



# Impact of Melatonin on Improving Productive Traits of Broiler Exposed to Environmental Stress

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**Abstract** | During a five-week trial, this study investigated the impact of various levels and methods of adding melatonin to water and diets on the growth performance of broiler chickens. One day old, 400 Ross 308 broiler chicks were randomly allocated into five random groups, with each group being quadruplicated and 20 birds allocated to each replication. Melatonin was supplemented in the G1 control group. G2 received 10 mg of melatonin per kg of diet, G3 received 20 mg of melatonin per kg of diet, G4 received 10 mg of melatonin per liter, and G5 received 20 mg of melatonin per liter, respectively. All groups' chicks were exposed to heat stress (30-35-30 °C) during the breeding period. Our results showed a significant increase ( $p \leq 0.01$ ) for the G5 group in live body weight (LBW) and cumulative weight gain (W.G.). The study compared the feed intake (F.I.) and relative growth rate (RGR) of this group with those of other groups. It found a significant improvement in feed intake (F.I.) for all melatonin additions. When comparing total mortality to the G1 group, there was a significant improvement for the G3, G4, and G5 groups, with rates of 1.449, 1.467, and 1.425 respectively, in feed conversion ratio (FCR) compared to the G1 group, which had a rate of 1.563.

**Keywords** | Broiler, Environmental, Heat stress, Melatonin, Productive performance.

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## INTRODUCTION

It is well-documented that heat stress can impact various tissues of a bird's body. It can compromise the integrity of the intestines by reducing oxygen levels in the intestinal epithelium (Dokladny *et al.*, 2016). This can lead to disruptions in the digestion and absorption of nutrients, as well as an imbalance in the microbial population within the intestinal tubule (Burkholder *et al.*, 2008; Deng *et al.*, 2012). This, in turn, leads to a decrease in the birds' productivity, as well as an imbalance in their physiological state, as the

hormone corticosterone increases during stress, impacting the depletion of energy sources and lowering the birds' immunity (Al-Jebory *et al.*, 2023; AL-Jaryan *et al.*, 2023). Melatonin, a hormone secreted from the pineal gland, is known for its antioxidant properties, which help capture free radicals both inside and outside the body (Bonfont-Rousselot *et al.*, 2011). In addition to its antioxidant properties and ability to scavenge free radicals, melatonin protects cells both inside the body (*in vivo*) and outside the body, directly and indirectly. Melatonin's antioxidant capacity is attributed to its amphiphilic nature, allowing it

to traverse physical barriers and mitigate oxidative damage in both fat cells and cells with high water content (Jou, 2010). This is accomplished through various mechanisms, including direct effects as an antioxidant and the neutralization of the toxic effects of active oxygen species, as well as indirect effects through the activation of antioxidant enzymes, which efficiently reduce the activity of enzymatic oxidants (Russel *et al.*, 2016), additionally, melatonin plays a significant part in removing the harmful effects of certain heavy metal elements that are involved in Haber-Weiss and Fenton reactions (Taha *et al.*, 2020; Al-Samrai *et al.*, 2023). Sahin *et al.* (2003) noted supplemental melatonin significantly increased live weight gain and carcass characteristics under stress conditions heat exposure increased the excretion of N, Ca, P, Zn, Fe, and Cr and decreased retention rates for them, and dietary melatonin supplementation returned these values to normal, while Kadim and Alhamdani, (2023) observed that adding melatonin to the feed reduced the negative effects of heat stress on broilers when added at a concentration of 0.50 mg/kg feed. Consequently, the current study investigates the impact of varying amounts of the hormone melatonin on enhancing the productive performance of broilers under ambient heat stress. Melatonin can thereby mitigate the detrimental effects of heat stress.

## MATERIALS AND METHODS

Four hundred broiler chickens (Ross 308) were utilized in this experiment, which was carried out at the Al-Anwar Poultry Company farm in the Babylon/Al-Muradiyah Governorate between July 25, 2023, and August 29, 2023. Each group had four replicates, and each one had 20 chicks. Melatonin was added to the diet and drinking water according to the following groups: G1 control group, G2 added 10 mg melatonin/ kg diet, G3 added 20 mg melatonin/ kg diet, G4 added 10 mg melatonin/ litter drinking water, G5 added 20 mg melatonin/ litter drinking water, Diets (The composition and nutrient content) were designed to meet the nutrient requirements for the birds during stages of growth (starter, grower, and finisher) based on NRC, (1994). All groups of chicks were exposed to heat stress (Table -1- °C) during the breed period.

**Table 1:** Explain the temperature (°C) during the breed period (time/ week).

Weeks	Hour			
	6 morning	12 evening	6 evening	12 morning
1	34.12	35.61	34.21	34.26
2	31.22	35.87	35.36	33.12
3	32.56	34.36	35.42	34.57
4	32.33	35.91	35.33	33.21
5	31.46	35.19	35.16	33.69

## PERFORMANCE TRAITS

The LBW, WG, feed intake, FCR, and mortality were measured according to (Al-Fayadh and Naji, 1989). The RGR was measured according to (Regassa *et al.*, 2014).

## LIVE BODY WEIGHT AND WEIGHT GAIN

The body weight and weight gain were calculated according to:

$$LBW = \text{birds weight} / \text{birds count}$$

$$WG = \text{final weight} - \text{initial weight}$$

## RELATIVE GROWTH RATE

The relative growth rate was measured by:

$$RGR = ((W2 - W1) / (0.5 (W2 + W1))) * 100 \text{ as } W1 = \text{initial weight and } W2 = \text{final weight.}$$

## FEED INTAKE

Feed intake per week was indicated according to:

$$FI = \text{feed intake} / \text{birds count}$$

## FEED CONVERSION RATIO

The feed conversion ratio was indicated according to:

$$FCR = \text{weight gain} / \text{feed intake} * 100$$

## MORTALITY %

The mortality was indicated throughout the whole study and calculated as follows.

$$\text{Total mortality percent} = (\text{The number of mortality birds during the experiment period}) / (\text{the total number of birds}) * 100$$

The data were analyzed using a completely randomized design (CRD) to study the effect of the studied parameters on the various characteristics, and the significant differences between the means were compared using the multinomial test (Duncan, 1955). The program SAS (2012) was used in statistical analysis according to the following mathematical model:

$$Y_{ij} = \mu + T_i + e_{ij}$$

So:  $Y_{ij}$ : the value of the  $j$ th view belonging to transaction  $i$ .

$\mu$ : the general average of the trait.

$T_i$ : The effect of treatment  $i$  (as the study included the effect of the 5 aforementioned treatments).

$e_{ij}$ : Random error that is normally distributed with a mean of zero and a variance of  $2 e \sigma$ .

## RESULTS AND DISCUSSION

### LIVE BODY WEIGHT

The effect of study groups on LBW is presented in Table 2. The results show no significant difference in initial weight; in the first week, the G3 groups demonstrated a significant

**Table 2:** Effect of melatonin in LBW of broiler exposed to heat stress (mean ± standard error)

Groups	Initial weight	1 <sup>st</sup> W	2 <sup>nd</sup> W	3 <sup>rd</sup> W	4 <sup>th</sup> W	5 <sup>th</sup> W
G1	37.61±0.23	159.97± 2.75 b	450.36± 4.07 b	846.16± 1.32 c	1398.44± 15.90 b	2007.44± 3.54 c
G2	37.82±0.39	168.33± 1.02 ab	457.34± 1.34 b	873.34± 1.26 b	1429.97± 15.11 ab	2075.43± 1.89 bc
G3	38.38±1.21	169.18± 2.24 a	470.02± 0.60 a	914.44± 2.93 a	1466.71± 12.67 a	2156.24± 6.44 ab
G4	37.94±0.14	167.34± 3.35 ab	472.54± 3.49 a	931.30± 2.73 a	1447.86± 6.07 a	2169.94± 2.84 ab
G5	38.52±0.72	167.50± 1.24 ab	479.29± 3.01 a	892.54± 4.23 b	1459.79± 11.17 a	2207.14± 3.27 a
Significant	N.S	*	**	**	**	**

N.S. not significant, \* at level (0.05≤p), \*\* at level (0.01≤p).

**Table 3:** Effect of melatonin in weekly W.G. of broiler exposed to heat stress (mean ± standard error)

Groups	1 <sup>st</sup> W	2 <sup>nd</sup> W	3 <sup>rd</sup> W	4 <sup>th</sup> W	5 <sup>th</sup> W	Cumulative W.G.
G1	122.36± 2.95	290.39± 3.23 b	395.79± 1.95 b	552.75± 4.51 ab	608.52± 2.96 c	1969.83± 3.75 c
G2	130.51± 0.99	289.01± 3.15 b	415.99± 8.34 b	556.63± 2.89 ab	645.45± 6.23 bc	2037.60± 1.66 bc
G3	130.80± 1.43	300.83± 2.40 ab	444.42± 1.25 a	552.26± 1.42 ab	689.53± 4.58 abc	2117.86± 4.98 ab
G4	129.39± 4.30	305.20± 4.75 ab	458.75± 1.57 a	516.55± 2.21 b	722.08± 5.93 ab	2131.99± 2.77 ab
G5	128.97± 2.03	311.79± 3.11 a	413.24± 2.69 b	567.45± 3.15 a	747.35± 3.39 a	2168.82± 3.45 a
Significant	N.S.	*	**	*	*	**

N.S. not significant, \* at level (0.05≤p), \*\* at level (0.01≤p).

**Table 4:** Effect of melatonin in RGR of broiler exposed to heat stress (mean ± standard error)

Groups	1 <sup>st</sup> W	2 <sup>nd</sup> W	3 <sup>rd</sup> W	4 <sup>th</sup> W	5 <sup>th</sup> W
G1	123.85±2.53	95.15±0.01 a	61.04±0.31 bc	46.57±1.25 a	35.73±0.33 b
G2	126.62±2.64	92.38±0.21 b	62.52±0.09 b	48.33±1.13 a	36.82±0.27 b
G3	126.05±2.35	94.12±0.30 ab	64.20±0.19 ab	28.90±1.07 b	38.06±0.04 ab
G4	126.06±1.79	95.39±0.11 a	65.37±0.72 a	43.42±1.23 a	39.91±0.61 ab
G5	125.20±1.85	96.41±0.31 a	60.24±0.49 c	48.24±1.15 a	40.76±1.00 a
Significant	N.S.	*	*	*	*

N.S. not significant, \* at level (0.05≤p).

**FEED INTAKE**

superiority (p ≤ 0.05) compared to the G1 group, while in the second week, there was a significant increase (p ≤ 0.01) in the G3, G4, and G5 groups compared to the G1 and G2 groups. By the third week, there was a significant increase (p ≤ 0.01) for the G2, G3, G4, and, G5 groups compared to the G1 group. The fourth week also showed a significant superiority (p ≤ 0.01) in the G3, G4, and G5 groups compared to the G1 group. Additionally, in the fifth week, there was a significant increase (p ≤ 0.01) in the G5 group compared to the G1 and G2 groups.

**WEEKLY WEIGHT GAIN**

Table 3 shows the effect of the experiment on weekly weight gain. The first week noted no significant difference between treatments. Second week appeared significant increase (p ≤ 0.05) for the G5 group compared to G1, and, G2 groups, in the third week had significant superiority (p ≤ 0.01) for the G3, and, G4 groups compared to the G1, G2, and, G5 groups, while in fourth the G5 groups had significant superiority (p ≤ 0.05) compared to G4 group

and not a significant difference between the G1, G2, G3, and, G4 groups, the G5 group appear height significant increase compared to G1, and, G2 groups in the fifth week and cumulative weight gain.

**RELATIVE GROWTH RATE**

The effect of different methods of adding melatonin to the RGR is noted in Table 4. There was no significant difference between groups in the first week the second week showed a significant increase (p ≤ 0.05) for the G1, G4, and G5 groups on the G2 group. In comparison, the G4 group had a higher significance (p ≤ 0.05) compared to the G1, G2, and G5 groups in the third week, in the fourth had were significant increase (p ≤ 0.05) for the G1, G2, G4, and, G5 groups on G3 group, as well in fifth week noted significant increase (p ≤ 0.05) for the G5 group compared to G1, and, G2 groups.

Table 5 shows the effect of the study on F.I. During the first week, there was a significant increase (p ≤ 0.01) for the G1, G2, G4, and G5 groups on the G3 group, second

**Table 5:** Effect of melatonin in F.I. of broiler exposed to heat stress (mean ± standard error)

Groups	1 <sup>st</sup> W	2 <sup>nd</sup> W	3 <sup>rd</sup> W	4 <sup>th</sup> W	5 <sup>th</sup> W	Total F.I.
G1	129.36± 1.89 a	312.71± 2.34 b	618.02± 8.56	964.61± 7.92 a	1400.87± 6.75 b	3425.58± 36.18
G2	130.01± 0.34 a	332.10± 1.12 a	611.06± 9.36	947.08± 12.21 ab	1468.34± 14.83 a	3488.61± 31.84
G3	122.66± 1.28 b	333.91± 2.48 a	593.27± 3.88	916.17± 13.12 b	1465.25± 12.98 a	3431.27± 17.85
G4	128.36± 0.90 a	339.15± 2.36 a	594.63± 7.21	964.91± 19.95 a	1483.90± 10.49 a	3510.96± 23.30
G5	127.52± 0.86 a	331.34± 1.00 a	597.49± 13.38	929.69± 10.42 ab	1487.38± 4.87 a	3473.43± 45.38
Significant	**	**	N.S.	*	*	N.S.

N.S. not significant, \* at level (0.05≤p), \*\* at level (0.01≤p).

**Table 6:** Effect of melatonin in FCR of broiler exposed to heat stress (mean ± standard error)

Groups	1 <sup>st</sup> W	2 <sup>nd</sup> W	3 <sup>rd</sup> W	4 <sup>th</sup> W	5 <sup>th</sup> W	Total FCR
G1	1.081± 0.05 a	1.112± 0.02 ab	1.561± 0.03 a	1.748± 0.05 ab	2.314± 0.12	1.563± 0.02 a
G2	0.996± 0.01 ab	1.149± 0.02 a	1.470± 0.03 ab	1.707± 0.06 ab	2.279± 0.07	1.524± 0.02 ab
G3	0.937± 0.01 b	1.110± 0.02 ab	1.386± 0.03 bc	1.664± 0.06 b	2.148± 0.13	1.449± 0.03 bc
G4	0.994± 0.03 ab	1.113± 0.02 ab	1.296± 0.02 c	1.868± 0.04 a	2.068± 0.10	1.467± 0.01 bc
G5	0.988± 0.01 b	1.062± 0.02 b	1.437± 0.03 b	1.639± 0.04 b	1.998± 0.08	1.425± 0.02 c
Significant	*	*	**	*	N.S.	**

N.S. not significant, \* at level (0.05≤p), \*\* at level (0.01≤p).

and fifth weeks appeared to significant increase (p ≤ 0.01) for the G2, G3, G4, and, G5 groups other G1 group, while in the third week and total F.I. there was no significant difference among groups, in fourth week shown significant increase (p ≤ 0.01) for the G1 and G4 groups on the G3 group.

**FEED CONVERSION RATIO**

The effect of the experiment in FCR is shown in (Table 6) during the first week significant improvement (p ≤ 0.05) for the G3 and G5 groups on the G1 group, and in the second week appeared to be a significant improvement (p ≤ 0.05) for the G5 group on the G2 group, while in the third week and total FCR significant improvement (p ≤ 0.01) for the G3, G4, and G5 groups on G1 group, in fourth-week significant improvement (p ≤ 0.05) for the G3, and G5 groups on G4 group, meanwhile no significant difference in the fifth week.

**Table 7:** Effect of melatonin in total mortality of broiler exposed to heat stress (mean ± standard error)

Groups	Total mortality
G1	18.75±6.25 a
G2	11.25±7.50 b
G3	12.50±6.25 b
G4	10.00±2.50 b
G5	12.50±1.25 b
Significant	*

\* at level (0.05≤p).

**TOTAL MORTALITY**

From 1-5 weeks of chick’s age (Table 7), the mortality was significantly higher (p ≤ 0.05) in the G1 group compared to the G2, G3, G4, and G5 groups.

Melatonin is important for circadian rhythms, and intestinal mucosal immune cells, this explains the productive performance improvement in melatonin groups. Melatonin can potentially support intestinal disease therapy significantly, but its exact mechanism remains unknown (Gil-Martín *et al.*, 2019), because melatonin in the colon typically does not enter the systemic circulatory system, melatonin is a multifunctional chemical with a special place in the bidirectional communication between the neuroendocrine and immunological systems (Paulose *et al.*, 2016). It seems intestinal melatonin has a localized influence on the mucosal immune system. Mucosal epithelial tissue, gut-associated lymphoid tissues, and commensal microbiota make up the intestinal mucosal immune system, which acts as the first line of defense against gastrointestinal infections and carries out the mucosal local defense to preserve immunological homeostasis (Nie *et al.*, 2019). Gut melatonin, which is impacted by feed intake, plays a significant role in the function of the gut epithelium (Ma *et al.*, 2017). The location of immune cells, microbial colonization, and nutrition digestion occur in the gastrointestinal intestine tract (Arnoldini *et al.*, 2018; Ma *et al.*, 2018). The intestinal epithelial barrier forms the mucosal immune system, shields surrounding tissues from intestinal contents and the luminal environment, and allows specific molecules to be absorbed for nutritional purposes; if this intestinal bal-

ance is upset, an overreaction to the interaction of gut microbes with the host barrier created by intestinal epithelial cells may result in the development of several gut-related disorders (Sommer *et al.*, 2017). This could explain why the melatonin addition groups' productive performance improved due to better intestinal health, epithelium histological features, and the microbial community, which enhanced nutrient absorption and digestion and consequently enhanced chick growth. Prior research on chickens has also demonstrated the connection between cellular oxidative stress and heat stress (Estévez, 2015; Surai *et al.*, 2019) since excess free radicals produced during oxidative stress damage DNA, lipids, proteins, and other components of every cell. Depending on how severe the oxidative stress is, the results might range from minor, reversible alterations to apoptosis and cell death (Lennon *et al.*, 1991). Therefore, in poultry, oxidative stress is linked to biological harm, severe health conditions, slower growth rates, and financial losses (Estévez, 2015). Melatonin functions by controlling the activity of mitochondria to lower the temperature of the cell and mitochondria, which lowers the generation of free radicals and lessens the impact of oxidative stress, studies have shown a decrease in the physiological and productive performance of birds raised at high temperatures, suggesting that the control group birds' worsening productivity may have been caused by heat stress (Ohtsu *et al.*, 2015), since the alterations are also caused by the chronic heat stress-related process of thermoregulation, behavioral and physiological harm to several tissues and organs, including the liver (Jastrebski *et al.*, 2017) and heart tissue (Aengwanich and Simaraks, 2004). The liver is crucial for preserving the body's metabolic equilibrium and plays a significant role in synthesizing antioxidants, contributing to the liver's low heat stress tolerance (Santana *et al.*, 2021). Apart from the fact that stress raises the hormone corticosterone, which in turn causes growth hormone to drop due to feedback, the hormone corticosterone also directs the body's energy resources to resist stress, which also results in lower growth (Jastrebski *et al.*, 2017).

Melatonin is usually associated with reduced oxidative damage (Taha, 2016). Melatonin's metabolites could eliminate reactive oxygen species and active nitrogen types, setting it apart from other antioxidants. This ongoing defense mechanism provided by melatonin and its metabolites is known as free radical defense (Hardeland and Pandi-Perumal, 2020), this means that even at low concentrations, melatonin is highly effective at protecting cells from oxidative damage (Tan *et al.*, 2015). One of the activities of melatonin is its antioxidant potential, which is attribute to two side chains: an indoleamine, a 3-amide group, and a 5-methoxy group (Galano *et al.*, 2016). Because of its high lipophilicity and hydrophobicity, melatonin and its metabolites can easily cross cell membranes, making them signif-

icant for direct and indirect antioxidant activities (Tamura *et al.*, 2012). Melatonin shields many biomolecules from damage caused by reactive oxygen species, nitrogen species, and free radicals through direct scavenging in oxidative stress situations; this reduces the quantity of glutathione in many cells (Pandi-Perumal *et al.*, 2013). The actions of enzymes that increase intracellular levels of decreased GSH are preserved by melatonin; glutathione is oxidized to its disulfide glutathione, which is swiftly reduced back to GSH by glutathione reductase; melatonin stimulates these enzymes, which is one of the critical ways that melatonin reduces oxidative stress, a nuclear binding site appears to be the site of action for melatonin's ability to modulate enzyme activities, which is one of the antioxidant activities that regulate the GSH/GSSG balance (Reiter *et al.*, 2018), as a result of increased melatonin, numerous GSH-metabolizing enzymes, such as CAT (Marshall *et al.*, 1996). Furthermore, melatonin increases gamma glutamylcysteine synthesizes to regulate glutathione formation, which considerably raises glutathione levels (Karasek and Winczyk, 2006), so that, in contrast to the traditional antioxidants (vitamins C and E), one melatonin molecule can scavenge up to ten ROS (Taha *et al.*, 2020; Al-Samrai *et al.*, 2023).

## CONCLUSION

Overall, heat stress decreased broiler production, while the concentration of 20 mg melatonin/liter of drinking water gave the best results in terms of improvement in LBW, WG., and relative growth efficiency, at the same time the other melatonin groups improved in productive performance compared to the control group.

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## CONFLICT OF INTEREST

The authors have declared no conflict of interest.

## NOVELTY STATEMENT

This study is the first one by using melatonin hormone in Iraq/ Babylon government, to decrease heat stress acute in broiler.

## AUTHORS CONTRIBUTION

All authors contributed equally for this work.

All animals in this study were handled and cared for according to the appropriate biosecurity procedures. The study was performed by the rules of the 'Guide for the Care and Use of Laboratory Animals, and Broiler ROSS 308 guide that were approved by the Ethics Committee of the College of Agriculture, Al-Qasim Green University, Iraq (Number 20 A.P. at 20/6/2023) before starting this study.

## REFERENCES

- Aengwanich W, Simaraks S (2004). Pathology of heart, lung, liver, and kidney in broilers under chronic heat stress. Songklanakarin J. Sci. Technol. 26: 417–424.
- Al-Fayad HAA, Naji SAH, Hajo NNA (2011). Poultry Products Technology. Press of the Ministry of Higher Education and Scientific Research - second edition. Baghdad, Iraq.
- AL-Jaryan IL, AL-Thuwaini TM, AL-Jebory HH (2023). Heat shock protein 70 and its role in alleviating heat stress and improving livestock performance. Rev. Agricult. Sci., 11: 234–242, 2023 [https://doi.org/10.7831/ras.11.0\\_234](https://doi.org/10.7831/ras.11.0_234).
- Al-Jebory HH, Al-Saeedi MKI, Al-Jaryan IL, Al-Khfaji FR (2023). Impact of neem (*Azadirachta indica*) leaves powder on growth performance of broiler (Ross 308) exposed to H.S. Res. J. Agricult. Biolog. Sci, 15(2): 1-5. <https://doi.org/10.22587/rjabs.2023.15.2.1>.
- Al-Samrai MK, Al-Jumaily TKH, Taha AT (2023). Impact light regimen and melatonin on growth performance, welfare and physiological parameters of broiler chickens. ICASE-2022 IOP Conf. Series: Earth Environ. Sci. 1225 (2023) 012046 IOP Publishing <https://doi.org/10.1088/1755-1315/1225/1/012046>.
- Arnoldini M, Cremer J, Hwa T (2018). Bacterial growth, flow, and mixing shape human gut microbiota density and composition. Gut Microbes. 9(6):1-8. <https://doi.org/10.1080/19490976.2018.1448741>
- Bonnefont-Rousselot D, Collin F, Jore D, Gardès-Albert M (2011). Reaction mechanism of melatonin oxidation by reactive oxygen species *in vitro*. J. Pineal Res., 50(3): 328–335. <https://doi.org/10.1111/j.1600-079X.2010.00847.x>
- Burkholder KM, Thompson KL, Einstein ME, Applegate TJ, Patterson JA (2008). Influence of stressors on normal intestinal microbiota, intestinal morphology, and susceptibility to *Salmonella enteritidis* colonization in broilers. Poult. Sci. 87: 1734–1741. <https://doi.org/10.3382/ps.2008-00107>.
- Deng X, Li Z, Zhang W (2012). Transcriptome sequencing of *Salmonella enterica serovar* Enteritidis under desiccation and starvation stress in peanut oil. Food Microbiol. 30: 311–315. <https://doi.org/10.1016/j.fm.2011.11.001>.
- Dokladny K, Zuhl MN, Moseley PL (2016). Intestinal epithelial barrier function and tight junction proteins with heat and exercise. J. Appl. Physiol. 120: 692–701. <https://doi.org/10.1152/jappphysiol.00536.2015>.
- Duncan DB (1955). Multiple Rang and Multiple F-test, Biometrics. 11. <https://doi.org/10.2307/3001478>
- Estévez M (2015). Oxidative damage to poultry: From farm to fork. Poult. Sci. 94: 1368–1378. <https://doi.org/10.3382/ps/pev094>
- Galano A, Castañeda-Arriaga R, Pérez-González A, Tan DX, Reiter RJ (2016). Phenolic melatonin-related compounds: Their role as chemical protectors against oxidative stress. Molecules. 21(11):1-42 <https://doi.org/10.3390/molecules21111442>
- Gil-Martín E, Egea J, Reiter RJ, Romero A (2019). The emergence of melatonin in oncology: focus on colorectal cancer. Med. Res. Rev. <https://doi.org/10.1002/med.21582> [published online ahead of print April 04, 2019].
- Hardeland R, Pandi-Perumal SR (2020). Melatonin, a potent agent in antioxidative defense: Actions as a natural food constituent, gastrointestinal factor, drug and prodrug. Nutrit. Metabol (London). (52;2):1-42 <https://doi.org/10.1186/1743-7075-2-22>
- Jastrebski SF, Lamont SJ, Schmidt CJ (2017). Chicken hepatic response to chronic heat stress using integrated transcriptome and metabolome analysis. PLoS One. 12: e0181900. <https://doi.org/10.1371/journal.pone.0181900>
- Jou MJ, Peng TI, Hsu LF, Jou SB, Reiter RJ, Yang CM, Chen CC (2010). Visualization of melatonin's multiple mitochondrial levels of protection against mitochondrial Ca<sup>2+</sup>-mediated permeability transition and beyond in rat brain astrocytes. J. Pineal Res., 48(1): 20-38. <https://doi.org/10.1111/j.1600-079X.2009.00721.x>
- Kadim EM, Alhamdani AA (2023). Study Reactive Effects of lighting systems and addition melatonin and L - tryptophane on heat shock proteins (HSP70), melatonin and the antioxidant status in broiler chicken managed in hot climates. IOP Conf. Series: Earth Environ. Sci. 1252 (2023) 012110. <https://doi.org/10.1088/1755-1315/1252/1/012110>.
- Karasek M, Winczyk K (2006). Melatonin in humans. J. Physiol. Pharmacol. 57(5):19-39.
- Lennon SV, Martin SJ, Cotter TG (1991). Dose-dependent induction of apoptosis in human tumour cell lines by widely diverging stimuli. Cell Prolif. 24: 203–214. <https://doi.org/10.1111/j.1365-2184.1991.tb01150.x>
- Ma N, Guo P, Zhang J (2018). Nutrients mediate intestinal bacteria-mucosal immune crosstalk. Front Immunol. 9:5. <https://doi.org/10.3389/fimmu.2018.00005>
- Ma N, Tian Y, Wu Y, Ma X (2017). Contributions of the interaction between dietary protein and gut microbiota to intestinal health. Curr. Protein Pept. Sc. 18(8):795-808. <https://doi.org/10.2174/1389203718666170216153505>
- Marsha KA, Reiter RJ, Poeggeler B, Aruoma OI, Halliwe B (1996). Evaluation of the antioxidant activity of melatonin *in vitro*. Free Rad. Biol. Med., 21(3): 307-315. [https://doi.org/10.1016/0891-5849\(96\)00046-9](https://doi.org/10.1016/0891-5849(96)00046-9)
- Nie P, Li Z, Wang Y (2019). Gut microbiome interventions in human health and diseases. Med. Res. Rev., <https://doi.org/10.1002/med.21584>.
- NRC (1994). Nutrient requirement of poultry gthEdn. National Academy Press. Washington. D.C.USA.
- Ohtsu H, Yamazaki M, Abe H, Murakami H, Toyomizu M (2015). Heat stress modulates cytokine gene expression in the spleen of broiler chickens. J. Poult. Sci. 52: 282–287. <https://doi.org/10.2141/jpsa.0150062>
- Pandi-Perumal SR, BaHammam AS, Brown GM, Spence DW, Bharti VK, Kaur C, Hardeland R, Cardinali DP (2013). Melatonin antioxidative defense: therapeutic implications for aging and neurodegenerative processes. Neurotox. Res., 23(3): 267-300. <https://doi.org/10.1007/s12640-012-9337-4>
- Paulose JK, Wright JM, Patel AG, Cassone VM (2016). Human

- gut bacteria are sensitive to melatonin and express endogenous circadian rhythmicity. *PLOS One*. 11(1):e0146643. <https://doi.org/10.1371/journal.pone.0146643>
- Regassa SL, Bekana E, Geleta T (2014). Production performance of fayoumi chicken breed under backyard management condition in Mid Rift Valley of Ethiopia. *Her J Agric Food Sci. Res.* 2(1):078–081.
- Reiter RJ, Tan DX, Rosales-Corral S, Galano A, Zhou XJ, Xu B (2018). Mitochondria: central organelles for melatonin's antioxidant and anti-aging actions. *Molecules.*, 23(2):.509 <https://doi.org/10.3390/molecules23020509>.
- Russel JR, Mayo JC, Tan D, Sainz RM, Alatorre-Jimenez M, Qin L (2016). Melatonin as an antioxidant: under promises but over delivers. *J. Pineal Res.* 61: 253–278. <https://doi.org/10.1111/jpi.12360>
- Sahin N, Onderci M, Sahin K (2003). Melatonin supplementation can ameliorate the detrimental effects of heat stress on performance and carcass traits of Japanese quail. *Biol. Trace Elem. Res.* 96: 169–177. <https://doi.org/10.1385/BTER:96:1-3:169>.
- Santana TP, Gasparino E, de Sousa FCB, Khatlab AS, Zancanela V, Brito CO, Barbosa LT, Fernandes RPM, Del Vesco AP (2021). Effects of free and dipeptide forms of methionine supplementation on oxidative metabolism of broilers under high temperature. *Animal; Int. J. Anim. Biosci.* 15: 100-173. <https://doi.org/10.1016/j.animal.2021.100173>.
- SAS (2012). Statistical Analysis System, User's Guide, Statistical. Version 9.1<sup>th</sup> ed. SAS. Inst. Inc. Cary. N.C. USA.
- Sommer F, Anderson JM, Bharti R, Raes J, Rosenstiel P (2017). The resilience of the intestinal microbiota influences health and disease. *Nat. Rev. Microbiol.* 15(10):630–638. <https://doi.org/10.1038/nrmicro.2017.58>
- Surai PF, Kochish II, Fisinin VI, Kidd MT (2019). Antioxidant Defence Systems and Oxidative Stress in Poultry Biology: An Update. *Antioxidants.* 20: 8, 235. <https://doi.org/10.3390/antiox8070235>
- Taha AT (2016). Effect adding different levels of Panax ginseng powdered on some reproductive traits and antioxidant status of Japanese quail males. *Euphrates J. Agricult. Sci.*, 8(3)86-92
- Taha AT, Al-Jumaily TKH, Al-Samrai MK (2020). Effect of melatonin in adult quail males exposed to oxidative stress induced by H<sub>2</sub>O<sub>2</sub>. 1<sup>st</sup> Scientific International Virtual Agricultural Conference Iop Conf. Series: Earth Environ. Sci. 553 (2020) 012013 IOP Publishing <https://doi.org/10.1088/1755-1315/553/1/012013>.
- Tamura H, Takasaki A, Taketani T, Tanabe M, Kizuka F, Lee L, Tamura I, Maekawa R, Aasada H, Yamagata Y, Sugino N (2012). The role of melatonin as an antioxidant in the follicle. *J. Ovarian Res.*,5:1-9 <https://doi.org/10.1186/1757-2215-5-5>
- Tan DX, Manchester LC, Esteban-Zubero E, Zhou Z, Reiter RJ (2015). Melatonin as a potent and inducible endogenous antioxidant: Synthesis Metabol. *Molecul.* 20(10): 18886-18906. <https://doi.org/10.3390/molecules201018886>