

# Prevalence and Public Health Significance of Anisakis Larvae in some Marketed Marine Fish in Egypt

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**Abstract** | This study investigated the prevalence rate of Anisakis larvae in two marine water fish commonly consumed in Egypt, namely, herrings and sardine. Samples were collected equally from Suez, Ismailia, Damietta, Port Said and Alexandria. The obtained results revealed overall incidence rates of Anisakis larvae in herrings and sardine at 70% and 50%, respectively. The parasite infested mainly (100%) the visceral organs of the positive samples in the two fish species; while infested the muscles in 30% and 10% of herrings and sardine, respectively. The highest prevalence rates for the two species were recorded in the collected samples from Damietta, followed by Alexandria, Port Said, Ismailia, and Suez, respectively. The public health significance of Anisakis larvae was further discussed.

Keywords | Herrings; Sardine; Anisakis larvae; Egypt

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### **INTRODUCTION**

Fish is considered as a major source for high quality protein, essential amino acids, polyunsaturated fatty acids, minerals, and vitamins (El-Ghareeb et al., 2021; Morshdy et al., 2013, 2019). However, fish is considered as a potential source for foodborne pathogens including bacteria, molds, and parasites. The World Health Organization (WHO, 2012) stated that about 56 million cases of human infection with parasitic diseases are associated with the consumption of fish.

Anisakids are nematodes that associated with a parasitic zoonotic disease named anisakiasis which occurs due to

ingestion of the third larval stage of *Anisakis spp*. This parasite infests mainly marine fish and parasitizes crustaceans, cephalopods, and fish as intermediate hosts (Nieuwenhuizen and Lopata, 2013). The disease is caused mainly by *Anisakis simplex, Anisakis pegreffii*, and *Anisakis physeteris* (Mattiucci and Nascetti, 2008).

Humans acquire anisakis infestation via ingestion of inadequately cooked fish or raw fish such as sushi and sashimi (famous dishes in Japan) (Pampiglione et al., 2002). In addition, several anisakid antigens are thermostable and therefore, they constitute a major health concern (EFSA, 2010; Caraballo et al., 2011). Human anisakiasis is characterized by acute abdominal pain, nausea, vomiting, and

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the disease might progress to develop peptic and duodenal ulcers, appendicitis, and peritonitis. Hypersensitivity, urticaria, and anaphylaxis are also among the critical symptoms in some highly susceptible people (Villazanakretzer et al., 2016).

The third Anisakis larval stage is observed in many fish species worldwide. Herrings are among the important fish species in Egypt and Mediterranean countries and play important roles in the economy and fish trading in those countries. Herrings are sold in Egypt either as raw herrings or as smoked ones. The latter is very popular in Egypt because of its specific aroma and flavor. However, herrings are also considered as among the important hosts for *Anisakis spp.* (Bao et al., 2017; Guardone et al., 2017). *Sardine spp.* is another important fish species of high nutritive value that is very common in the fish menu in Egypt. At the same time, this species is among the natural hosts for *Anisakis spp.* (Debenedetti et al., 2019).

Studying the prevalence of anisakis larvae in fish in Egypt, particularly among the marketed herrings and sardine has received less attention. In sight of the previous facts, this study aimed at investigation the prevalence rates of *Anisakis spp.* in the retailed raw herrings and sardines. Discussion of the public health significance of this nematode was followed.

### MATERIAL AND METHODS

#### **COLLECTION OF SAMPLES**

A total of 100 random whole fish, including 50 fish from each of herrings (*Clupea harengus Linnaeus*), and sardine (*Sardinella aurita*). Fish samples were collected equally from fish markets (n = 20 fish/from each locality) in Suez, Ismailia, Port Said, Damietta, and Alexandria, Egypt on a daily basis in the morning and within 2 h from the arrival of the fish into the market. Fish samples were intact, with fresh smell, and reddish gills. Samples were transferred cooled directly without delay to the Laboratory of Food Hygiene, Faculty of Veterinary Medicine, Zagazig University, Egypt for parasitological examination.

#### **FISH EXAMINATION**

Fish samples were dissected and analyzed for the presence of Anisakid larvae by careful inspection of the fish viscera under a stereoscopic microscope. Fish muscles were exposed to an artificial enzymatic digestion according to Llarena-Reino et al. (2013) and the resultant product was examined under a stereoscopic microscope.

#### MORPHOLOGICAL IDENTIFICATION

All the detected anisakids were identified according to the morphological characters described before (Gibbons,

2010). The main criteria considered for anisakids classifications were the position of the excretory pore, the shape of the tail, the arrangement and separation of the digestive tract into esophagus, ventricle and the presence/absence of structures such as intestinal caeca and esophageal appendix.

#### MOLECULAR IDENTIFICATION

**DNA extraction, quantification, and quality assessment:** Total DNA extraction and quantitative estimation of DNA from 1 to 4 larvae per sample was performed according to Guardone et al. (2016). In addition, DNA integrity was further evaluated according to Giusti et al. (2019).

Amplification of the mitochondrial cytochrome c oxidase subunit II gene: Amplification of a 629 bp fragment of the mitochondrial cytochrome c oxidase subunit II (cox2) gene was performed as described in Guardone et al. (2018). The cycling conditions used for the amplification procedures started with an initial denaturation at 95 °C for 3 min, followed by 45 cycles at 95 °C for 20 s, 52 °C for 20 s, 72 °C for 25 s, and a final extension cycle at 72 °C for 10 min was followed.

#### STATISTICAL ANALYSIS

The prevalence and the distribution frequencies were analyzed according to Bush et al. (1997) for the total anisakid larvae in relation to the place of infestation either the muscle or the viscera and to the origin of the sample collection.

### RESULTS

The obtained results in the present study revealed detection of Anisakis larvae in herrings and sardine with overall prevalence rates of 70% and 50%, respectively. Anisakis larvae were detected at 70% and 50% in the viscera of the two species, respectively. The larvae infested the muscles at lower rates 30% (herrings), and 10% (sardine), respectively (Figure 1).

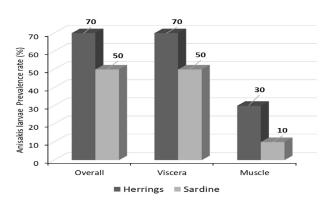


Figure 1: Prevalence rates (%) of Anisakis larvae in the examined herrings and sardine

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The results recorded in Figure 2 showed that fish sampled at Damietta had had the highest infestation rate with Anisakis larvae, while that sampled at Suez had the lowest infestation rate.

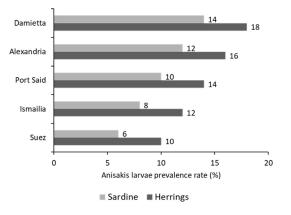


Figure 2: Prevalence rates (%) of Anisakis larvae in the collected herrings and sardine from different localities in Egypt

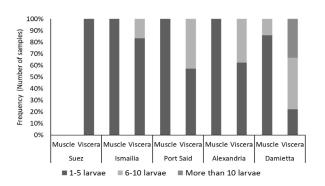
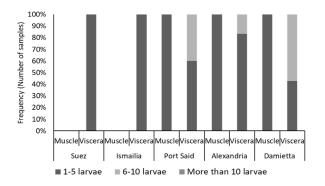


Figure 3: Frequency distribution of Anisakis larvae from viscera and muscle of herrings collected from different localities in Egypt



**Figure 4:** Frequency distribution of Anisakis larvae from viscera and muscle of sardine collected from different localities in Egypt

The presented data in Figure 3 showed detection of more than 10 anisakis larvae in the viscera of 30% of the her-

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rings collected from Damietta representing a high-risk group, while 6 to 10 larvae (intermediate-risk group) were detected at 0%, 10%, 30%, 30%, and 40% in the viscera of the herrings collected from Suez, Ismailia, Port Said, Al-exandria, and Damietta, respectively. Sardine located at the intermediate-risk group as Anisakis larvae was detected at only 10%, 20%, and 40% in the viscera of sardine collected from Alexandria, Port Said, and Damietta (Figure 4).

#### DISCUSSION

Herrings and sardine are among the most important fish species in the Egyptian markets. These fish kinds are used as fried or grilled or pass into further manufacture and processing steps to make new types of fish products such as salted sardine, or smoked herrings. The obtained results in Figure 1 revealed that herrings had higher infestation rate with Anisakis larvae compared with sardine. Anisakis larvae infested the viscera at higher rates compared with the muscles in the two fish species examined. In agreement with the obtained results in the current study, Unger et al. (2014) detected Anisakis larvae in the viscera of the herrings collected from the Baltic-sea at 100%. Bao et al. (2017) detected Anisakis larvae in the visceral organs of the herrings collected from the North Sea at 76%. Similarly, Levsen et al. (2018) detected the parasite in the body cavity of the herrings collected from the North Sea at 81.2% and in the muscles at 17.4%. In addition, Guardone et al. (2019) detected Anisakis larvae at 41.5% from the marketed herrings and herrings; products in Italy. They further reported that visceral organs were the most favorable site for the parasite where, they can detect the larvae at 98% from the positive samples. Anisakis larvae were also detected in sardine collected from Spain at 6.76% (Debenedetti et al., 2019). Generally, sardine is considered as a fish with low risk of anisakiosis because of the low prevalence rate (Cavallero et al., 2015; Gutiérrez-Galindo et al., 2010).

The prevalence of Anisakis larvae in the two examined fish species varied according to the place of the collection. As the highest prevalence rates of the parasite from the two fish species came in the following order: Damietta > Alexandria > Port Said (Mediterranean Sea source) > Ismailia (Suez Canal source) > Suez (Red Sea source) (Figure 2). In agreement with this observation, Debenedetti et al. (2019) observed a clear variation in the prevalence rates of Anisakis larvae in different fish species (hake, mullet, sardine, mackerel, and anchovy) collected from the Atlantic Ocean than same fish species collected from the Mediterranean Sea. Therefore, the origin of the fish should be considered as a critical factor in order to reduce the infection risk by this zoonosis. This assumption agrees with Abollo et al. (2001), Silva and Eiras (2003), and Herrador et al. (2019). Higher infestation rates were observed in the viscera com-

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pared with the muscles, particularly the viscera of herrings which represented a high-risk group (Figure 3, 4). Unlikely, there was no high-risk group in sardine, i.e., more than 10 larvae per fish (Debenedetti et al., 2019). The obtained result in the present study agrees with the report of Debenedetti et al. (2019) who demonstrated a clear variation in the frequency of distribution of anisakis larvae in different fish species including anchovy, hake, sardine, mullet, and mackerel. In all data analyzed, the high prevalence of the larvae in the viscera compared with the flesh was observed. People usually consume the fish flesh than the viscera; however cross contamination of the muscle can take place during any step of fish preparation starting from eviscera to the flesh (Abollo et al., 2001).

Parasitic infestation of the fish with anisakis larvae has several adverse health effects on the fish starting from inflammation during larval penetration to the different visceral organs with significant reduction in the physiological functions of such organs. In addition, the larvae can excrete some chemical compounds such as pentanols and pentanones which have local anaesthetic effects on the fish muscles and therefore affecting the swimming ability of the fish making them easy targets for their predatory fishes (Buchmann and Mehrdana, 2016; Haarder et al., 2013; Rohlwing et al., 1998).

Consumption of fish contaminated with anisakis larvae might have several adverse health effects including abdominal anisakiasis in the humans due to penetration of the gastric or abdominal mucosa (Audicana et al., 1997; Di Azevedo et al., 2017). In addition, allergic reactions that might reach to anaphylaxes might occur, particularly in the high-risk groups including children, elderly, and debilitating patients (Song et al., 2019; Pozio, 2013).

Among the effective strategies to prevent anisakis infection are to prevent their migration postmortem from viscera to flesh by visual inspection by the fishermen or the consumers and collecting the larvae manually, avoid consumption of undercooked or raw fish, efficient cooking of the fish by allowing an internal temperature of 60°C for 1-3 minutes, adequate freezing of the fish for 24 hours at -20°C, and pickling of the fish in vinegar and salt is also considered as a suitable method for reducing the hazards of the anisakis larvae (EC, 2011).

In conclusion, the obtained results in the present study revealed detection of anisakis larvae in both herrings and sardine collected from different locations in Egypt at variable percentages. Herrings had higher prevalence rates compared with the sardine. The nematodes mainly infested the viscera than the muscle. Therefore, efficient cooking of the fish is recommended as a standard preventive strategy to reduce the risk of human anisakiasis in Egypt.

# **CONFLICT OF INTEREST**

None.

# **AUTHORS CONTRIBUTION**

All authors contributed equally.

### REFERENCES

- Abollo E, Gestal C, Pascual S (2001). *Anisakis* infestation in marine fish and cephalopods from Galician waters: An updated perspective. Parasitol. Res. 87: 492-499. https://doi.org/10.1007/s004360100389
- Audicana L, Audicana MT, Fernández de Corres L, Kennedy MW (1997). Cooking and freezing may not protect against allergenic reactions to ingested Anisakis simplex antigens in humans. Vet. Rec. 140: 235. https://doi.org/10.1136/ vr.140.9.235
- Bao M, Pierce G, Pascual S, Gonzalez-Munoz M, Mattiucci S, Mladineo I, Cipriani P, Buselic I, Strachan NJC (2017). Assessing the risk of an emerging zoonosis of worldwide concern: anisakiasis. Sci. Rep. 7. https://doi.org/10.1038/ srep43699
- Buchmann K, Mehrdana F (2016). Effects of anisakid nematodes Anisakis simplex (s.l.), Pseudoterranova decipiens (s.l.) and Contracaecum osculatum (s.l.) on fish and consumer health. Food Waterborne Parasitol. 4: 13-22. https://doi. org/10.1016/j.fawpar.2016.07.003
- Bush AO, Lafferty KD, Lotz JM, Shostak AW (1997). Parasitology meets ecology on its own terms: Margolis et al. revisited. J. Parasitol. 83: 575-583. https://doi.org/10.2307/3284227
- Caballero ML, Umpierrez A, Moneo I, Rodriguez-Perez R (2011). Anis 10, a new Anisakis simplex allergen: cloning and heterologous expression. Parasitol. Int. 60: 209-212. https://doi.org/10.1016/j.parint.2011.01.003
- Cavallero S, Magnabosco C, Civettini M, Boffo L, Mingarelli G, Buratti P, Giovanardi O, Fortuna CM, Arcangeli G (2015). Survey of *Anisakis* sp. and *Hysterothylacium* sp. in sardines and anchovies from the North Adriatic Sea. Int. J. Food. Microbiol. 200: 18-21. https://doi.org/10.1016/j. ijfoodmicro.2015.01.017
- Debenedetti ÁL, Madrid E, Trelis M, Codes FJ, Gil-Gómez F, Sáez-Durán S, Fuentes MV (2019). Prevalence and risk of anisakid larvae in fresh fish frequently consumed in Spain: An Overview. Fishes 4(1): 13. https://doi.org/10.3390/ fishes4010013
- Di Azevedo MIN, Carvalho VL, Iñiguez AM (2017). Integrative taxonomy of anisakid nematodes in stranded cetaceans from Brazilian waters: an update on parasite's hosts and geographical records. Parasitol. Res. 116: 3105-3116. https:// doi.org/10.1007/s00436-017-5622-8
- European Commission (2011). Commission Regulation (EU) No 1276/2011 of 8 December 2011 amending Annex III to Regulation (EC) No 853/2004 of the European Parliament and of the Council as regards the treatment to kill viable parasites in fishery products for human consumption. Off. J.

#### Advances in Animal and Veterinary Sciences

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Eur. Union L 327: 39-41.

- European Food Safety Association (EFSA) (2010). Scientific opinion on risk assessment of parasites in fishery products. EFSA J 8: 1543. https://doi.org/10.2903/j.efsa.2010.1543
- El-Ghareeb WR, Elhelaly AE, Abdallah KME, El-Sherbiny HM, Darwish WS (2021). Formation of biogenic amines in fish: dietary intakes and health risk assessment. Food Sci. Nutr. 9(6): 3123-3129. https://doi.org/10.1002/fsn3.2271
- Gibbons LM (2010). Keys to the Nematode Parasites of Vertebrates, Supplementari Volume; CAB International: Wallingford, England, 2010; p. 416. https://doi. org/10.1079/9781845935719.0000
- Giusti A, Tinacci L, Sotelo CG, Acutis PL, Ielasi N, Armani A (2019). Authentication of ready-to-eat anchovy products sold on the Italian market by BLAST analysis of a highly informative cytochrome b gene fragment. Food Control 97: 50-57. https://doi.org/10.1016/j.foodcont.2018.10.018
- Guardone L, Malandra R, Costanzo F, Castigliego L, Tinacci L,
  Gianfaldoni D, Guidi A, Armani A (2016). Assessment of
  a sampling plan based on visual inspection for the detection
  of anisakid larvae in fresh anchovies (*Engraulis encrasicolus*).
  A first step towards official validation? Food Anal. Methods
  9: 1418-1427. https://doi.org/10.1007/s12161-015-0316-2
- Guardone L, Nucera D, Lodola LB, Tinacci L, Acutis PL, Guidi A, Armani A (2018). Anisakis spp. larvae in different kinds of ready to eat products made of anchovies (*Engraulis encrasicolus*) sold in Italian supermarkets. Int. J. Food Microbiol. 268: 10-18. https://doi.org/10.1016/j. ijfoodmicro.2017.12.030
- Guardone L, Nucera D, Pergola V, Costanzo F, Costa E, Guidi A, Armani A (2017). A rapid digestion method for the detection of anisakid larvae in European anchovy (*Engraulis encrasicolus*): Visceral larvae as a predictive index of the overall level of fish batch infestation and marketability. Int. J. Food Microbiol. 250: 12-18. https://doi.org/10.1016/j. ijfoodmicro.2017.03.011
- Guardone L, Nucera D, Rosellini N, Tinacci L, Acutis PL, Guidi A, Armani A (2019) Occurrence, distribution and viability of Anisakis spp. larvae in various kind of marketed herring products in Italy. Food Control 101: 126-33. https://doi. org/10.1016/j.foodcont.2019.02.030
- Gutiérrez-Galindo JF, Osanz-Mur AC, Moraventura MT (2010). Occurrence and infection dynamics of anisakid larvae in Scomber scombrus, Trachurus trachurus, Sardina pilchardus, and Engraulis encrasicolus from Tarragona (NE Spain). Food Control 21: 1550-1555. https://doi.org/10.1016/j. foodcont.2010.03.019
- Haarder S, Kania PW, Bahlool QZM, Buchmann K (2013). Expression of immune relevant genes in rainbow trout following exposure to live Anisakis simplex larvae. Exp. Parasitol. 135: 564-569. https://doi.org/10.1016/j. exppara.2013.09.011
- Herrador Z, Daschner A, Perteguer MJ, Benito A (2019). Epidemiological scenario of anisakidosis in Spain based on associated hospitalizations: The tipping point of the iceberg. Clin. Inf. Dis. 69(1): 69-76. https://doi.org/10.1093/cid/ ciy853
- Levsen A, Svanevik CS, Cipriani P, Mattiucci S, Gay M,

Hastie LC (2018). A survey of zoonotic nematodes of commercial key fish species from major European fishing grounds—introducing the FP7 PARASITE exposure assessment study. Fisheries Res. 202: 4-21

- Llarena-Reino M, Piñeiro C, Antonio J, Outeriño L, Vello C, González AF, Pascual S (2013). Optimization of the pepsin digestion method for anisakids inspection in the fishing industry. Vet. Parasitol. 191: 276-283. https://doi. org/10.1016/j.vetpar.2012.09.015
- Mattiucci S, Nascetti G (2008). Advances and trends in the molecular systematics of anisakid nematodes, with implications for their evolutionary ecology and host parasite co-evolutionary processes. Adv. Parasitol. 66: 47-148. https://doi.org/10.1016/S0065-308X(08)00202-9
- Morshdy AE, Hafez AE, Darwish WS, Hussein MA, Tharwat AE (2013). Heavy metal residues in canned fishes in Egypt. Jpn. J. Vet. Res. 61: S54-S57.
- Morshdy AEMA, Darwish WS, Daoud JRM, Hussein MAM, Sebak MAM (2019). Monitoring of organochlorine pesticide residues in *Oreochromis niloticus* collected from some localities in Egypt. Slov. Vet. Res. 55: 303-311.
- Nieuwenhuizen N, Lopata A (2013). Anisakis a foodborne parasite that triggers allergic host defences. Int. J. Parasitol. 43(12-13): 1047-57. https://doi.org/10.1016/j. ijpara.2013.08.001
- Pampiglione S, Rivasi F, Criscuolo M, De Benedittis A, Gentile A, Russo S, Testini M, Villan M (2002). Human anisakiasis in Italy: a report of eleven new cases. Pathol. Res. Pract. 198: 429-434. https://doi.org/10.1078/0344-0338-00277
- Pozio E (2013). Integrating animal health surveillance and food safety: the example of Anisakis. Rev. Sci. Tech. 32: 487-496 https://doi.org/10.20506/rst.32.2.2246.
- Rohlwing T, Palm HW, Rosenthal H (1998). Parasitation with Pseudoterranova decipiens (Nematoda) influences the survival rate of the European smelt Osmerus eperlanus retained by a screen wall of a nuclear power plant. Dis. Aquat. Org. 32: 233-236. https://doi.org/10.3354/dao032233
- Silva M, Eiras J (2003). Occurrence of *Anisakis* sp. in fishes off the Portuguese West coast and evaluation of its zoonotic potential. Bull. Eur. Assoc. Fish Pathol. 23: 13-17.
- Song H, Jung B-K, Cho J, Chang T, Huh S, Chai J-Y (2019). Molecular identification of Anisakis larvae extracted by gastrointestinal endoscopy from health checkup patients in Korea. Korean J. Parasitol. 57: 207. https://doi.org/10.3347/ kjp.2019.57.2.207
- Unger P, Klimpel S, Lang T, Palm HW (2014). Metazoan parasites from herring (*Clupea harengus L.*) as biological indicators in the Baltic Sea. Acta Parasitologica 59(3): 518-528. https://doi.org/10.2478/s11686-014-0276-5
- Villazanakretzer DL, Napolitano PG, Cummings KF, Magann EF (2016). Fish parasites: a growing concern during pregnancy. Obstet. Gynecol. Surv. 71: 253-259. https://doi. org/10.1097/OGX.00000000000303
- World Health Organization (WHO)(2012). Soiltransmitted Helminths. World Health Organization. http://www.who.int/intestinal\_worms/en/.