		nopanoge			ongre Dannotan

Species		C	G	т
D2D3 region	А	C	G	1
S. feltiae Pak. G. S. 358	0.29848	0.16190	0.32381	0.22381
S. affine Pak. G. S. 356	0.23810	0.17619	0.33810	0.24762
S. feltiae Pak. G. S. 357	0.25238	0.30000	0.21905	0.22857
S. feltiae Pak. G. S. 354	0.24762	0.31429	0.20952	0.22857
S. feltiae	0.24286	0.17619	0.33333	0.24762
S. hebeiense	0.24268	0.17619	0.33333	0.24762
S. oregonense	0.24286	0.17619	0.33810	0.24286
S. cholashanense	0.24286	0.17619	0.33333	0.24762
S. tielingense	0.23810	0.17619	0.33333	0.25238
S. kushidai	0.25238	0.6190	0.32857	0.25714
S. ashiuense	0.23333	0.17143	0.33810	0.25714
S. glesseri	0.23810	0.19524	0.33810	0.22857
S. cubanum	0.23810	0.19048	0.33810	0.23333
S. diaprepsi	0.24286	0.17619	0.33333	0.24762
S. citrai	0.24762	0.17619	0.33333	0.24762
S. scarabei	0.25238	0.16667	0.32857	0.30476
S. unicornum	0.29524	0.14762	0.33810	0.33333
S. affine	0.26667	0.14286	0.30000	0.29048
S. intermedium	0.26667	0.14762	0.30476	0.28095
S. sichuanense	0.27619	0.13810	0.30476	0.28095
S. everstense	0.29524	0.16667	0.20476	0.33333
S. carpocapsae	0.25238	0.14762	0.29524	0.30476
S. siamkayai	0.24286	0.17619	0.33333	0.24762
S. balochiense	0.22857	0.19048	0.34286	0.23810
S. websteri	0.21857	0.16762	0.29524	0.24762
S. rarum	0.23810	0.14286	0.32384	0.2462

<b>S.</b> #	Species	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22
1	S. cholashanense	-																					
	Pak. G. S.350																						
2	S. cholashanense	1/15																					
	Pak. G. S.355	145	-																				
3	S. jollieti	96	2	-																			
4	S. weiseri	96	2	130	-																		
5	S. ichnuase	96	3	130	17	-																	
6	S. feltiae	96	5	132	17	19	-																
7	S. xueshanense	96	2	131	14	16	32	-															
8	S. litorale	96	3	130	15	17	33	19	-														
9	S. cholashanense	95	3	132	13	15	31	17	15	-													
10	S. kraussei	96	9	134	15	20	31	22	18	17	-												
11	S. oregonense	96	1	131	15	17	34	19	17	15	25	-											
12	S. taxanum	95	6	130	16	18	33	22	20	18	27	29	-										
13	S. thani	96	9	134	17	19	35	21	19	17	26	28	19	-									
14	S. akhursti	96	7	131	13	15	29	15	11	9	19	21	12	18	-								
15	S. everstense	95	61	131	13	15	29	15	11	9	19	21	12	18	13	-							
16	S. kushidai	95	10	131	13	14	18	30	18	14	12	22	24	13	21	16	-						
17	S. monticolum	97	10	131	18	21	34	21	19	17	27	29	20	26	22	25	21	-					
18	S. ashiuense	98	13	132	21	21	34	23	21	19	29	31	22	28	25	27	23	22	-				
19	S. scapterisci	98	35	131	35	31	41	38	34	33	39	41	32	36	33	37	36	33	34	-			
20	S. carpocapsae	94	36	132	33	32	44	38	34	35	41	41	34	36	35	40	37	34	34	36	-		
21	S. siamkayai	96	38	129	37	36	48	44	38	39	45	45	40	39	38	45	41	38	39	36	38	-	
22	S. tami	97	38	133	39	38	51	45	40	41	47	45	42	42	41	46	42	39	43	38	40	41	-
23	S. balochiense	98	34	131	34	33	34	40	34	35	42	42	36	36	35	41	32	34	36	33	32	34	35

# Table 4. Pairwise distance of ITs regions of *Steinernema* species.

S. #	Species	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22
1	S. feltiae	-																					
	Pak. G. S. 358																						
2	S. affine	00																					
	Pak. G. S. 356	90	-																				
3	S. feltiae	144	144																				
	Pak. G. S. 357	144	144	-																			
4	S. feltiae	144	141	142																			
	Pak. G. S. 354	144	141	142	-																		
5	S. feltiae	82	2	34	8	-																	
6	S. litorale	128	124	120	144	141	-																
7	S. hebeiense	82	2	34	144	141	0	-															
8	S. oregonense	81	5	34	144	142	4	144	-														
9	S. cholashanense	81	8	31	144	141	6	125	6	-													
10	S. tielingense	82	4	33	144	140	2	121	2	121	-												
11	S. kushidai	82	10	32	145	142	9	124	9	124	9	-											
12	S. ashiuense	85	16	39	143	140	15	126	16	126	15	13	-										
13	S. arenarium	85	20	40	143	140	19	127	19	127	19	20	18	-									
14	S. boemeri	84	19	40	146	143	18	127	18	127	18	17	17	16	-								
15	S. glesseri	86	19	38	142	139	19	127	19	127	19	20	18	19	19	-							
16	S. affine	90	35	00	135	142	34	120	34	120	34	35	34	32	33	30	-						
17	S. intermedium	90	35	2	145	142	34	120	34	120	34	35	34	32	33	30	120	-					
18	S. sichuanense	88	35	4	143	140	34	119	34	119	34	35	34	32	33	30	119	119	-				
19	S. carpocapsae	96	87	90	147	146	88	134	88	134	88	86	88	84	89	86	134	134	87	-			
20	S. siamkayai	84	18	37	145	142	17	125	17	125	17	16	16	17	17	17	125	125	18	21	-		
21	S. balochiense	85	21	37	144	141	21	127	21	127	21	22	20	19	21	19	127	127	16	25	24	-	
22	S. websteri	82	29	37	144	141	28	126	28	126	28	29	26	27	26	28	126	126	32	25	32	29	-
23	S. rarum	81	20	34	143	140	20	125	20	125	21	20	19	18	20	18	125	125	15	24	23	22	22

 Table 5. Pairwise distance of D2 D3 regions of Steinernema species.

COST 812 Biotechnology: Genetics of Entomopathogenic Nematodes-Bacterium Complexes, Proceedings of Symposium & Workshop. St. Patrick's College, Maynooth, Co., Kildare, Ireland. European Commission, DGXII, Luxembourg, pp. 178-187.

- Khatri-Chhetri, H. B., Waeyenberge, L., Spiridonov, S., Manandhar, H. K. & Moens, M. (2011). Two new species of *Steinernema* Travassos, 1927 with short infective juveniles from Nepal. *Russian Journal of Nematology*, 19, 53-74.
- Mracek, Z., Kindlmann, P. & Webster, J. M. (2005a). *Steinernema affine* (Nematoda: Steinernematidae), a new record for North America and its distribution relative to other entomopathogenic nematodes in British Columbia. *Nematology*, 7, 495-501.
- Mracek, Z., Becvar, S., Kindlmann, P. & Jersakova, J. (2005b). Habitat preference for entomopathogenic nematodes, their insect hosts and new faunistic records for the Czech Republic. *Biological Control*, 34, 27-37.
- Nguyen, K. B., Puza, V. & Mracek, Z. (2008). *Steinernema cholashanense* n. sp. (Rhabditida: Steinernematidae) a new species of entomopathogenic nematode from the province of Sichuan, Chola Shan Mountains, China. *Journal of Invertebrate Pathology*, 97, 251-264.
- Nguyen, K. B. & Hunt, D. (2007) Entomopathogenic Nematodes: Systematics, Phylogeny and Bacterial Symbionts. Brill, NV, Leiden, The Netherlands, 816 pp.
- Nguyen, K.B., Malan, A.P. and Gozel, U. (2006) *Steinernema khoisanae* n. sp. (Rhabditida: Steinernematidae), a new entomopathogenic nematode from South Africa. *Nematology*. 8, 157–175.
- Poinar, G. O. (1976). Description and biology of new insect parasitic rhabditoid, *Heterorhabditis bacteriophora* gen. n., sp. n. (Rhabditida: Heterorhabditidae n. fam.). *Nematologica*, 21, 463-470.
- Poinar, Jr, G. O. (1979). Nematodes for biological control of insects. Boca Raton, FL, USA, CRC Press.

- Poinar, G. O. (1988). "Re-description of Neoaplectana affinis Bovien (Rhabditida: Steinernematidae)." Revue de Nematology, 11, 143-147.
- Seinhorst, J. W. (1959). A rapid method for the transfer of nematodes from fixative to anhydrous glycerin. *Nematologica*, 4, 67-69.
- Shahina, F. Firoza, K. & Tabassum, K. A. (2017) Status of Entomopathogenic Nematodes in Integrated Pest management Stretegies in Pakistan. In: *Biocontrol Agents: Entomopathogenic and Slug Parasitic Nematodes*. (Edt.) Elgawad, M. M. M., Askary, T. H. and Coupland, J. Publised by CAB International, CPI Group (UK) Ltd, Croydon, CR0 4 YY. 383-408 pp.
- Sturhan, D. & Liskova, M. (1999). Occurrence and distribution of entomopathogenic nematodes in the Slovak Republic. *Nematology*, 1, 273-277.
- Sturhan, D. & Mracek, Z. (2000). Comparison of the *Galleria* baiting technique and a direct extraction method for recovering *Steinernema* infective juveniles from soil. *Folia Parasitologica*, 47, 315-318.
- Swofford, D. L. (2002). PAUP\* Phylogenetic Analysis Using Parsimony (\*and Other Methods). Sunderland, Massachusetts, Sinauer Associates.
- Tabassum, K. A. & Shahina, F. (2004). Redescription of *Steinernema feltiae* Filipjev
  1934 (Nematoda: Steinernematidae) from Pakistan. *Pakistan Journal of Nematology*, 22, 1-8.
- Tarasco, E., Clausi, M., Rappazzo, G., Panzavolta, T., Curto, G., Sorino, R., Oreste, M., Longo, A., Leone, D., Tiberi, R., Vinciguerra, M. T. & Triggiani, O. (2015). of **Biodiversitv** entomopathogenic nematodes Journal in Italy. of Helminthology, 89, 359-366.
- Thompson, J. D., Gibson, T. J., Plewniak, F., Jeanmougin, F. & Higgins, D. G. (1997).The CLUSTAL\_X windows interface: flexible strategies for multiple sequence alignment aided by quality analysis tools. *Nucleic Acid Research*, 25, 4876-4882.

- White, G. F. (1927). A method for obtaining infective nematode larvae from cultures. *Science*, 66, 302-303.
- Wouts, W. M., Mracek, Z., Gerdin, S. & Bedding, R. A. (1982). *Neoaplectana* Steiner, 1929 a junior synonym of

*Steinernema* Travassos, (1927). (Nematoda: Rhabditida). *Systematic Parasitology*, 4, 147-154.

Zuccon, A. & Zuccon, D. (2010). *MrEnt v.2.1*. Program distributed by the authors, available online at: http://www.mrent.org.

(Received: 10 March, 2017)