## **Short Communication**

## Community analysis of phytoparasitic nematodes associated with ornamental plants at Jimma University Agriculture Campus, Ethiopia

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Under the hypothesis, this work was carried out to determine that a complex of plant-parasitic nematode groups are involved in the growth suppression and death of the ornamental plants and hence establishing records of their type and abundance is necessary to minimize the risk to future plantings.

To assess the incidence of plant-parasitic nematodes, soil samples were collected in December 2016, from three locations of Agriculture Campus of Jimma University located at 356 km Southwest of Addis Ababa. Ethiopia, at 7°68'N latitude and 36°83'E longitude with an altitude of 1752 m.a.s.l. The annual temperature ranges from 11.8-26.8°C with a relative humidity and mean rainfall of 91% and 1500mm, respectively. Collections were made from the rhizosphere of eight species of ornamental plants viz., Rosa spp., Colocasia esculenta, Salvia splendens, Ctenanthe oppenheimiana, Phoenix abyssinica, Aster Washingtonia rubusta and novea-angliae. Doronicum grandiflorum growing along the roadsides in the campus showing severe infestation of phytoparasitic nematodes.

Following the procedure of Baermann modified techniques (Hooper *et al.*, 2005) nematodes were extracted and identified (Siddiqi, 2001) up to genus level and enumerated from a 1 ml capacity counting dish using a Leica compound microscope.

A total of ten genera of plant parasitic nematodes (Helicotylenchus, Hemicycliophora, Meloidogyne, Pratylenchus, Scutellonema, Paratrichodorus, Rotylenchulus, Trichodorus, Mesocriconema and Tylenchorhynchus) were recorded associated with these ornamental plant species. Helicotylenchus and Scutellonema were found associated with all ornamental plants followed by *Meloidogyne* that was not found in the rhizophere of A. novea-angliae. On the other hand, S. splendens harbored eight of the detected Rotylenchulus genera other than and Pratylenchus. Moreover, Helicotylenchus and Meloidogyne were the most frequently encountered nematode genera with relatively high population densities.

Total mean population densities (PD), frequency of occurrence, prominence value of each

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nematode genus from all ornamental species were determined based on Norton (1978). Prominence value (PV) was calculated as PV= PD\*(FO-1/2)\*10-1(De Waele *et al.*, 1998). The frequency of occurrence (FO), mean population density (PD) and prominence value (PV) of the afore mentioned phytoparasitic genera is shown (Table 1). This result revealed that density and occurrence of each nematode PPN genera varied with host species. The highest mean population density recorded for *Meloidogyne* was 130 J2in *S. splendes*, followed by 73 J2 in *P. abyssinica*.

The host species, C. oppenheimiana was found to be the least host for *Meloidogyne* with a mean population density of 13 J2. Helicotylenchus became the most abundant among all genera recorded with mean population density of 380 nematodes recorded on D. grandiflorum while the least host for this genus was C. esculenta with a population density of 13 nematodes per100 ml soil. D. grandiflorum and S. splendes appeared to be the best host for Scutellonema which was detected with a mean population density of 183 and 180 nematodes 100 ml-1 soil, respectively. P. abyssinica hosted the lowest population of Scutellonema i.e. 7 nematodes ml<sup>-1</sup>soil. The mean population density of rest of the PPN genera ranged between 7 and 27 nematodes 100 ml<sup>-1</sup>soil.

Trichodorus, Rotylenchulus and Pratylenchus that were only found in a single sampling site of their respective hosts had generally low mean population density of 17, 13 and 7 nematodes, respectively. Rosa spp., S. splendes and C. oppenheimiana equally hosted Mesocriconema (10 nematodes  $100 \text{ml}^{-1}$  soil) while C. esculenta hosted the lowest mean density (7)nematodes100ml<sup>-1</sup>soil). P. abyssinica hosted Tylenchorhynchus better than D. grandiflorum and S. splendes with a mean density of 27 nematodes. Similar to their density, the frequency of occurrence of each PPN genera varied among host species (Table 1). Helicotylenchus was detected (FO=100%) in all examined perennial ornamental crops except on Rosa spp. and C. esculenta (33%). The highest 112

frequency of occurrence (100%) of *Meloidogyne* was recorded on Rosa spp., C. esculenta, and S. splendes followed by (67%) in W. robusta while the least (33%) recorded in D. grandiflorum, C. oppenheimiana and P. abyssinica plant species. The highest frequency of occurrence of Scutellonema (100%) was found on C. esculenta followed by D. grandiflorum and S. splendes (67%) and occurred uniformly across the rest of the plant species (FO= 33%). *Tylenchorhynchus* was detected from three sampling sites (100%) associated with P. abyssinica 67% was associated with S. splendens and with D. grandiflorum (33%). The rest of nematode (Hemicyclophora, genera Mesocriconema, Pratylenchus, Rotylenchulus, Paratrichodorus and Trichodorus) were all with least frequency of occurrence (33%) in all plant species.

Helicotylenchus was most prominent the nematode (PV =1,202) followed by Scutellonema (PV =473) on D. grandiflorum. Meloidogyne was prominent on S. splendens (PV=411). The other nematode genera such as Hemicycliophora, Mesocriconema, Paratrichodorus, Trichodorus and Pratylenchus were less prominent in all the plant species. It has already been reported that Salvia cultivars are as suitable host for root-knot nematodes (Goff, 1936). Four cultivars of S. splendens were evaluated for their responses to an isolate of Meloidogyne incognita and M. javanica showed higher nematode population on all cultivars (McSorley & Frederick, 2001). W. robusta was reported to support the reproduction of Rotylenchulus reniformis (Inserra et al., 1994), in the present study this nematode was only recorded from C. esculenta known to parasitize impact the production. it and Higher susceptibility of Washingtonia spp. to M. javanica is also documented (Brito et al., 2010).

The root-knot nematodes are well known to infect nearly all plant species causing root galls (Moens *et al.*, 2009). However, the presence of differential hosts indicates that a host known to be susceptible to one *Meloidogyne* species could be resistance to the same species (Hartman &

Ornamental plant species	Nematode genera	PD	FO (%)**	<b>PV</b> ***
Rosa spp.	Meloidogyne	48	100	151
	Helicotylenchus	57	33	104
	Scutellonema	13	33	24
	Hemicycliophora	7	33	12
	Mesocriconema	10	33	18
Colocasia esculenta	Meloidogyne	60	100	190
	Scutellonema	67	100	211
	Rotylenchulus	13	33	24
	Mesocriconema	7	33	12
	Helicotylenchus	13	33	24
Salvia splendens	Scutellonema	180	67	465
	Hemicycliophora	13	33	24
	Tylenchorhynchus	20	67	52
	Meloidogyne	130	100	411
	Helicotylenchus	187	100	590
	Paratrichodorus	23	33	43
	Trichodorus	17	33	30
	Mesocriconema	10	33	18
Ctenanthe oppenheimiana	Helicotylenchus	173	100	548
	Scutellonema	60	33	110
	Meloidogyne	13	33	24
	Mesocriconema	10	33	18
	Paratrichodorus	7	33	12
Phoenix abyssinica	Helicotylenchus	43	100	137
	Meloidogyne	73	33	134
	Scutellonema	7	33	12
	Tylenchorhynchus	27	100	84
Aster novae-angliae	Helicotylenchus	70	100	221
	Scutellonema	27	33	49
Washingtonia robusta	Helicotylenchus	133	100	422
	Meloidogyne	57	67	146
	Pratylenchus	7	33	12
	Scutellonema	40	33	73
Doronicum grandiflorum	Helicotylenchus	380	100	1202
	Meloidogyne	23	33	43
	Scutellonema	183	67	473
	Tylenchorhynchus	10	33	18

Table 1. Frequency of occurrence and abundance in 100 of ml soil and prominence value of major
nematode genera from the soil rhizosphere of ornamental plant species at the Agriculture
College Campus of Jimma University, Ethiopia

\*Population density = mean number of individual nematodes genus per 100 ml soil/number of sampling sites (n=3) \*Frequency of occurrence (FO%) = number of sites with positive detected/total number of sites sampled\*100. \*\*\*\*Prominence Value (PV) = Mean population density  $(FO)^{1/2} *10^{-1}$ .

Sasser,1985). In this respect, the fact *Meloidogyne* was not recovered from *A.novae-angliae* could be ascribed to the chemical characteristics of the family Asteraceae so is does this species (Furtado *et al.*, 2011).

Except *Meloidogyne*, *Helicotylenchus* and *Hemicycliophora* which previously reported from *Rosa* species (Meressa *et al.*, 2014), no host-nematode relationship of these plants species along with their respective associated nematode genera has been reported from Ethiopia. The fact that this study was conducted for the first time, it is difficult to associate a particular nematode genus to the frequent death of these ornamental plants in the campus.

However, in Brazil, Pratylenchus, Rotylenchulus, Meloidogyne and Helicotylenchus were reported to be responsible for serious injuries to most ornamental plants species reducing their ornamental properties (Gimenses et al., 2010). The loss of beauty caused by Mesocriconema, Helicotylenchus, and Pratylenchus damage on various ornamental species was also reported in Tehran city (Saeedizadeh, 2016). Similarly, a greenhouse study in the city of Kyiv (Sigarovova & Karplivk, 2015) indicated that Tylenchorhynchus, Pratylenchus, Rotylenchus and Helicotylenchus can cause a severe damage to landscape plants reducing their economic value. Recently, there have been deaths of some trees of *P. abyssinica* in the sampling area.

A study by Mani *et al.*, (2005) reported *Helicotylenchus diagonicus* and *Meloidogyne javanica* as important parasitic nematode species of date palm trees in Sultanate of Oman. The fact that these two species were recorded in the campus may signify their potential to kill the plant.

*Doronicum* spp. are known to be infected by different species of *Aphelenchoides* (Kohl, 2011), although their economic damage is not known. Thus, this study adds four new host-parasite relationships that need exploration for 114

extent of damage that can be caused by those associated nematode genera.

The present study revealed the presence of various economically important phytoparasitic associated with different ornamental plant species that could be vital for future management plan. Meloidogyne, Helicotylenchus and Scutellonema can be considered more potential constraints for the establishment of most of the ornamental plants at the campus. Nevertheless, it is paramount important that their population dynamics and damage thresholds to the host plants is further studied. The frequent occurrence of the other genera in less population levels probably indicates that they are likely to parasitize the ornamentals but cause less damage.

## Acknowledgments

This work was supported by Jimma University. The authors are also thankful to Mr Nasir A. for his assistance during identification of the ornamental plants.

## References

- Brito, J. A., Kaur, R., Cetintas, R., Stanley, J. D., Mendes, M. L., Powers, T. O. & Dickson, D. W. (2010). *Meloidogyne* spp. infecting ornamental plants in Florida. *Nematropica*, 40, 87-103.
- De Waele, D., McDonald, A. H., Jordaan, E. M., Orion, D., Van Den Berg, E. & Loots, G. C. (1998). Plant-parasitic nematodes associated with maize and pearl millet in Namibia. *African Plant Protection*, 4, 113-117.
- Furtado, S. K., Silva, A. L. P., Miguel, O. G., Dias, J. F. G., Miguel, M. D., Costa, S. S. & Raquel, N. R. B. (2011). Effectiveness of Asteraceae extracts on Trichostrongylidae eggs development in sheep. *Revista Brasileira de Parasitologia Veterinária*, 20, 215-218.
- Gimenses, R., Batista, G. S., Pivetta, K. F. L., Santos, J. M., Soares, P. L. M. & Martins, T. A. (2010). Occurrence of plant-parasitic

nematodes in ornamental and flowering plants at UNESP/FCAV, Campus of Jaboticabal, São Paulo State, Brazil. *Acta Horticulturae*, 881, 607-610.

- Goff, C. C. (1936). Relative susceptibility of some annual ornamentals to root- knot. Univ. of Florida Agriculture Extension Station Buellton. 291.
- Hartman, K. M., & Sasser, J. N. (1985). Identification of *Meloidogyne* species on the basis of differential host test and perineal pattem morphology. In: *An Advanced Treatise on Meloidogyne*, *Vol. 2, Methodology*. Ed. by.K. R. Barker, C. C. Carter, & J. N. Sasser, North Carolina State Univ. Graphics, Raleigh.
- Hooper, D., Hallmann, J. & Subbotin, S. (2005). Extraction, processing and detection of plant and soil nematodes. In: *Plant parasitic nematodes in subtropical and tropical agriculture*. Ed. by Luc, M., Sikora, R. A., Bridge, J. CABI Publishing, Wallingford, UK, pp. 53-86.
- Inserra RN, Duncan LW, O'Bannon JH, Fuller SA. (1994). Citrus nematode biotypes and resistant citrus rootstocks in Florida. *Nematology Circular No. 205.* Florida Department of Agriculture and Consumer Services. Division of Plant Industry.
- Kohl, L. M. (2011). Foliar nematodes: A summary of biology and control with a compilation of host range. Plant Health Progress doi:10.1094/PHP-2011-1129-01-RV.
- Mani, A., Handoo, Z. A. & Livingston, S. (2005). Plant-parasitic nematodes associated with

date palm trees (*Phoenix dactylifera* L.) in the sultanate of Oman. *Nematropica*, 35, 135-143.

- McSorley, R. & Frederick, J. J. (2001). Host suitability of some *Vinca* and *Salvia* cultivars to two isolates of root-knot nematodes. *Proceedings of the Florida State Horticultural Society*. 114, 239-241.
- Meressa, B. H., Dehne, H. W. & Hallmann, J. (2014). Plant parasitic nematodes of cutflowers in Ethiopia. *International Journal of Nematology*, 24, 1-10.
- Moens, M., Perry, R. N. & Starr, J. L. (2009). *Meloidogyne* species- a diverse group of novel and important plant parasites. In: *Root-knot nematodes*. Ed. by Perry, R. N. Moens, M., & Starr, J. L. CABI International. Wallingford. UK, CABI Publishing, pp. 1-17.
- Norton, D. C. (1978). *Ecology of plant parasitic nematodes*. John Wiley, New York. pp.266.
- Saeedizadeh, A. (2016). Identification and distribution of plant-parasitic nematodes in landscape of Tehran City, Iran. *Iranian Journal of Plant Protection Science*, 47, 43-49.
- Siddiqi, M. R. (2000). Tylenchida: parasites of plants and insects 2<sup>nd</sup> Edition. CABI Publishing, Wallingford, UK, pp. 833.
- Sigarovova, D. D. & Karpliyk, V. G. (2015). Parasitic nematodes of flowering and ornamental plants: effect of parasite on the plants and response of the plants to the presence of nematodes. *Vestnik Zoology*, 49, 427-432.